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# Detection of multidrug-resistant *Enterobacterales* isolated from buffalo species

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#### Abstract

The study was conducted to investigate the detection of multidrug-resistant *Enterobacterales* isolated in buffalo species during the period of August, 2019 to January, 2020 at the Department of Veterinary Pathology, LUVAS, Hisar. For bacteriological isolation, the samples were collected aseptically from the heart blood, liver, and mesenteric lymph nodes of buffalo species suspected of gastrointestinal tract disorders. The bacterial species were identified through the *Vitek-2* Compact system. Out of 31 Gram negative bacteria, a total 17 different bacterial species were identified. Most of the bacterial species showed multiple drug resistance patterns.

Keywords: Buffalo, multidrug, Enterobacterales, bacteriological, and vitek-2

#### Introduction

Multidrug-resistant Enterobacterales (MDRE) are a major cause of hospital-acquired infections associated with high morbidity and mortality, as well as rising the cost of healthcare (Vrancianu et al., 2021)<sup>[1]</sup>. In several species of Enterobacteriaceae, there is an increase in antimicrobial resistance (Karlowsky et al., 2003)<sup>[2]</sup>. The generic E. coli, which is commonly found in raw meat, has the capability to transfer antibiotic resistance to other bacteria residing in the intestine (Dunowska et al., 2006)<sup>[3]</sup>. Kim et al. (2005)<sup>[4]</sup> reported that enteric bacteria develop resistance to common antibiotics used in human and veterinary gentamicin, medicine. like tetracycline, kanamycin, and streptomycin. In Enterobacteriaceae, the E. coli serotype (O157:H7) and Salmonella caused most of the foodborne illness and were often transmitted through raw meats (Kennedy et al., 2005) [5]. Cogan et al. (1999)<sup>[6]</sup> reported that Salmonella could potentially spread on kitchen surfaces during the preparation of the contaminated poultry. The spread of antibiotic-resistant foodborne pathogens threatens the successful treatment of infectious diseases (Andersson, 2003)<sup>[7]</sup>.

#### **Materials and Methods**

The present investigation was undertaken on 38 buffalo carcasses (17 adult buffalo and 21 buffalo calves) suspected of gastrointestinal tract disorders that were brought to the Department of Veterinary Pathology, LUVAS, Hisar for post-mortem during the period of six months (from August, 2019 to January, 2020). The study included the detection of multi-drug resistant *Enteriobacterales* identified in buffalo carcasses. The isolated microorganisms were subjected to in-vitro antibiotic sensitivity testing using antibiotic discs with appropriate conc. (Table 1) by the single disc diffusion technique (Bauer *et al.*, 1966)<sup>[8]</sup>. The results were then interpreted on the basis of the inhibition zone as per the manufacturer's recommendations (Hi-Media, India). The bacterial isolates showing resistance to more than three antibiotic groups or combinations were placed in the MDR category (Oteo *et al.*, 2005)<sup>[9]</sup>.

#### Results

The results of *in vitro* antibiotic sensitivity are depicted in Table 2. The results revealed that *E. coli* showed maximum resistance to cephalosporin/ $\beta$ -lactamase inhibitors, monobactams,

(polymyxin-B), penicillin+\beta-lactamase polymyxins followed fluoroquinolone inhibitors, by and aminoglycosides (Table 2). Likewise, the Salmonella spp. was found to be most resistant to cephalosporin/B-lactamase inhibitors cefepime). monobactams. (except aminoglycosides (except streptomycin), and penicillin+βlactamase inhibitors. Serratia plymuthica was found to be 100% resistant to cephalosporin/β-lactamase inhibitors, fluoroquinolone, monobactams, and followed by penicillin+ $\beta$ -lactamase inhibitors, but no resistance to polymyxins and aminoglycosides. Proteus mirabilis, Ochrobactrum anthropi, and Gardnerella vaginalis were found to be resistant to most of the antibiotics except aminoglycosides. Polymyxins also showed no resistance against Proteus mirabilis. In the case of Sphingomonas paucimobilis, which was found to be most resistant to monobactams and the fluoroquinolone group of antibiotics, it showed no resistance to other groups of antibiotics. Likewise, Stenotrophomonas multophila, Pseudomonas mendocina, and Pseudomonas aeruginosa were showed no resistant to most of the antibiotics except monobactams, which showed 100% resistance against Stenotrophomonas multophila and were 50% resistant to polymyxins and fluoroquinolone. Pseudomonas mendocina was found to be 33.3% resistant to the monobactam group of antibiotics. The multidrug resistance (MDR) pattern of Escherichia coli and other Enterobacterales was depicted in Tables 3 and 4, which revealed that all isolates of E. coli, Ochrobactrum anthropi, Gardnerella vaginalis, Proteus mirabilis, and Serratia plymuthica showed MDR patterns, but other Enterobacterales did not show any multidrug resistance (MDR) patterns.

# Discussion

The current study was conducted for detection of multi-drug resistant Enteriobacterales identified in buffalo carcasses (38) including adult and calves. The results revealed that E. coli showed maximum resistance to cephalosporin/βinhibitors, monobactams, lactamase polymyxins (polymyxin-B), penicillin+ $\beta$ -lactamase inhibitors, followed by fluoroquinolone and aminoglycosides. In support of our study, Londhe (2010) <sup>[10]</sup> also reported the resistance of E. coli to cephalosporin but in contrast to our study, Londhe (2010)<sup>[10]</sup> and Lehreena (2008)<sup>[11]</sup> reported the sensitivity of E. coli to polymyxins and cephalosporin/ $\beta$ -lactamase inhibitors, respectively. More or less similar findings were reported by Dinesh et al. (2018)<sup>[12]</sup> regarding the resistance of *E. coli* to penicillin+ $\beta$ -lactamase inhibitors and aminoglycosides, but in contrast to our study, they found that E. coli showed sensitivity towards fluoroquinolone. Brennan et al. (2016)<sup>[13]</sup>, in their study in bovine, found that one isolate of *E. coli* was resistant to the  $\beta$ -lactamase and fluoroquinolone groups of antibiotics. The results showed that, Salmonella spp. was found to be most resistant to cephalosporin/ $\beta$ -lactamase inhibitors (except cefepime), monobactams, aminoglycosides (except streptomycin), and penicillin+\beta-lactamase inhibitors. More or less similar results regarding the susceptibility of Salmonella spp. were reported by Sushma et al. (2016)<sup>[14]</sup>. The study of sensitivity pattern in Serratia plymuthica showed that, it was found to be 100% resistant to cephalosporin/β-lactamase inhibitors, monobactams, and fluoroquinolone, followed by penicillin+ $\beta$ -lactamase inhibitors, but no resistance to polymyxins and aminoglycosides. Likewise, Proteus

mirabilis, Ochrobactrum anthropi, and Gardnerella vaginalis were found to be resistant to most of the antibiotics except aminoglycosides. Polymyxins also showed no resistance against *Proteus mirabilis*. Similar findings were reported by Londhe (2010)<sup>[10]</sup> regarding the susceptibility of Proteus spp. to antibiotics. In the case of Sphingomonas paucimobilis, which was found to be most resistant to monobactams and the fluoroquinolone group of antibiotics, it showed no resistance to other groups of antibiotics. But in contrast to our study, Cengiz et al. (2015) <sup>[15]</sup> reported the greatest resistance of Sphingomonas paucimobilis to gentamicin. In the current study, Stenotrophomonas multophila, Pseudomonas mendocina, and *Pseudomonas aeruginosa* were showed no resistant to most of the antibiotics except monobactams, which showed 100% resistance against Stenotrophomonas multophila and were 50% resistant to polymyxins and fluoroquinolone. Pseudomonas mendocina was found to be 33.3% resistant to the monobactam group of antibiotics. The study of multidrug resistance (MDR) pattern of Escherichia coli and other Enterobacterales revealed that all isolates of E. coli, Ochrobactrum anthropi, Gardnerella vaginalis, Proteus mirabilis, and Serratia plymuthica showed MDR patterns, but other Enterobacterales did not show any multidrug resistance (MDR) patterns. More or less similar results were reported by Shrestha et al. (2017) [16] regarding the MDR pattern of total isolated bacterial species.

### Conclusion

In the present study, 61.29% of the of the bacterial isolates exhibit the MDR pattern and are mostly resistant to cephalosporin+ $\beta$ -lactamase inhibitors, monobactams, penicillin+ $\beta$ -lactamase inhibitors, fluoroquinolone, and aminoglycosides. At present, regulation on the misuse of antibiotics should be necessary.

# References

- 1. Vrancianu CO, Dobre EG, Gheorghe I, Barbu I, Cristian RE, Chifiriuc MC. Present and future perspectives on therapeutic options for carbapenemaseproducing *Enterobacterales* infections. Microorganisms. 2021;9(4):730.
- Karlowsky JA, Jones ME, Thornsberry C, Friedland IR, Sahm DF. Trends in antimicrobial susceptibilities among *Enterobacteriaceae* isolated from hospitalized patients in the United States from 1998 to 2001. Antimicrobial agents and chemotherapy. 2003;47(5):1672-1680.
- 3. Dunowska M, Morley PS, Traub-Dargatz JL, Hyatt DR, Dargatz DA. Impact of hospitalization and antimicrobial drug administration on antimicrobial susceptibility patterns of commensal *Escherichia coli* isolated from the feces of horses. Journal of the American Veterinary Medical Association. 2006;228(12):1909-1917.
- 4. Kim SH, Wei CI, Tzou YM, An H. Multidrug-resistant *Klebsiella pneumoniae* isolated from farm environments and retail products in Oklahoma. Journal of food protection. 2005;68(10):2022-2029.
- Kennedy J, Blair IS, McDowell DA, Bolton DJ. The microbiological status of none/food contact surfaces in domestic kitchens and the growth of *Staphylococcus aureus* in domestic refrigerators. Food Prot. Trends. 2005;25:974-980.

- 6. Cogan TA, Bloomfield SF, Humphrey TJ. The effectiveness of hygiene procedures for prevention of cross-contamination from chicken carcases in the domestic kitchen. Letters in applied microbiology. 1999;29(5):354-358.
- 7. Andersson DI. Persistence of antibiotic resistant bacteria. Current opinion in microbiology. 2003;6(5):452-456.
- 8. Bauer AW. Antibiotic susceptibility testing by a standardized single disc method. Am. J. of Clinc. Path. 1996;45:149-158.
- Oteo J, Lázaro E, de Abajo FJ, Baquero F, Campos J, members of EARSS S. Antimicrobial-resistant invasive *Escherichia coli*, Spain. Emerging Infectious Diseases. 2005; 11(4):546.
- 10. Londhe MS Studies on pathological investigation of gastrointestinal tract of bovines. M.V.Sc. Thesis, Chaudhary Charan Singh Haryana Agricultural University, Hisar, India; c2010.
- 11. Lehreena. Etio-pathological studies on calf mortality with special reference to hepatic disorders in bovine calves. M.V.Sc. Thesis, CCS HAU, Hisar; c2008.
- 12. Mittal D, Grakh K, Prakash A, Moudgil P, Devi B, Jadhav V. Isolation, Identification and characterization of enteric bacteria from post weaning diarrheic pigs and their resistance to multiple antibiotics. International Journal of Current and Applied Microbiology. 2018;7:2377-2384.
- 13. Brennan E, Martins M, McCusker MP, Wang J, Alves BM, Hurley D, *et al.* Multidrug-resistant *Escherichia coli* in bovine animals, Europe. Emerging infectious diseases. 2016;22(9):1650.
- 14. Sushma S, Vikas Nehra VN, Deepika Lather DL. Aetiopathological studies of digestive and respiratory affections in buffalo calves. Haryana Vet. 2016;55(2):170-175.
- 15. Cengiz S, Seyitoglu S, Altun SK, Dinler U. Detection of *Sphingomonas paucimobilis* infections in domestic animals by VITEK® Compaq 2 and Polymerase Chain Reaction. Archivos de medicina veterinaria. 2015;47(1):117-119.
- 16. Shrestha A, Bajracharya AM, Subedi H, Turha RS, Kafle S, Sharma S, *et al.* Multi-drug resistance and extended spectrum beta lactamase producing Gram negative bacteria from chicken meat in Bharatpur Metropolitan, Nepal. BMC research notes. 2017;10:1-5.