



ISSN Print: 2617-4693
 ISSN Online: 2617-4707
 IJABR 2024; SP-8(3): 431-437
www.biochemjournal.com
 Received: 01-12-2023
 Accepted: 05-01-2024

Nitesh Kumar Singh
 Department of Soil Science and
 Agricultural Chemistry,
 Institute of Agricultural
 Sciences, Banaras Hindu
 University, Varanasi,
 Uttar Pradesh, India

Anand Prakash Singh
 Department of Soil Science and
 Agricultural Chemistry,
 Institute of Agricultural
 Sciences, Banaras Hindu
 University, Varanasi,
 Uttar Pradesh, India

Corresponding Author:
Nitesh Kumar Singh
 Department of Soil Science and
 Agricultural Chemistry,
 Institute of Agricultural
 Sciences, Banaras Hindu
 University, Varanasi,
 Uttar Pradesh, India

Evaluation of different insoluble zinc sources along with zinc solubilizing bacteria on growth of wheat (*Triticum aestivum*) in an inceptisol having marginal Zn availability

Nitesh Kumar Singh and Anand Prakash Singh

DOI: <https://doi.org/10.33545/26174693.2024.v8.i3Sf.777>

Abstract

The present study intended to evaluate the effect of insoluble Zn sources when applied with and without zinc solubilizing bacteria, on the growth and zinc content of the wheat crop. For this purpose, a pot experiment was designed and conducted during 2016-17 and 2017-18 using factorial completely randomized design (FCRD). Three zinc compounds ($ZnSO_4$ (Zn_1), ZnO (Zn_2), and $ZnCO_3$ (Zn_3)) with two distinct doses (1 mg kg^{-1} (D_1) and 2 mg kg^{-1} (D_2) Zn) were used in the experiment. In response to ZSB application, the data revealed an increase in wheat growth and, as a result, an increase in wheat yields also. Significant differences among the effects of sources and bacterial inoculations were noticed during the experiment. Performance of all the Zn sources including $ZnSO_4$ was found to improve when applied with ZSB. In contrast to the control, all isolates resulted in a nearly 1.5- to 2-fold increase in Zn content in grains and straw. In case of zinc solubilizing bacteria, inoculation of ZSB₂ showed best results as compared to other two sources. It was concluded that the application of $ZnSO_4$ @ 2 mg kg^{-1} Zn along with ZSB₂ was had the best zinc management strategy to conflict the zinc deficiency.

Keywords: Growth, yield, zinc solubilizing bacteria, Zn sources, pot experiment, Zn content

1. Introduction

Despite the massive increase in wheat production due to Green Revolution, the present average yield is nowhere near the potential yield (Liu *et al.*, 2017) [23]. One of the limiting factors being the cultivation of wheat in soils with severely low or marginal levels of Zn (Alloway, 2008) [2]. Over the years, various methods for increasing zinc availability in soil to plants have been used (Sharma *et al.*, 2015) [34]. Though it is a long shot, several approaches have been suggested to address the aforementioned problem, one of which is the use of zinc containing chemical fertilizers such as $ZnSO_4$, ZnO , $ZnCO_3$, $Zn_3(PO_4)_2$ and $ZnCl_2$ (Joy *et al.* 2017; Kumar *et al.*, 2017) [17, 21], use of organic farming (Bhattacharya *et al.*, 2006) [5], using transgenic approach (Bashir *et al.*, 2010) [4], and the use of microbial inoculants (PGPR) (Tariq *et al.*, 2007) [37]. The major problem with the use of chemical fertilizers are its quick fixation in soil. About 96% of the exogenously applied Zn fertilizers are converted into various unavailable forms depending upon the soil types and physicochemical reactions within a week of application (Gontia-Mishra *et al.*, (2017) [11]. Organic farming and transgenic approaches to addressing zinc deficiency have struggled due to their high production costs and implementation methods. Recently, some researchers have begun to look into the possibility of using microbial inoculants to convert the inaccessible form of zinc into the usable form. Use of these microbial inoculants in sustainable agriculture and soil regeneration is generating a lot of buzz. These microbial inoculants are referred as zinc solubilizers (Hussain *et al.*, 2015; Mumtaz *et al.*, 2017) [14, 27]. Among the bacterial genera *Bacillus*, *Acinetobacter*, *Pseudomonas*, and *Thiobacillus ferrooxidans*, *Enterobacter*, *Arthrobacter*, *Burkholderia*, *Exiguobacterium*, *Gluconacetobacter*, *Mycobacterium*, *Ralstonia*, *Stenotrophomonas*, *Sphingomonas* and *Xanthomonas* have been reported as zinc solubilizers and shown to promote plant growth (Saravanan *et al.* 2007; Gandhi and Muralidharan, 2016) [32, 10].

Based on these facts a pot experiment was planned to be conducted in the rabi seasons of 2016 and 2017 with an objective to evaluate the effect of different insoluble zinc sources along with zinc solubilizing bacteria on growth parameters of wheat crop grown in an Inceptisol of Varanasi, having Zn availability just above critical value.

2. Materials and Methods

2.1 Bacterial isolates

Forty bacterial isolates were taken for the experiment from the Soil Microbiology Laboratory, Department of Soil Science and Agricultural Chemistry, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi. Based on the morphological characters and assayed *in vitro* for Zn solubilization on Bunt and Rovira media (Bunt and Rovira, 1955) [40] supplemented with ZnO (ZnO). Out of all, three bacterial isolates were selected as Zn solubilizers and tested in the pot for *in situ* Zn solubilizing activity. In addition to the bacterial inoculation, changes in the growth parameters as well as zinc status of the plants were also observed.

2.2 Plant Material and Treatment

Wheat variety (HUW 234) was used for the experiment which was obtained from the Department of Genetics and Plant Breeding, Banaras Hindu University, Varanasi, Uttar

Pradesh, India. The selected wheat variety (HUW 234) is commonly grown in entire wheat area of the North Eastern Plains Zone of India. The experiment was steered in net house of the Department of Soil Science and Agricultural Chemistry, I. Ag. Sc., Banaras Hindu University, Varanasi in two succeeding *rabi* seasons (2016–17 and 2017–18). The experiment was laid out in FCRD with three factors *viz.*, sources of Zn ($ZnSO_4-Zn_1$, $ZnO-Zn_2$, and $ZnCO_3-Zn_3$); two doses 1 mg kg^{-1} - D_1 and 2 mg kg^{-1} - D_2 Zn) and three bacterial isolates (ZSB_1 , ZSB_2 and ZSB_3) treatments were replicated thrice. The recommended dose of fertilizers N, P, K (120:60:60) were mixed with soil in suitable quantities. Seeds were treated with selected bacterial isolates with the help of sticker solution (10% of Jaggery solution). Bacteria treated seeds of wheat were sown in each pot. After full emergence of first leaves, four healthy plants were maintained per pot.

2.3 Experimental soil

The experimental soil was collected from Agricultural Research Farm, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi. The initial soil properties are present in Table 1. After processing, the soil was filled in earthen pots of 10 Kg capacity which were lined with polythene sheet for checking the loss of water and nutrients from the pots.

Table 1: Physico-chemical properties of the initial soil

Particulars	Value	
	2016-17	2017-18
A. Mechanical separates		
Texture		
a) Sand (%)	63.20	60.15
b) Silt (%)	25.10	21.34
c) Clay (%)	15.44	16.20
Textural class	Sandy Clay Loam	
B. Physico-chemical characteristics		
pH	7.8	8.1
Electrical conductivity (dSm^{-1})	0.2	0.18
Bulk density	1.39	1.35
CEC	19.6	20.2
WHC (%)	42.35	40.22
Organic carbon (%)	0.40	0.38
Available N (mg/kg)	81.22	83.14
Available P (mg/kg)	8.24	7.56
Available K (mg/kg)	80.2	84.1
DTPA extractable-Zn (mg/kg)	0.68	0.71
C. Biological properties		
Soil microbial biomass carbon ($mg\ C\ kg^{-1}\ soil$)	92.6	90.1
Dehydrogenase activity ($\mu g\ TPF\ g^{-1}\ soil\ day^{-1}$)	25.64	22.2
D. Soil microbial count		
Bacteria ($cfu \times 10^5\ g^{-1}\ soil$)	15.4	13.8
ZSB ($cfu \times 10^4\ g^{-1}\ soil$)	7.8	7.2

2.4 Pot experiment

Wheat variety (HUW 234) was used for the experiment which was obtained from Agriculture Farm, Banaras Hindu University, Varanasi, Uttar Pradesh, India. The selected wheat variety (HUW 234) is commonly grown in entire wheat area of the North Eastern Plains Zone of India. The experiment was conducted in net house of the Department of Soil Science and Agricultural Chemistry, I. Ag. Sc., Banaras Hindu University, Varanasi for two consecutive *rabi* seasons (2016–17 and 2017–18). The experiment was

laid out in FCRD with three factors *viz.*, sources of Zn ($ZnSO_4-Zn_1$, $ZnO-Zn_2$, and $ZnCO_3-Zn_3$); two doses 1 mg kg^{-1} - D_1 and 2 mg kg^{-1} - D_2 Zn) and three bacterial isolates (ZSB_1 , ZSB_2 and ZSB_3) treatments were replicated thrice. The recommended dose of fertilizers N, P, K (120:60:60) were mixed with soil in suitable quantities. Wheat seed were treated with selected bacterial isolates with the help of sticker solution (10% of Jaggery solution). Bacteria treated seeds of wheat were sown in each pot. After full emergence of first leaves, four healthy plants were maintained per pot.

2.5 Analysis of growth attributes of wheat

Wheat growth parameters including plant height, number of leaves and chlorophyll content were recorded at 30, 60 and 90 DAS. Plant height was recorded with the help of meter scale and expressed in centimeter while the chlorophyll content of the leaves was measured by SPAD meter. The spike length of the plant was measured from base to top most tip of the spikelet and it was expressed in cm after computation.

2.6 Statistical analyses

The raw data generated during the whole experiment were subject to statistical analysis by following the Factorial Complete Randomized Design (FCRD) to draw the valid differences among the treatments.

3. Results

3.1 Chlorophyll content (SPAD)

Chlorophyll content (SPAD value) recorded during two years (2016-17 and 2017-18) of the pot experiment at three growth stages (30, 60, and 90 DAS) of wheat crop are presented in Table 2. The content increased up to 60 DAS and thereafter decreased. At 30 DAS, highest chlorophyll (36.24) was observed with application of Zn through ZnSO₄ during 2016-17. Between the two doses of Zn, higher SPAD

value, (33.99) was recorded with application of zinc @ 2 mg kg⁻¹ (Zn₂) while among the zinc solubilizing bacterial (ZSB) strains, highest solubilization was observed in case of ZSB₂ (33.30). In 2017-18, SPAD value (35.65) of plants receiving ZnSO₄ was higher than SPAD value of plants receiving other Zn sources. Application of Zn₂ resulted in 5.05% higher SPAD value than Zn₁. Highest chlorophyll (42.82), at 60 DAS was observed with application of Zn through the ZnCO₃ during 2016-17. Between the zinc doses, Zn₂ exhibited highest SPAD value (40.82) as compared to Zn₁ while in case of zinc solubilizing bacterial strains; ZSB₂ recorded the highest chlorophyll (41.17). In 2017-18, chlorophyll content (42.53) in plants receiving ZnSO₄ superior to other sources. Zn₂ caused higher SPAD value over Zn₁ which produced 4% lesser SPAD value than Zn₂. ZSB₂ resulted in higher chlorophyll (40.80) over ZSB₁ (40.01). At 90 DAS, highest chlorophyll (33.36) was observed with the application of Zn through ZnCO₃ during 2016-17. Among the dose and zinc solubilizing bacterial strains, Zn₂ and ZSB₂ recorded the highest SPAD value, i.e. 30.86 and 31.12, respectively. In 2017-18, the SPAD value (34.16) of plants receiving ZnSO₄ was superior to SPAD value of plants receiving other sources. Application of Zn₂ resulted in 10% higher content than Zn₁.

Table 2: Effect of zinc with zinc solubilizing bacteria on chlorophyll content of wheat

Treatment	Chlorophyll (SPAD)					
	2016-17			2017-18		
	30 DAS	60 DAS	90 DAS	30 DAS	60 DAS	90 DAS
Zn ₀	31.90	37.76	26.32	32.22	37.48	27.09
Zn ₁	36.24	39.84	28.21	35.65	42.53	34.16
Zn ₂	32.03	39.63	30.21	32.31	39.18	28.80
Zn ₃	33.86	42.82	33.36	33.89	39.48	30.80
Sem±	0.17	0.18	0.29	0.16	0.16	0.31
CD (0.05)	0.48	0.50	0.81	0.47	0.46	0.88
D ₁	33.02	39.21	28.18	32.69	38.92	28.83
D ₂	33.99	40.82	30.86	34.34	40.42	31.60
Sem±	0.12	0.13	0.20	0.12	0.12	0.22
CD (0.05)	0.34	0.36	0.57	0.33	0.33	0.62
B ₀	31.72	38.45	27.58	31.90	38.09	28.36
B ₁	33.74	40.26	30.18	33.63	40.01	31.00
B ₂	35.27	41.17	31.12	34.67	40.80	31.70
B ₃	33.30	40.17	29.21	33.87	39.77	29.81
Sem±	0.17	0.18	0.29	0.16	0.16	0.31
CD (0.05)	0.48	0.50	0.81	0.47	0.46	0.88

Where Zn₀ = no zinc, Zn₁=ZnSO₄, Zn₂= ZnO, Zn₃= ZnCO₃, D₁ and D₂= Doses of Zn, B₀= no bacteria, B₁, B₂, B₃= Treatment with bacterial strain 1,2 and 3 respectively

3.2 Plant height

The data pertaining to plant height of wheat during the two-year pot experiment (2016-17 and 2017-18) at three growth stages (30, 60, and 90 DAS) are presented in Table 3. The highest plant height (19.15 cm) at 30 DAS was observed with the application of Zn through ZnSO₄ during 2016-17. Application of Zn @ 2 mg kg⁻¹ (Zn₂) resulted in higher plant height (18.67 cm) over-application of Zn @ 1 mg kg⁻¹ (Zn₁). Among the zinc solubilizing bacterial strains (ZSB), the highest increment was observed in the case of ZSB₂ (18.86 cm). In 2017-18, plants receiving ZnSO₄ showed the highest plant height (21.41 cm) over plant receiving other sources. The application of Zn₂ (dose) exhibited 11.41% higher plant

height than Zn₁. The highest plant height (45.43 cm) was observed at 60 DAS when Zn was applied through ZnSO₄ in 2016-17. Comparing the two doses of Zn (Zn₁ and Zn₂) pointed out that Zn₂ resulted in 5% higher effect on plant height. Among the zinc solubilizing bacteria, highest plant height (44.43 cm) was noted in ZSB₁. In 2017-18, the highest plant height (45.54 cm) was noted when Zn was applied through ZnSO₄. At 90 DAS, ZnSO₄ @ 2 mg kg⁻¹ treated plants showed the highest plant height during both the years (2016-17 and 2017-18). Among the ZSB treated plants, highest plant height (48.57cm) was observed with ZSB₁. In 2017-18, ZSB₂ showed the highest plant height (48.76 cm).

Table 3: Effect of zinc with zinc solubilizing bacteria on plant height of wheat

Treatment	Plant height (cm)					
	2016-17			2017-18		
	30 DAS	60 DAS	90 DAS	30 DAS	60 DAS	90 DAS
Zn ₀	18.18	38.24	43.77	17.44	38.83	43.57
Zn ₁	19.15	45.43	50.87	21.41	45.54	50.52
Zn ₂	18.38	41.24	48.27	20.57	41.05	47.32
Zn ₃	18.61	42.47	48.50	21.35	40.76	46.79
SEm±	0.11	0.15	0.32	0.27	0.17	0.18
CD (0.05)	0.32	0.43	0.90	0.77	0.48	0.52
D ₁	18.49	40.87	46.74	19.10	40.59	45.87
D ₂	18.67	42.82	48.97	21.28	42.49	48.23
SEm±	0.08	0.11	0.22	0.19	0.12	0.13
CD (0.05)	0.22	0.30	0.64	0.55	0.34	0.37
B ₀	18.27	39.07	46.63	18.83	38.97	44.85
B ₁	18.58	44.43	48.57	20.78	42.44	47.55
B ₂	18.86	42.52	48.33	21.13	43.41	48.76
B ₃	18.60	41.36	47.89	20.03	41.36	47.04
SEm±	0.11	0.15	0.32	0.27	0.17	0.18
CD (0.05)	0.32	0.43	0.90	0.77	0.48	0.52

Where Zn₀ = no zinc, Zn₁=ZnSO₄, Zn₂= ZnO, Zn₃= ZnCO₃, D₁ and D₂= Doses of Zn, B₀= no bacteria, B₁, B₂, B₃= Treatment with bacterial strain 1,2 and 3 respectively

3.3 Number of leaves

The data related to the effect of different treatments on the number of leaves of wheat crop at different growth stages during 2016-17 and 2017-18 are presented in Table 4. At 30 DAS, application of Zn through ZnO and ZnCO₃ was at par in its effect on the no. of leaves in first year. Whereas, significantly higher number of leaves (3.81) was recorded with Zn₂ (2 mg kg⁻¹). In case of zinc solubilizing bacterial strains, ZSB₂ followed by ZSB₃ caused highest number of leaves (3.84 and 3.78, respectively) over uninoculated control (3.47). In second year of pot experiment, application of Zn through ZnO showed highest leaves (3.67). Zn₂ significantly increase the no. of leaves in comparison to Zn₁. Inoculation with bacterial strains ZSB₂, ZSB₁ and ZSB₃ significantly increased the number of leaves as compared to un-inoculated control. Application of ZSB₂ showed 10% higher leaves than ZSB₀. Results revealed that application Zn through ZnSO₄ showed highest number of leaves (10.09) followed by ZnO (9.80) at 60 DAS in 2016-17. Higher number of leaves was recorded with Zn₂ which was ~5% higher in its effect on number of leaves than Zn₁. Among the ZSB, Highest increase in plant leaves was exhibited by ZSB₂ (10.50) followed by ZSB₁ (9.64). During the second year of pot experiment, plants receiving ZnSO₄ showed highest number of leaves (10.01). Significantly higher (9.54) leaves were recorded with application of Zn₂. The ZSB₂ caused highest no. of leaves (10.48) and showed its significant superiority over the other bacterial strains. ZSB₂ caused an increment of 30% in number of leaves over ZSB₀. Among the zinc sources, the highest number of leaves (9.98) at 90 DAS was recorded with ZnSO₄ during 2016-17. Application of ZnO and ZnCO₃ @ Zn₂ exhibited 9.96 and 9.53 leaves, respectively. Between the two doses (Zn₁ and Zn₂) of Zn, Zn₂ produced significantly higher number of leaves (9.74) which was ~4% higher than the effect caused by Zn₁. Among the ZSB, ZSB₂ produced the highest number of leaves (10.66) followed by ZSB₁ (9.58) inoculated plants. In the second year (2017-18) of pot experiment, the plants receiving ZnO produced highest number of leaves (10.47). While in the case of doses, the application of Zn₂ resulted in significantly higher number of leaves (9.94) over application of Zn₁ (9.92). Among the ZSB, plants inoculated

with ZSB₂ produced the highest number of leaves (11.10) which was 26.27% higher than the effect caused by uninoculated control.

Table 4: Effect of zinc with zinc solubilizing bacteria on no. of leaves of wheat

Treatment	No. of Leaves					
	2016-17			2017-18		
	30 DAS	60 DAS	90 DAS	30 DAS	60 DAS	90 DAS
Zn ₀	3.55	8.47	8.78	3.48	8.14	8.60
Zn ₁	3.66	10.09	9.98	3.63	10.01	10.02
Zn ₂	3.83	9.80	9.96	3.67	9.85	10.47
Zn ₃	3.82	9.49	9.53	3.59	9.38	10.65
SEm±	0.04	0.11	0.10	0.08	0.10	0.06
CD (0.05)	0.13	0.30	0.27	0.22	0.30	0.17
D ₁	3.62	9.24	9.39	3.61	9.15	9.94
D ₂	3.81	9.68	9.74	3.57	9.54	9.92
SEm±	0.03	0.08	0.07	0.06	0.07	0.04
CD (0.05)	0.09	0.21	0.19	0.16	0.21	0.12
B ₀	3.47	8.31	8.52	3.44	8.07	8.79
B ₁	3.77	9.64	9.58	3.69	9.55	10.10
B ₂	3.84	10.50	10.66	3.80	10.48	11.10
B ₃	3.78	9.40	9.49	3.43	9.28	9.74
SEm±	0.04	0.11	0.10	0.08	0.10	0.06
CD (0.05)	0.13	0.30	0.27	0.22	0.30	0.17

Where Zn₀ = no zinc, Zn₁=ZnSO₄, Zn₂= ZnO, Zn₃= ZnCO₃, D₁ and D₂= Doses of Zn, B₀= no bacteria, B₁, B₂, B₃= Treatment with bacterial strain 1,2 and 3 respectively

3.4 Spike length

The data about the spike length (cm) of wheat during 2016-17 and 2017-18 are presented in table 5. During the 1st year of the pot experiment, the highest spike length (8.10 cm) was observed with the application of Zn through ZnCO₃ followed by ZnO (7.98 cm) and ZnSO₄ (7.80 cm). Similarly, the highest increase in spike length was observed due to the application of Zn @ 2mg kg⁻¹ (Zn₂). In the case of plants inoculated with ZSB, ZSB₃ inoculated plants showed the highest spike length (8.08 cm). In the second year of the pot experiment, the application of Zn through ZnSO₄ and ZnO caused the significant increase in spike length (8.26 cm and 7.81 cm, respectively). Zn₂ produced the significantly higher spike length (8.04 cm) which was ~5% higher than the

effect caused by Zn₁. Among the zinc solubilizing bacterial strains, highest spike length (8.04 cm) was observed in plants inoculated with ZSB₂ which was 19.16% higher than the effect caused by uninoculated control.

Table 5: Effect of zinc with zinc solubilizing bacteria on spike length of wheat

Treatment	Spike length (cm)	
	2016-17	2017-18
Zn ₀	7.73	7.59
Zn ₁	7.80	8.26
Zn ₂	7.98	7.81
Zn ₃	8.10	7.80
Sem	0.14	0.10
CD	0.41	0.28
D1	7.83	7.69
D2	7.97	8.04
Sem	0.10	0.07
CD	0.29	0.20
B0	7.77	7.20
B1	7.85	7.92
B2	7.90	8.58
B3	8.08	7.75
Sem	0.14	0.10
CD	0.41	0.28

Where Zn₀ = no zinc, Zn₁=ZnSO₄, Zn₂= ZnO, Zn₃= ZnCO₃, D₁ and D₂= Doses of Zn, B₀= no bacteria, B₁, B₂, B₃= Treatment with bacterial strain 1,2 and 3 respectively

4. Discussion

The increment in SPAD value may be due to the action of zinc solubilizing bacteria on different insoluble zinc sources resulting in enhanced supply and uptake of zinc to the plant. Zinc plays a very important role in the enhancement of chlorophyll content in the plant because it acts as a structural and catalytic component of proteins and enzymes responsible for pigment biosynthesis (Samreen *et al.*, 2013) [30]. Similarly, Othman *et al.* (2017) [28] reported that inoculation with zinc-solubilizing bacterium (TM56) at 0.2 mg/L ZnSO₄ produced the highest chlorophyll content (12.34 mg/cm²) followed by plants inoculated with (TM9) with a chlorophyll content of 10.97 mg/cm² at 0.2 mg/L ZnSO₄. Similar findings were also reported by Sarathambal *et al.* (2009) [31] and Majumdar and Chakraborty (2015) [24]. Various sources and doses produced similar variation in plant height recorded at 30, 60 and 90 DAS during both years of pot experiment. In case of zinc solubilizing bacteria, the trend was not similar at different stages. Application of ZnSO₄ @ Zn₂ and ZSB₂ significantly increased the plant height in wheat at 30 DAS in first year and 30, 60 and 90 DAS in second year. In first year at 60 and 90 DAS highest height was observed in plants treated with ZnSO₄ @ Zn₂ and ZSB₁. This finding shows that application of zinc with zinc solubilizing bacteria causes significant effect on plant height. Similarly, Goteti *et al.* (2013) [12] reported that seed bacterization with zinc solubilizing bacteria resulted in increased plant height. Ramesh *et al.* (2014) [29] reported that inoculation with zinc solubilizing bacteria *Bacillus aryabhatai* significantly increases the plant height. The increase in plant height may be attributed to the adequate supply of zinc which could meet the enhanced zinc demand by the action of zinc solubilizing bacteria on different zinc sources. Zinc solubilizing bacteria increase the plant growth by making the nutrient available through N₂ fixation, phosphate

solubilization, Zn solubilization, siderophores and phytohormones production and antagonistic effect on plant pathogens (Iqbal *et al.*, 2010; Wang *et al.*, 2014 and Gandhi and Muralidharan 2016) [16, 39, 10]. Kamran *et al.* (2017) also observed that inoculation of zinc solubilizing bacteria *P. agglomerans* in jute plant significantly increased the plant height as compared to uninoculated control. Othman *et al.* (2017) [28] reported that Inoculation of plant with Zn solubilizing bacterium *Acinetobacter* sp. (TM56) at 0.2 mg/L ZnSO₄ produced the significantly higher plant height. Gandhi and Muralidharan (2016) [10] reported that application of zinc solubilizing bacterial strains (AGM₃ + AGM₉) with insoluble source of zinc significantly enhanced the growth and yield of rice crop over control under pot experiment. This might be due to the production of different types of growth hormones by ZSB.

Zinc solubilizing bacterial strains ZSB₂ and ZSB₁ significantly increased the number of leaves in wheat under both the years of pot experiment at 30, 60 and 90 DAS. Among the sources, plants receiving Zn through ZnO recorded highest number of leaves at 30 DAS which was at par with ZnCO₃ treated plants and both were higher than the control in both the years. At 60 and 90 DAS, ZnSO₄ treated plants recorded highest number of leaves. Results showed that zinc solubilizing bacterial isolates with zinc sources significantly increased the number of leaves in wheat. This increment in number of leaves may be attributed to the solubilization of nutrients by zinc solubilizing bacteria. Improvement in different plant parameters by seed bacterization with zinc solubilizing bacteria has been reported by Goteti *et al.* (2013) [12].

The spike length of wheat was significantly higher in the plants receiving zinc through ZnCO₃ and ZnSO₄ during 2016-17 and 2017-18, respectively. However, the similar trend was observed in case of doses where Zn₂ produced the highest spike length in both the years of pot experiment. Plants inoculated with ZSB₃ the highest spike length in 1st year of pot experiment. But in 2nd year of pot experiment, plants inoculated with ZSB₂ recorded highest spike length. Results of our study showed that inoculation of ZSB along with different zinc sources enhanced spike length of wheat under net house condition. The increase in spike length may be associated with the more availability of other nutrients in the rhizosphere due to zinc (Tarique *et al.*, 2016) [37]. Kenbaev and Sack (2002) [18] also reported that increase in zinc availability to the plant increases the number of spikes m⁻². Dewal and Pareek (2004) [8] also reported that zinc application improved spike length. Similar findings were reported by Asad and Rafique (2000) [3] and Curtin *et al.* (2008) [6].

5. Conclusion

The result demonstrated that the growth attributes *viz.* chlorophyll content (SPAD), plant height, number of leaves and spike length of wheat crop were enhanced by the application of zinc with zinc solubilizing bacteria over uninoculated control. Hence, on the basis of above findings it was concluded that the application of Zn @ 2mg kg⁻¹ through ZnSO₄ along with bacterial isolate ZSB₂ exhibited the best Zn management strategy for improving growth of wheat in inceptisol having marginal Zn availability.

6. Acknowledgements

The authors are thankful to the Department of Soil Science and Agricultural Chemistry, I. Ag. Sc., Banaras Hindu

University (BHU) for providing necessary facilities to conduct this experiment. Authors are also thankful to University Grants Commission (UGC), New Delhi for providing financial support during research work in the form of Fellowship.

7. References

- Alloway BJ. Zinc in soils and crop nutrition. Brussels, Belgium: International Zinc Association; c2004. p. 130.
- Alloway BJ. Zinc in soils and crop nutrition. Brussels, Belgium and Paris, France: IZA and IFA; c2008. p. 139.
- Asad A, Rafique R. Effect of zinc, copper, iron, manganese and boron on the yield and yield components of wheat crop in Tehsil Peshawar. Pakistan Journal of Biological Sciences. 2000;3(10):1615-1620.
- Bashir K, Ishimaru Y, Nishizawa NK. Iron uptake and loading into rice grains. Rice. 2010;3(2-3):122-130.
- Bhattacharyya P, Chakraborty A, Chakrabarti K, Tripathy S, Powell MA. Copper and zinc uptake by rice and accumulation in soil amended with municipal solid waste compost. Environmental Geology. 2006;49(7):1064-1070.
- Curtin D, Martin RJ, Scott CL. Wheat (*Triticum aestivum*) response to micronutrients (Mn, Cu, Zn, B) in Canterbury, New Zealand. New Zealand Journal of Crop and Horticultural Science. 2008;36(3):169-181.
- Desai S, Kumar P, Sultana U, Pinisetty S, Reddy G. Potential microbial candidate strains for management of nutrient requirements of crops. African Journal of Microbiology Research. 2012;6(17):3924-3931.
- Dewal GS, Pareek RG. Effect of phosphorus, sulphur and zinc on growth, yield and nutrient uptake of wheat (*Triticum aestivum*). Indian Journal of Agronomy. 2004;49(3):160-162.
- Epstein E, Bloom AJ. Mineral nutrition of plants: principles and perspectives; c2005.
- Gandhi A, Muralidharan G. Assessment of zinc solubilizing potentiality of *Acinetobacter* sp. isolated from rice rhizosphere. European Journal of Soil Biology. 2016;76:1-8.
- Gontia-Mishra I, Sapre S, Tiwari S. Zinc solubilizing bacteria from the rhizosphere of rice as prospective modulator of zinc biofortification in rice. Rhizosphere. 2017;3:185-190.
- Goteti PK, Emmanuel LDA, Desai S, Shaik MHA. Prospective zinc solubilising bacteria for enhanced nutrient uptake and growth promotion in maize (*Zea mays* L.). International journal of microbiology; c2013.
- Hafeez FY, Abaid-Ullah M, Hassan MN. Plant growth-promoting rhizobacteria as zinc mobilizers: a promising approach for cereals biofortification. In: Maheshwari DK, Ed. Bacteria in Agrobiolgy: Crop Productivity. Berlin, Heidelberg: Springer; c2013. p. 217-235.
- Hussain A, Arshad M, Zahir ZA, Asghar M. Prospects of zinc solubilizing bacteria for enhancing growth of maize. Pakistan Journal of Agricultural Sciences. 2015;52(4).
- Hussain A, Zahir ZA, Asghar HN, Imran M, Ahmad M, Hussain S. Integrating the potential of *Bacillus* sp. Az6 and organic waste for zinc oxide bio-activation to improve growth, yield and zinc content of maize grains. Pakistan Journal of Agricultural Sciences. 2020;57(1).
- Iqbal U, Jamil N, Ali I, Hasnain S. Effect of zinc-phosphate-solubilizing bacterial isolates on growth of *Vigna radiata*. Annals of microbiology. 2010;60(2):243-248.
- Joy EJ, Ahmad W, Zia MH, Kumssa DB, Young SD, Ander EL, Broadley MR. Valuing increased zinc (Zn) fertiliser-use in Pakistan. Plant and Soil. 2017;411(1-2):139-150.
- Kenbaev B, Sade B. Response of field-grown barley cultivars grown on zinc-deficient soil to zinc application. Communications in Soil Science and Plant Analysis. 2002;33(3-4):533-544.
- Khande R, Sharma SK, Ramesh A, Sharma MP. Zinc solubilizing *Bacillus* strains that modulate growth, yield and zinc biofortification of soybean and wheat. Rhizosphere. 2017;4:126-138.
- Khangahi MY, Ricciuti P, Allegretta I, Terzano R, Crecchio C. Solubilization of insoluble zinc compounds by zinc solubilizing bacteria (ZSB) and optimization of their growth conditions. Environmental Science and Pollution Research. 2018;25(26):25862-25868.
- Kumar AS, Meenakumari KS, Anith KN. Screening for Zn solubilisation potential of soil bacteria from Zn deficient soils of Kerala. Journal of Tropical Agriculture. 2017;54(2):194.
- Kumar A, Dewangan S, Lawate P, Bahadur I, Prajapati S. Zinc-Solubilizing Bacteria: A Boon for Sustainable Agriculture. In: Kumar A, Singh D, Singh P, eds. Plant Growth Promoting Rhizobacteria for Sustainable Stress Management. Singapore: Springer; c2019. p. 139-155.
- Liu DY, Zhang W, Pang LL, Zhang YQ, Wang XZ, Liu YM, Zou CQ. Effects of zinc application rate and zinc distribution relative to root distribution on grain yield and grain Zn concentration in wheat. Plant and Soil. 2017;411(1-2):167-178.
- Majumdar S, Chakraborty U. Phosphate solubilizing rhizospheric *Pantoea agglomerans* Acti-3 promotes growth in jute plants. World J Agric. Sci. 2015;11:401-410.
- Martino E, Perotto S, Parsons R, Gadd GM. Solubilization of insoluble inorganic zinc compounds by ericoid mycorrhizal fungi derived from heavy metal polluted sites. Soil Biology and Biochemistry. 2003;35(1):133-141.
- Ministry of Agriculture and Farmers Welfare, Government of India. Advance Estimate/4th Adv Estimates 201718n Eng.pdf [Internet]. 2018 [cited 2018 Dec 25]. Available from: <https://eands.dacnet.nic.in/Advance Estimate/4th Adv Estimates 201718n Eng.pdf>
- Mumtaz MZ, Ahmad M, Jamil M, Hussain T. Zinc solubilizing *Bacillus* spp. potential candidates for biofortification in maize. Microbiological Research. 2017;202:51-60.
- Othman NMI, Othman R, Saud HM, Wahab PEM. Effects of root colonization by zinc-solubilizing bacteria on rice plant (*Oryza sativa* MR219) growth. Agriculture and Natural Resources. 2017;51(6):532-537.
- Ramesh A, Sharma SK, Sharma MP, Yadav N, Joshi OP. Inoculation of zinc solubilizing *Bacillus aryabhatai* strains for improved growth, mobilization and biofortification of zinc in soybean and wheat

- cultivated in Vertisols of central India. Applied Soil Ecology. 2014;73:87-96.
30. Samreen T, Shah HU, Ullah S, Javid M. Zinc effect on growth rate, chlorophyll, protein and mineral contents of hydroponically grown mungbeans plant (*Vigna radiata*). Arabian Journal of Chemistry. 2013;10:S1802-S1807.
 31. Sarathambal C, Thangaraju M, Gomathy M. Solubilization of insoluble zinc compounds by *Gluconacetobacter diazotrophicus* and its influence on maize. Asian Journal of Bio Science. 2009;4(1):110-112.
 32. Saravanan VS, Madhaiyan M, Thangaraju M. Solubilization of zinc compounds by the diazotrophic, plant growth promoting bacterium *Gluconacetobacter diazotrophicus*. Chemosphere. 2007;66(9):1794-1798.
 33. Sharma SK, Ramesh A, Joshi OP. Characterization of zinc-solubilizing *Bacillus* isolates and their potential to influence zinc assimilation in soybean seeds. J. Microbiol. Biotechnol. 2011;22:352-359.
 34. Sharma A, Patni B, Shankhdhar D, Shankhdhar SC. Evaluation of different PGPR strains for yield enhancement and higher Zn content in different genotypes of rice (*Oryza sativa* L.). Journal of Plant Nutrition. 2015;38(3):456-472.
 35. Shewry PR. Wheat. Journal of Experimental Botany. 2009;60(6):1537-1553.
 36. Sunithakumari K, Devi SP, Vasandha S. Zinc solubilizing bacterial isolates from the agricultural fields of Coimbatore, Tamil Nadu, India. Current Science. 2016:196-205.
 37. Tariq M, Hameed S, Malik KA, Hafeez FY. Plant root associated bacteria for zinc mobilization in rice. Pakistan Journal of Botany. 2007;39(1):245.
 38. Tavallali V, Rahemi M, Eshghi S, Kholdebarin B, Ramezani A. Zinc alleviates salt stress and increases antioxidant enzyme activity in the leaves of pistachio (*Pistacia vera* L. 'Badami') seedlings. Turkish Journal of Agriculture and Forestry. 2010;34(4):349-359.
 39. Wang Y, Yang X, Zhang X, Dong L, Zhang J, Wei Y, Lu L. Improved plant growth and Zn accumulation in grains of rice (*Oryza sativa* L.) by inoculation of endophytic microbes isolated from a Zn Hyperaccumulator, *Sedum alfredii* H. Journal of Agricultural and Food Chemistry. 2014;62(8):1783-1791.
 40. Bunt JS, Rovira AD. Microbiological studies of some subantarctic soils. Journal of Soil Science. 1955 Jan;6(1):119-128.