

ISSN Print: 2617-4693 ISSN Online: 2617-4707 IJABR 2024; 8(2): 566-570 www.biochemjournal.com Received: 21-12-2023 Accepted: 27-01-2024

Author's details are given below the reference section

Screening of high-yielding Swarna lines developed through MABB for resistance against bacterial blight and blast diseases

M Kousik, E Punniakotti, G Rekha, K Chaitra, T Dilip Kumar, SK Hajira, K Swapnil, SK Mastanbee, M Anila, M Ayyappadass, B Laxmi Prasanna, Pragya Sinha, Ravindra Kale, K Jyothi, G Vivek, D Aleena, P Senguttuvel, GS Laha, MS Prasad, P Sudhakar, A Krishna Satya, CN Neeraja, R Abdul Fiyaz and RM Sundaram

DOI: https://doi.org/10.33545/26174693.2024.v8.i2g.630

Abstract

Rice cultivation has two significant challenges: BB and blast diseases. There are a number of Indian rice variety that are extremely sensitive to these diseases, one of which is the well-known Swarna mega-rice variety. The resistance genes Xa21 and Pi54, which provide protection against bacterial blight (BB) and blast diseases, were incorporated into a Near Isogenic Line (NIL) of Swarna that already had the yield-boosting gene OsSPL14 using marker-assisted backcross breeding. Ten best introgressed lines were selected at the ICF₆ stage and subjected to phenotypic screening in a standardized blast nursery to evaluate resistance to blast disease and under controlled conditions for bacterial leaf blight. The lines exhibited outstanding performance and showed good tolerance to biotic stresses such as bacterial blight and blast.

Keywords: Bacterial blight, blast, yield, MABB

Introduction

Rice is among the three most extensively cultivated crops globally, alongside wheat and maize. India's rice demand is projected to reach 130 million tonnes by 2030 and 160 million tonnes by 2050, based on an estimated daily consumption of 189 g per person and an average annual population growth rate of 1.8%. To address this need, it is essential to enhance rice productivity and production (Guru *et al.* 2018) ^[6]. Abiotic and biotic stresses are the primary causes of significant yield reductions in several crops. One of the most harmful types of biotic stress that rice is subjected to is bacterial leaf blight (BB), which is caused by *Xanthomonas oryzae* pv. *Oryzae* (*Xoo*). According to Kumar *et al.* (2012) ^[13], the severity of BB can result in a reduction of rice yield of up to the extent of 85 percent. Researchers have found a total of 48 resistance genes from various rice sources to date (Jiaxin Xing *et al.*, 2021; Fiyaz *et al.*, 2022; Chen *et al.*, 2020) ^[10, 5, 4]. Sundaram *et al.* (2008) ^[28] and Hue ThiNguyen *et al.* (2018) ^[9] have demonstrated that the *Xa21* gene, which is both dominant and resistant, derives from the wild species *Oryza longistaminata* that is native to Africa. This gene has been found to offer resistance that is both widespread and extensive.

The fungus *Magnaporthe oryzae* produces rice blast disease, which can lead to significant yield losses of 70-80% in severe cases, posing a substantial threat to rice production. Sahu PK *et al.* 2022 ^[22] reported the existence of around 146 important rice blast resistance genes (R-genes). The *Pi54* gene, derived from the Tetep cultivar, has demonstrated strong effectiveness against Indian blast isolates (Sharma *et al.*, 2002) ^[23]. In 1982, Maruteru, Andhra Pradesh, India, introduced Swarna, also known as MTU7029, a long-duration, medium-bold grain type mega-rice variety. Swarna experiences significant reductions in crop output because of its high vulnerability to bacterial blight and its low resistance to blast disease. With the use of the marker-assisted backcross breeding (MABB) technique, an effort was made to incorporate *Xa21* and *Pi54* genes into the high-yielding Swarna variety that already contains the yield improving gene *OsSPL14*.

Corresponding Author: Dr. R Abdul Fiyaz Senior Scientist, ICAR-Indian Institute of Rice Research, Rajendra Nagar, Hyderabad, Telangana, India This action was taken in an attempt to enhance the yield of the line.

Materials and Methods Plant materials

A NIL of Swarna (IR121047-2-2-1) susceptible to bacterial blight and blast but possessing *OsSPL14* for panicle branching was crossed with a NIL of ISM (ISM-12-305-106) containing resistance genes *Xa21* and *Pi54* against BB and blast, respectively (Rekha *et al.* 2018)^[21]. The crossing programme began in the Kharif season of 2015 and concluded in the Kharif season of 2021. During the ICF₆ generation, 10 best lines were selected for assessing their resistance to BB and blast disease, along with resistant and susceptible checks.

Assessing the intercross developed lines of Swarna for resistance to bacterial blight.

The enhanced intercross derived Swarna lines were planted in the field and assessed for their resistance to BB, in comparison with their parents in the Rabi 2020-21 and Kharif 2021. The negative parent (Swarna) and positive parent (ISM) checks for bacterial leaf blight were also included in the evaluation. A highly infectious strain, IX0-20, sourced from Telangana, of Xanthomonas oryzae pv. Oryzae was used for the screening process. The bacterial culture was cultivated on Hayward's agar media and placed in an incubator at 28 °C for 96 hours. Following the completion of the incubation time, the bacterial culture was collected and diluted to a final concentration of 108 colonyforming units per milliliter (Preece et al., 1982)^[18]. In 1973, Kauffman et al. developed a leaf clipping approach that involves cutting leaf tips with Xoo-infected scissors to expose crosscut veins to Xoo suspension. The IRRI SES score was used to assess the average percentage of diseased leaf area on the top three leaves of the plants 15 days after inoculation.

Assessing the intercross developed lines of Swarna for blast resistance

The Swarna intercross derived lines, together with their parent lines NIL of Swarna (IR121047-2-2-1) and NIL of ISM (ISM-12-305-106), were assessed for blast resistance in UBN beds during the Rabi 2020-21 and Kharif 2021 seasons, compared to the negative check (HR12) and positive check (Tetep). Pathogen cultures were prepared by breaking down 10-day-old mycelia slants in 5 milliliter of water and then placing them on Mathur's medium for sporulation. After incubating for 8 to 10 days at a temperature range of 25 to 28 degrees Celsius, the plates were rinsed with 10 milliliters of water to create a spore suspension. The concentration of the pathogen Magnaporthe oryzae spore suspension was adjusted to 1×105 spores/ml. 15-day-old seedlings were inoculated with around 40 cc of spore suspension (Local IIRR isolate-SPI 40) of the blast infectious agent using a low-volume plastic sprayer. Sprinklers were employed to moisten the environment with water 3-4 times a day to maintain elevated humidity levels. Disease resistance and susceptibility scores were recorded on a 0-9 scale when standard blast lesions were observed on each line.

Results and Discussion

The yield of rice is considerably reduced by various biotic stresses such as bacterial blight (BB) and blast infections (Tanweer *et al.*, 2015)^[28]. Because of the country's rapidly

evolving environment, these two diseases have been identified in many regions, and widely grown varieties such as Swarna are especially susceptible to them. Resistance genes can be incorporated into popular cultivars like Swarna to provide improved varieties for a range of races/isolates of BB and blast diseases.

Swarna (MTU7029) is a mega rice variety developed and released by ANGRAU in 1982. It is known for its adaptability, brown hull color, dark green foliage, high tillering ability, semi-dwarf plant stature, long duration, and medium bold grains. Indian farmers utilize Swarna for farming in kharif and Rabi seasons. It is derived from the cross Vasistha and Mahsuri, as reported by Revathi *et al.* (2020) ^[15] and Anila *et al.* (2018) ^[1]. In 2019, there was a notable request for 397.25 quintals of breeder seed.

Phenotypic assessment for resistance to bacterial leaf blight

A robust resistance to the disease was demonstrated by each of the ten intercross-derived lines, with lesion lengths ranging from one centimeter to six centimeters and SES scores which ranged from one to three (Fig 1; Table 1). With a disease score of nine and a lesion length that was greater than twelve centimeters, the recurrent parent Swarna demonstrated that it was also susceptible to bacterial blight disease. It was determined that the donor parent NIL of ISM (ISM-12-305-106) had an immune level of resistance. The disease score was one, and the lesion length ranged from zero to less than one centimeter. The diseases known as bacterial blight and blast are extremely damaging and have the potential to reduce plant output by as much as fifty percent (Sharma et al., 2017). Xa21 gene, which encodes a leucine-rich repeat receptor-like kinase (LRR-RLK) gene, was the first R gene to be cloned. This gene was originally derived from Oryza Longistaminata, according to Song et al. 1995^[25].

In spite of the fact that it is typically preferable to combine many genes into high-quality rice varieties, there have been examples in which a prominent BB resistance gene, such as Xa21, has demonstrated the ability to exhibit strong resistance to the disease. It is well known that Xa21 is significantly resistant to a number of different strains of the bacterial blight pathogen that is prevalent in India. To provide resistance to the bacterial blight disease in the Swarna variety, we utilised the Xa21 gene in our research work.



Fig 1: Phenotypic screening of improved Swarna lines at ICF₆ generation for resistance to BB disease

Fig 1. Phenotypic screening of the selected ICF₆ lines exhibited resistance to BB with a score of '1'. P1- Swarna (susceptible check), P2- ISM-12-305-106 (NIL of Improved Samba Mahsuri (Xa21+Pi54); resistance check), P3-

https://www.biochemjournal.com

IR121047-2-2-1 (OsSPL14), IL-1 to IL-5- Intercross derived

lines of Swarna.

| Table 1: Screening Improved breeding lines of Swarna (at ICF ₆) along with the parents, with DX020 isolate of <i>Xanthomonas oryzae</i> pv. |
|--|
| oryzae to assess their bacterial blight resistance |

| | | Reaction against bacterial leaf blight disease | | |
|--------|---|--|-------|--|
| S. No. | Parents and Checks | Lesion length (cm) | Score | Immune/ Resistant/Moderately resistant/Susceptible/ Moderately susceptible/Highly Susceptible |
| 1. | Swarna | 12 | 9 | Susceptible |
| 2. | ISM-12-305-106 | Less than 1 | 1 | Resistant |
| 3. | IR121047-2-2-1 | Less than 12 | 9 | Susceptible |
| S. No. | Improved breeding lines (ICF ₆) | Lesion length (cm) | Score | Resistant/Moderately resistant/Susceptible |
| 1. | Swarna-IL-1 | Less than 3 | 1 | Resistant |
| 2. | Swarna-IL-2 | Less than 2 | 1 | Resistant |
| 3. | Swarna-IL-3 | Less than 1 | 1 | Resistant |
| 4. | Swarna-IL-4 | Less than 1 | 1 | Resistant |
| 5. | Swarna-IL-5 | Less than 2 | 1 | Resistant |
| 6. | Swarna-IL-6 | Less than 2 | 1 | Resistant |
| 7. | Swarna-IL-7 | Less than 2 | 1 | Resistant |
| 8. | Swarna-IL-8 | Less than 1 | 1 | Resistant |
| 9. | Swarna-IL-9 | Less than 1 | 1 | Resistant |
| 10. | Swarna-IL-10 | Less than 1 | 1 | Resistant |

Phenotypic screening for blast resistance

There was a high level of resistance to blast disease across all ten intercross-derived lines, with scores ranging from 2-3. Both the negative check, HR12,TN1, and the recurrent parent, NIL of Swarna (IR121047-2-2-1), displayed a high susceptibility to blast disease, as indicated by a score of 9. The positive check, Tetep, and NIL of ISM (ISM-12-305-106) on the other hand, revealed a high level of resistance to blast disease, with scores of 1 and 3, respectively (Figure 2; Table 2). According to Khush and Jena (2009)^[12], rice blast is a particularly severe disease that is brought on by the fungus Magnaporthe grisea. This disease can cause yield losses of between 70 and 80 percent. As demonstrated by Sharma et al. (2010) ^[24] and Ramkumar et al. (2011) ^[20], Pi54 is a major gene that confers resistance to a large number of blast pathogen isolates in India. The Pi54 gene, which codes for an NBS-LRR protein, was successfully isolated and duplicated from the indica cv. Tetep strain for the first time. According to Rai et al. (2011) [19], it offers resistance against Indian rice blast isolates that is extremely broad. The efficiency of the gene in giving a high level of resistance was demonstrated in a previous study that was conducted by Balachiranjeevi et al (2015)^[3]. According to the findings of Prasad et al. (2011) [17], Samba Mahsuri breeding lines that included the gene showed a high level of resistance to blast diseases in trials conducted in many locations. When compared to the original parent, Swarna, the improved cultivars of Swarna had much better levels of resistance to bacterial leaf blight and blast.



Fig 2: Phenotypic screening of improved Swarna lines at ICF₆ generation for resistance to blast disease

Figure 2. Phenotypic screening of five selected intercross derived lines IL-1 to IL-5 showed a resistance against blast disease with the score of 2-3 as per IRRI-SES Scale, P1-ISM-12-305-106 (NIL of Improved Samba Mahsuri (*Xa21+Pi54*); resistance check) P2- Swarna (susceptible check), P3-Tetep (Resistance check), P4- IR121047-2-2-1 (*OsSPL14*), P5- TN1, P6- HR-12 (Susceptible check), IL-1 to IL-5- Intercross derived lines of Swarna.

| Table 2: Screening Improved breeding lines of Swarna (at ICF ₆) along with the parents, with SPI-40 isolate of <i>Magnaporthe oryzae</i> to assess |
|---|
| their blast resistance a (scoring done as per IRRI-SES scale; IRRI 2013) |

| | Parents and Checks | Reaction against blast disease SPI-40 | | |
|--------|---|--|--|--|
| S. No. | | | | |
| | | Score | Resistant/Moderately Resistant/Susceptible | |
| 1. | Swarna | 9 | Susceptible | |
| 2. | ISM-12-305-106 | 3 | Resistant | |
| 3. | Tetep (Resistance check) | 1 | Resistant | |
| 4. | HR-12 (Susceptible check) | 9 | Susceptible | |
| 5. | IR121047-2-2-1 | 9 | Susceptible | |
| S. No. | Improved breeding lines (ICF ₆) | Score | Resistant/Moderately Resistant/Susceptible | |
| 1. | Swarna-IL-1 | 3 | Resistant | |
| 2. | Swarna-IL-2 | 3 | Resistant | |
| 3. | Swarna-IL-3 | 2 | Resistant | |
| 4. | Swarna-IL-4 | 3 | Resistant | |
| 5. | Swarna-IL-5 | 3 | Resistant | |
| 6. | Swarna-IL-6 | 3 | Resistant | |
| 7. | Swarna-IL-7 | 2 | Resistant | |
| 8. | Swarna-IL-8 | 3 | Resistant | |
| 9. | Swarna-IL-9 | 2 | Resistant | |
| 10. | Swarna-IL-10 | 2 | Resistant | |

Conclusion

The improved Swarna lines with resistance to bacterial leaf blight and blast, produced in this present study, could provide a significant benefit to farmers growing the Swarna rice variety in fields impacted by both diseases. The enhanced Swarna lines produced in this research can serve as donors to transmit resistance to BB and blast to other varieties.

Acknowledgements

The authors express gratitude to the Director of ICAR-IIRR for providing the necessary infrastructure for conducting this research. The authors appreciate the Department of Biotechnology for giving financial assistance for conducting the research on "Marker assisted introgression of yield enhancing genes to increase yield potential in rice" under grant No. BT/PR12168/AGR/2/894/2014.

References

- 1. Anila M, Mahadeva Swamy HK, Kale RR, *et al.* Breeding lines of the Indian mega-rice variety, MTU 1010, possessing protein kinase OsPSTOL (Pup1), show better root system architecture and higher yield in soils with low phosphorus. Mol Breeding. 2018;38:147. DOI: 10.1007/s11032-018-0903-1
- 2. Balachiranjeevi CH, Bhaskar Naik S, Abhilash Kumar V, *et al.* Marker-assisted pyramiding of two major, broad-spectrum bacterial blight resistance genes, *Xa21* and Xa33 into an elite maintainer line of rice, DRR17B. PLoS ONE. 2018;13(10):e0201271.
- 3. Balachiranjeevi CH, Bhaskar S, Abhilash V, *et al.* Marker-assisted introgression of bacterial blight and blast resistance into DRR17B, an elite, fine-grain type maintainer line of rice. Mol. Breeding, 2015, 35(151). DOI: 10.1007/s11032-015-0348-8
- 4. Chen S, Wang C, Yang J, *et al.* Identification of the novel bacterial blight resistance gene Xa46 (t) by mapping and expression analysis of the rice mutant H120. Sci. Rep. 2020;10:12642. DOI: 10.1038/s41598-020-69639-y
- Fiyaz RA, Shivani D, Chaithanya K, *et al.* Genetic Improvement of Rice for Bacterial Blight Resistance: Present Status and Future Prospects. Rice Science. 2022;9(2):118-132. DOI: 10.1016/j.rsci.2021.08.002
- 6. Guru P, Chhuneja N, Dixit A, Tiwari P, Kumar A. Mechanical transplanting of rice in India: Status, technological gaps and future thrust. ORYZA- An International Journal on Rice; c2018. p. 55.
- 7. Hari Y, Srinivasarao K, Viraktamath BC, *et al.* Markerassisted improvement of a stable restorer line, KMR-3R and its derived hybrid KRH2 for bacterial blight resistance and grain-quality. Plant Breeding. 2011;130(60):608-616.
- 8. Hari Y, Srinivasarao K, Basavaraj C, *et al.* Marker assisted introgression of bacterial blight and blast resistance into IR58025B, an elite maintainer line of rice. Plant Breeding. 2013;132(6):586-594.
- Nguyen HT, Vu QH, Mai TV, *et al.* Marker-Assisted Selection of *Xa21* Conferring Resistance to Bacterial Leaf Blight in indica Rice Cultivar LT2. Rice Science. 2018;25(1):52-56. DOI: 10.1016/j.rsci.2017.08.004
- Xing J, Zhang D, Yin F, *et al.* Identification and Fine-Mapping of a New Bacterial Blight Resistance Gene, Xa47(t), in G252, an Introgression Line of Yuanjiang

Common Wild Rice (*Oryza rufipogon*). An International Journal of Applied Plant Pathology; c2021. DOI: 10.1094/PDIS-05-21-0939-RE

- 11. Kauffman HE, Reddy APK, Hsieh SPY, Merca SD. An improved technique for evaluating resistance of rice varieties to *Xanthomonas oryzae*. Plant Dis Rep. 1973;56:537-540.
- Khush GS, Jena KK. Current status and future prospects for research on blast resistance in rice (*Oryza* sativa L.). In: Advances in Genetics, Genomics and Control of Rice Blast Disease; c2009. p. 1-10. DOI: 10.1007/978-1-4020-9500-9_1
- Kumar PN, Sujatha K, Laha GS, *et al.* Identification and fine mapping of Xa33, a novel gene for resistance to *Xanthomonas oryzae* pv. *Oryzae*. Phytopathology. 2012;102(2):222-228. DOI: 10.1094/PHYTO-03-11-0075
- 14. Mathur RS, Barnet HL, Lily VG. Sporulation of *Collectotrichum lindemuthianum* in culture. Phytopathol. 1950;40:104-114.
- 15. Ponnuswamy R, Singh AK, Raman MS, *et al.* Conversion of partial restorer Swarna into restorer by transferring fertility restorer Rf gene (s) through marker assisted back cross breeding (MABB) in rice. Sci. Rep. 2020;10:1101. DOI: 10.1038/s41598-020-58019-1
- 16. Sharma P, Bora LC, Puzari KC, *et al.* Review on Bacterial Blight of rice caused by *Xanthomonas oryzae* pv. *oryzae*: different management approaches and role of Pseudomonas fluorescens as a potential Bio. Control. agent. Int. J Curr Microbiol. App Sci. 2017;6(3):982-1005. DOI: 10.20546/ijcmas.2017.603.117
- 17. Prasad MS, Madhav MS, Laha GS, *et al.* Rice Blast Disease and Its Management. Hyderabad, India: Directorate of Rice Research (ICAR); c2011. p. 52.
- Preece TF, Rhodes ME, Skinner FA. Progression of bacterial disease within plants. In: Bacteria and plants, eds. Academic Press; c1982. p. 71-83.
- Rai AK, Kumar SP, Gupta SK, *et al.* Functional complementation of rice blast resistance gene Pi-k (h) (*Pi54*) conferring resistance to diverse strains of *Magnaporthe oryzae*. J Plant Biochem. Biotechnol. 2011;20:55-65. DOI: 10.1007/s13562-010-0026-1
- Ramkumar G, Srinivasa Rao K, Madhan Mohan K, *et al.* Development and validation of functional marker targeting an In Del in the major rice blast disease resistance gene *Pi54* (Pikh). Mol. Breeding. 2011;27:129-135.
- 21. Rekha G, Abhilash Kumar V, Viraktamath BC, *et al.* Improvement of blast resistance of the popular highyielding, medium slender-grain type, bacterial blight resistant rice variety, Improved Samba Mahsuri by marker-assisted breeding. J Plant Biochem Biotechnol. 2018;27:463-472. DOI: 10.1007/s13562-018-0455-9
- 22. Sahu PK, Sao R, Choudhary DK, *et al.* Advancement in the Breeding, Biotechnological and Genomic Tools towards Development of Durable Genetic Resistance against the Rice Blast Disease. Plants. 2022;11(18):2386. DOI: 10.3390/plants11182386
- 23. Sharma TR, Chauhan RS, Singh BM, *et al.* RAPD and pathotype analysis of Magnaporthe grisea population from North-western Himalayan region of India. J Phytopathol. 2002;150:649-656. DOI: 10.1046/j.1439-0434.2002.00812.x

- 24. Sharma TR, Rai AK, Gupta GK, Singh NK. Broad spectrum blast resistance gene Pikh cloned from the rice line Tetep designated as *Pi54*. J Plant Biochem Biotechnol. 2010;19:1.
- 25. Song WY, Wang GL, Chen LL, *et al.* A receptor kinase-like protein encoded by the rice disease resistance gene, *Xa21*. Science. 1995;270:1804-1806. DOI: 10.1126/science.270.5243.1804
- 26. Sundaram RM, Priya MRV, Laha GS, *et al.* Introduction of bacterial blight resistance into Triguna, a high yielding, mid-early duration rice variety by molecular marker assisted breeding. Biotechnology. 2009;4:400-407. DOI: 10.1002/biot.200800310
- 27. Sundaram RM, Vishnupriya MR, Biradar SK, *et al.* Marker assisted introgression of bacterial blight resistance in Samba Mahsuri, an elite indica rice variety. Euphytica. 2008;160:411-422.
- Tanweer FA, Rafii MY, Sijam K, *et al.* Current advance methods for the identification of blast resistance genes in rice. C R Biol. 2015;338:321-334. DOI: 10.1016/j.crvi.2015.03.001

Authors Details,

M Kousik

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

E Punniakotti

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

G Rekha ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

K Chaitra

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

T Dilip Kumar

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

SK Hajira ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

K Swapnil ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

SK Mastanbee

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

M Anila

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

M Ayyappadass ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India B Laxmi Prasanna

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

Pragya Sinha

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

Ravindra Kale

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

K Jyothi

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

G Vivek

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

D Aleena

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

P Senguttuvel

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

GS Laha

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

MS Prasad

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

P Sudhakar

Acharya Nagarjuna University, Guntur, Andhra Pradesh, India

A Krishna Satya

Acharya Nagarjuna University, Guntur, Andhra Pradesh, India

CN Neeraja

ICAR-Indian Institute of Rice Research, Hyderabad, Telangana, India

R Abdul Fiyaz

Senior Scientist, ICAR-Indian Institute of Rice Research, Rajendra Nagar, Hyderabad, Telangana, India

RM Sundaram

Director, ICAR-Indian Institute of Rice Research, Rajendra Nagar, Hyderabad, Telangana, India