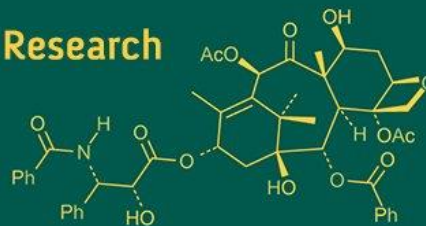
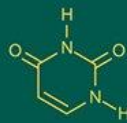
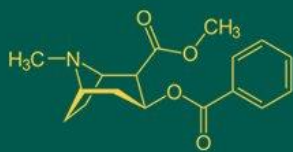


International Journal of Advanced Biochemistry Research



ISSN Print: 2617-4693
 ISSN Online: 2617-4707
 IJABR 2022; 6(2): 38-41
www.biochemjournal.com
 Received: 11-04-2022
 Accepted: 13-05-2022

Muhammad Torequl Islam
 Department of Pharmacy,
 Bangabandhu Sheikh Mujibur
 Rahman Science and
 Technology University,
 Gopalganj, Bangladesh

Anti-diabetic potential of *Ocimum sanctum* Linn.

Muhammad Torequl Islam

DOI: <https://doi.org/10.33545/26174693.2022.v6.i2a.132>

Abstract

Ocimum sanctum L., also known as Tulsi or the Holy Basil in Bangladesh and India. It is a vastly underutilized potential traditional medicinal plant. Its leaves and seed oil are used for treating many diseases, including coughs and colds. According to scientific evidence, it has a variety of traditional uses and can be used in oxidative stress, hypertension, cancers, neurological diseases and disorders, inflammation and inflammatory syndromes, and so on. These, this medicinal plant and its derivatives can be used in diabetes and metabolic syndromes. In this literature-based study, it has been seen that *O. sanctum* has promising anti-diabetic effects in various experimental models using mice, rats, rabbits etc. For its significant anti-diabetic effect, a tetracyclic triterpenoid known as 16-hydroxy-4, 4, 10, 13-tetramethyl-17-(4-methyl-pentyl)-hexadecahydro-cyclopenta[a]phenanthren-3-one has been introduced. More research is needed on its anti-diabetic effects.

Keywords: *Ocimum sanctum*, diabetes, metabolic syndromes, protective effects

Introduction

Diabetes mellitus, a complex metabolic disease which has significant harmful effects on humans. Diabetes is managed with conventional drugs in conjunction with lifestyle changes. They are not, however, completely successful, which is never recovered. Many medicinal plants have been used in many traditional medical systems worldwide to control diabetes mellitus because they are rich in biological elements. Several of them have been shown to be beneficial in the treatment of diabetes. Plants with hypoglycemic properties are becoming increasingly popular due to their inexpensive cost and lack of adverse effects. To date, hundreds of anti-diabetic medicinal herbs have been recognized (Khan *et al.*, 2012) [16].

Ocimum sanctum Linn. (Figure 1) (Family: Lamiaceae) is known to act against obesity and diabetes (Satapathy *et al.*, 2017) [27]. For this purpose, the plant is also used in combination as a polyherbal preparations with other medicinal plants (Kumar *et al.*, 2016a, b) [17-18].



Fig 1: Different parts of *Ocimum sanctum* Linn.

Corresponding Author:
Muhammad Torequl Islam
 Department of Pharmacy,
 Bangabandhu Sheikh Mujibur
 Rahman Science and
 Technology University,
 Gopalganj, Bangladesh

This review aims to summarize anti-diabetic effects of *O. sanctum* (also called Tulsi in Bangladesh and India) on the basis of available literature in the PubMed database.

***Ocimum sanctum* against diabetes**

Traditional remedies made from medicinal plants are used by around 67 percent of the world's population. Tulsi (*O. sanctum*) has diverse usage in Bangladesh and India (Bhattacharya *et al.*, 1997; Chattopadhyay, 1993, 1999; Vats *et al.*, 2002; Grover *et al.*, 2002; Kar *et al.*, 2003; Gholap and Kar, 2004; Narendhirakannan *et al.*, 2006; Modak *et al.*, 2007; De *et al.*, 2015) [2, 4-5, 21, 8, 15, 7, 23, 20, 6]. It can influence streptozotocin-induced changes in glycogen content and carbohydrate metabolism in animals (Vats *et al.*, 2004a) [32]. One report suggests that in mild to severe non-insulin dependent diabetes mellitus (NIDDM), the plant can be administered as an adjuvant to food therapy and pharmacological treatment (Agrawal *et al.*, 1996) [1].

Over a month, a 1% dose of the leaf powder reduced fasting blood sugar, uronic acid, total amino acids, total cholesterol (TC), triglyceride, phospholipids, and total lipids in normal and diabetic rats. TC and triglycerides were dramatically reduced in the liver. Total lipids were dramatically decreased in the liver and kidney. A substantial decrease in TC and phospholipids was seen in the heart. All of these findings show that *O. sanctum* has a hypoglycemic and hypolipidemic effects on the experimental animals (Rai *et al.*, 1997). Fresh leaf supplementation (2 g/kg, b.w. for 30 days) when compared to the control group, considerably reduced blood glucose levels and raised superoxide dismutase (SOD), reduced glutathione (GSH), and total thiol levels, while dramatically reduced peroxidised lipid levels (Sethi *et al.*, 2004) [28]. The leaves have been demonstrated to offer a wide range of potential advantages in a number of stressful situations. In one research, the leaf fixed oil was shown to exert considerable hypoglycemic, lipid lowering, and antioxidant activities in streptozotocin-induced diabetic rats. These effects might be attributed to the alpha-linolenic acid found in the oil (Suanarunsawat *et al.*, 2015) [30]. The leaf extract (500 mg/kg b.w., p.o. for 15 days) shown antidyslipidemic and antioxidant activity, suggesting that it might be utilized to avoid diabetic dyslipidemia and accompanying problems (Husain *et al.*, 2015) [12]. In comparison to the control group, seed oil (0.8 g/kg, b.w. /day for four weeks) significantly reduced blood cholesterol, triacylglycerol, and low-density lipoprotein and very low-density lipoprotein (LDL+VLDL)-cholesterol in alloxan-induced diabetic rabbits (Gupta *et al.*, 2006) [9].

Chloroform extract (250 and 500 mg/kg) of *O. sanctum* areal parts when compared to untreated alloxan-induced diabetic mice, dramatically reduced the raised level of blood glucose and generated a reversal of cholesterol, triglyceride, high-density lipoprotein (HDL), and LDL levels (Patil *et al.*, 2011b) [25]. The administration of Tulsi to streptozotocin-induced diabetic rats for 30 days significantly reduced plasma thiobarbituric acid reactive substances (TBARS) levels and improved the status of the antioxidant enzymes catalase, SOD, and glutathione peroxidase in vital organs such as the liver and kidney (Muralikrishnan *et al.*, 2012) [21]. Methanolic extract of Tulsi at 500 mg/kg (p.o.) reduced diabetic manifestations/dysregulations in both non-diabetic and alloxan-induced diabetes in adult female Wistar rats (Singh *et al.*, 2012) [29].

Oral treatment *O. sanctum* extract at 500 mg/kg (b.w.) reduced the glucose level in blood as well as in streptozotocin-induced diabetic mice, it prevented the generation of lipid peroxides, reactivated antioxidant enzymes, and restored GSH and metal levels (Chandra *et al.*, 2008) [3]. Tetracyclic triterpenoid [16-hydroxy-4,4,10,13-tetramethyl-17-(4-methyl-pentyl)-hexadecahydro-cyclopenta[a]phenanthren-3-one] was isolated from the hydro alcoholic extract of *O. sanctum* aerial part was found to exert a significant anti-diabetic effect on the test system (Patil *et al.*, 2011a) [24].

Certain minerals (e.g., Cu, Ni, Zn, K, Na, Fe, Cr, V) are linked to beneficial effects in diabetes (Narendhirakannan *et al.*, 2005) [22]. Insulin resistance has been linked to zinc deficiency. Tulsi contains zinc, which may have a key role against insulin resistance in animals. Tulsi can increase activity of glucokinase, hexokinase, and phosphofructokinase. It might have a far larger role in both diabetes and other metabolic syndromes. Tulsi has been shown in studies to lower blood cholesterol and LDL levels while also lowering serum triglycerides and VLDL. Furthermore, Tulsi contains potent protective properties (e.g., anti-microbial, antioxidant, anti-inflammatory) that can prevent the abnormal oxidation and inflammatory processes in metabolic syndromes (Kapoor, 2008) [14].

Tulsi aqueous extract substantially decreased plasma glucose, HbA(1c), lipid profile, and lipid peroxidation (LPO), while increasing glutathione peroxidase (GPx), SOD, catalase (CAT), and glutathione-S-transferase (GST). Most of the measures tested, including plasma glucose levels, were reversed after 16 weeks of treatment with Tulsi and/or Vitamin E in diabetic animals (Halim and Mukhopadhyay, 2006) [10]. On the other hand, the alcoholic extract and five fractions of Tulsi leaves stimulated insulin secretion in experimental animals (Hannan *et al.*, 2006) [11]. *O. sanctum* aqueous extract (200 mg/kg, p.o.) mixed with diet for eight weeks in diabetic (streptozotocin-induced) rats considerably decreased fasting blood glucose, serum lipid profile, lipid peroxidation products, and glucose tolerance. It also boosted antioxidant enzymes SOD, CAT, GPx, glutathione transferase (GT), and one antioxidant GSH in plasma and rat liver, lung, kidney, and brain (Hussain *et al.*, 2001) [13]. In one research, alloxan-induced diabetic rats were fed an aqueous extract of Tulsi leaves at a dose of 200 mg/kg/day until cataracts developed. Serum glucose and body weight were measured on a regular basis, and cataracts were checked with the naked eye and a slit light 75, 100, and 115 days after alloxan administration. All three plant extracts had a positive effect on body weight and blood glucose, indicating that this plant has strong antihyperglycemic action (Vats *et al.*, 2004b) [33].

The function of silver nanoparticles (AgNps) in current diabetic therapeutics and applied research is an appealing idea. Using aqueous leaf extracts of *O. sanctum*, stable AgNps with sizes ranging from 3 to 25 nm were produced. The concentration of the extract assisted in the reduction of silver nitrate, resulting in the rapid formation of AgNps at room temperature, exhibiting a faster reaction rate when compared to harsh chemical procedures and high conversion energy generally associated with the synthesis. The AgNps produced from Tulsi inhibited the *Bacillus Stearothermophilus* alpha-glucosidase enzyme model by 89.31 5.32 percent, showing an improved bio-catalytic

potential as compared to its crude extract and the control (Malapermal *et al.*, 2017) [19].

Conclusion

O. sanctum has protective functions such as anti-microbial, antioxidant, and anti-inflammatory properties that may have an important role in the therapy of diabetes and metabolic disorders. Doctors should be alert of the medicinal plant's therapeutic properties and should advocate its frequent inclusion in the daily diet, particularly for diabetics and those with metabolic disorders.

Conflict of interest: None declared.

References

1. Agrawal P, Rai V, Singh RB. Randomized placebo-controlled, single blind trial of holy basil leaves in patients with noninsulin-dependent diabetes mellitus. *Int J Clin Pharmacol Ther.* 1996;34(9):406-9.
2. Bhattacharya SK, Satyan KS, Chakrabarti A. Effect of Trasina, an Ayurvedic herbal formulation, on pancreatic islet superoxide dismutase activity in hyperglycaemic rats. *Indian J Exp Biol.* 1997;35(3):297-9.
3. Chandra A, Mahdi AA, Singh RK, Mahdi F, Chander R. Effect of Indian herbal hypoglycemic agents on antioxidant capacity and trace elements content in diabetic rats. *J Med Food.* 2008;11(3):506-12. Doi: 10.1089/jmf.2007.0042.
4. Chattopadhyay RR. Hypoglycemic effect of *Ocimum sanctum* leaf extract in normal and streptozotocin diabetic rats. *Indian J Exp Biol.* 1993;31(11):891-3.
5. Chattopadhyay RR. A comparative evaluation of some blood sugar lowering agents of plant origin. *J Ethnopharmacol.* 1999;67(3):367-72.
6. De B, Bhandari K, Singla RK, Katakam P, Samanta T, Kushwaha DK, Gundamaraju R, Mitra A. Chemometrics Optimized Extraction Procedures, Phytosynergistic Blending and *in vitro* Screening of Natural Enzyme Inhibitors Amongst Leaves of Tulsi, Banyan and Jamun. *Pharmacogn Mag.* 2015;11(4):S522-32. Doi: 10.4103/0973-1296.172956.
7. Gholap S, Kar A. Hypoglycaemic effects of some plant extracts are possibly mediated through inhibition in corticosteroid concentration. *Pharmazie.* 2004;59(11):876-8.
8. Grover JK, Yadav S, Vats V. Medicinal plants of India with anti-diabetic potential. *J Ethnopharmacol.* 2002;81(1):81-100.
9. Gupta S, Mediratta PK, Singh S, Sharma KK, Shukla R. Antidiabetic, antihypercholesterolaemic and antioxidant effect of *Ocimum sanctum* (Linn) seed oil. *Indian J Exp Biol.* 2006;44(4):300-4.
10. Halim EM, Mukhopadhyay AK. Effect of *Ocimum sanctum* (Tulsi) and vitamin E on biochemical parameters and retinopathy in streptozotocin induced diabetic rats. *Indian J Clin Biochem.* 2006;21(2):181-8. Doi: 10.1007/BF02912939.
11. Hannan JM, Marenah L, Ali L, Rokeya B, Flatt PR, Abdel-Wahab YH. *Ocimum sanctum* leaf extracts stimulate insulin secretion from perfused pancreas, isolated islets and clonal pancreatic beta-cells. *J Endocrinol.* 2006;189(1):127-36.
12. Husain I, Chander R, Saxena JK, Mahdi AA, Mahdi F. Antidyslipidemic Effect of *Ocimum sanctum* Leaf Extract in Streptozotocin Induced Diabetic Rats. *Indian J Clin Biochem.* 2015;30(1):72-7. Doi: 10.1007/s12291-013-0404-2.
13. Hussain EH, Jamil K, Rao M. Hypoglycaemic, hypolipidemic and antioxidant properties of tulsi (*Ocimum sanctum* linn) on streptozotocin induced diabetes in rats. *Indian J Clin Biochem.* 2001;16(2):190-4. Doi: 10.1007/BF02864859.
14. Kapoor S. *Ocimum sanctum*: A therapeutic role in diabetes and the metabolic syndrome. *Horm Metab Res.* 2008;40(4):296. Doi: 10.1055/s-2008-1058102.
15. Kar A, Choudhary BK, Bandyopadhyay NG. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. *J Ethnopharmacol.* 2003;84(1):105-8.
16. Khan V, Najmi AK, Akhtar M, Aqil M, Mujeeb M, Pillai KK. A pharmacological appraisal of medicinal plants with antidiabetic potential. *J Pharm Bioallied Sci.* 2012;4(1):27-42. Doi: 10.4103/0975-7406.92727.
17. Kumar D, Trivedi N, Dixit RK. Evaluation of the potential effect of *Allium sativum*, *Momordica charantia*, *Eugenia jambolana*, *Ocimum sanctum*, and *Psidium guajava* on intestinal p-glycoprotein in rats. *J Intercult Ethnopharmacol.* 2016a;6(1):68-74. Doi: 10.5455/jice.20160902023435.
18. Kumar D, Trivedi N, Dixit RK. Evaluation of the synergistic effect of *Allium sativum*, *Eugenia jambolana*, *Momordica charantia*, *Ocimum sanctum*, and *Psidium guajava* on hepatic and intestinal drug metabolizing enzymes in rats. *J Intercult Ethnopharmacol.* 2016b;5(4):372-382.
19. Malapermal V, Botha I, Krishna SBN, Mbatha JN. Enhancing antidiabetic and antimicrobial performance of *Ocimum basilicum*, and *Ocimum sanctum* (L.) using silver nanoparticles. *Saudi J Biol Sci.* 2017;24(6):1294-1305. Doi: 10.1016/j.sjbs.2015.06.026.
20. Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TP. Indian herbs and herbal drugs used for the treatment of diabetes. *J Clin Biochem Nutr.* 2007;40(3):163-73. Doi: 10.3164/jcfn.40.163.
21. Muralikrishnan G, Pillai SK, Shakeel F. Protective effects of *Ocimum sanctum* on lipid peroxidation and antioxidant status in streptozotocin-induced diabetic rats. *Nat Prod Res.* 2012;26(5):474-8. Doi: 10.1080/14786419.2010.531016
22. Narendhirakannan RT, Subramanian S, Kandaswamy M. Mineral content of some medicinal plants used in the treatment of diabetes mellitus. *Biol Trace Elem Res.* 2005;103(2):109-15.
23. Narendhirakannan RT, Subramanian S, Kandaswamy M. Biochemical evaluation of antidiabetogenic properties of some commonly used Indian plants on streptozotocin-induced diabetes in experimental rats. *Clin Exp Pharmacol Physiol.* 2006;33(12):1150-7.
24. Patil R, Patil R, Ahirwar B, Ahirwar D. Isolation and characterization of anti-diabetic component (bioactivity-guided fractionation) from *Ocimum sanctum* L. (Lamiaceae) aerial part. *Asian Pac J Trop Med.* 2011a;4(4):278-82. Doi: 10.1016/S1995-7645(11)60086-2
25. Patil RN, Patil RY, Ahirwar B, Ahirwar D. Evaluation of antidiabetic and related actions of some Indian medicinal plants in diabetic rats. *Asian Pac J Trop Med.*

- 2011b;4(1):20-3. Doi: 10.1016/S1995-7645(11)60025-4.
26. Rai V, Iyer U, Mani UV. Effect of Tulasi (*Ocimum sanctum*) leaf powder supplementation on blood sugar levels, serum lipids and tissue lipids in diabetic rats. *Plant Foods Hum Nutr.* 1997;50(1):9-16.
 27. Satapathy S, Das N, Bandyopadhyay D, Mahapatra SC, Sahu DS, Meda M. Effect of Tulsi (*Ocimum sanctum* Linn.) Supplementation on Metabolic Parameters and Liver Enzymes in Young Overweight and Obese Subjects. *Indian J Clin Biochem.* 2017;32(3):357-363. Doi: 10.1007/s12291-016-0615-4.
 28. Sethi J, Sood S, Seth S, Talwar A. Evaluation of hypoglycemic and antioxidant effect of *Ocimum sanctum*. *Indian J Clin Biochem.* 2004;19(2):152-5. Doi: 10.1007/BF02894276.
 29. Singh PK, Baxi D, Banerjee S, Ramachandran AV. Therapy with methanolic extract of *Pterocarpus marsupium* Roxb and *Ocimum sanctum* Linn reverses dyslipidemia and oxidative stress in alloxan induced type I diabetic rat model. *Exp Toxicol Pathol.* 2012;64(5):441-8. Doi: 10.1016/j.etp.2010.10.011.
 30. Suanarunsawat T, Anantasomboon G, Piewbang C. Anti-diabetic and anti-oxidative activity of fixed oil extracted from *Ocimum sanctum* L. leaves in diabetic rats. *Exp Ther Med.* 2016;11(3):832-840.
 31. Vats V, Grover JK, Rathi SS. Evaluation of anti-hyperglycemic and hypoglycemic effect of *Trigonella foenum-graecum* Linn, *Ocimum sanctum* Linn and *Pterocarpus marsupium* Linn in normal and alloxanized diabetic rats. *J Ethnopharmacol.* 2002;79(1):95-100.
 32. Vats V, Yadav SP, Grover JK. Ethanolic extract of *Ocimum sanctum* leaves partially attenuates streptozotocin-induced alterations in glycogen content and carbohydrate metabolism in rats. *J Ethnopharmacol.* 2004a;90(1):155-60.
 33. Vats V, Yadav SP, Biswas NR, Grover JK. Anti-cataract activity of *Pterocarpus marsupium* bark and *Trigonella foenum-graecum* seeds extract in alloxan diabetic rats. *J Ethnopharmacol.* 2004b;93(2-3):289-94.