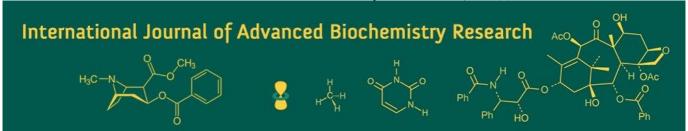
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Endophytic microbial advancement through NGS technology: Unlocking the power of the genome

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Abstract

The application of endophytic microorganisms for the protection of plant against to several diseases represents a significant contribution in sustainable agriculture. These endophytes live within internal plant parts and interact to their surrounds without causing any harmful effect and establishment through symbiotic relationship that can induce plant defence system. Endophytic bacteria and fungi have shown not only inhibit a wide range of phytopathogens, viz., bacteria, fungi, and viruses, through the production of bioactive metabolites substances, competition for habitat, space and nutrients, and the activation of plant immune responses but also actively participated to plant growth promoting activities like secretion of phytohormone, enhancing nutrient uptake and influences plant biochemical pathway. This review represents a scenario of the different types of endophytes, their mechanism of action as biocontrol agents and biofertilizers successful case studies, and their role in integrated pest and disease management systems with plant growth promotion and additionally, discusses the recent advanced Next Generation Technology, *viz.*, next generation sequencing, multi-omics studies for endophytic microbial identification and research, challenges, limitations, regulation and future scopes for the commercialization and application of endophytes as a biocontrol agents and biofertilizers to support sustainable agriculture in crop improvement with ecological resilience.

Keywords: Endophytic microorganisms, phytopathogens, sustainable agriculture, advanced NGS technology, bioactive metabolites

Introduction

Plant health is critically interconnected with microbial community that both above and below soil (Trivedi *et al.*, 2020; Van Elsas *et al.*, 2019) [194, 198]. Among the mega diversity of plant-associated microorganisms form of endophytes (Ali *et al.*, 2021; Afzal *et al.*, 2019) [16, 7], these organisms that reside internally within plant tissues leaves, stem and root (Mishra *et al.*, 2021) [175], and represent an underexplored frontier in microbial ecology and agricultural biotechnology (Dubey *et al.*, 2020) [61]. Endophytes has grown exponentially in recent years (Rana *et al.*, 2023) [11], not only for their plant growth-promoting activities (Srivastava *et al.*, 2024) [187], but also specially for their ability to work as natural biocontrol agents (Bhardwaj *et al.*, 2023; Baron *et al.*, 2022) [32, 25] for inhibit harmful virulent phytopathogens (Bhagya *et al.*, 2019; Emitaro *et al.*, 2024) [31, 67].

Today's agricultural sector is facing several types of biotic challenges (Anand *et al.*, 2023; Chaudhary *et al.*, 2022) [17, 45], including the rapid evolution like novel strain are developed of pest and phytopathogen (Anand *et al.*, 2023; Anand *et al.*, 2019; Adeleke *et al.*, 2022; Kumar *et al.*, 2021) [17, 18, 4, 202], control to conventional agrochemicals (Adetunji *et al.*, 2022; Bamisile *et al.*, 2021; Dubey *et al.*, 2020) [5, 23, 61], results, loss of soil-crop biodiversity (El-Saadony *et al.*, 2022) [66], and exceeded vulnerability of crops due to monoculture strategies (Bharti *et al.*, 2025; Negi *et al.*, 2025) [33, 134]. Chemical control measures are widely adapted, while effective in the short term, often environmental contamination, health risks, and nontarget effects, thereby necessitating more sustainable alternatives (Sharma *et al.*, 2025) [180]. Endophytic microorganisms especially endophytic bacteria and fungi (Akram *et al.*, 2023) [13] have emerged as potent in curing plant health (Wijesekara and Xu, 2023) [208], enhancing plant growth, and suppressing diseases (Basit *et al.*, 2021; Eid *et al.*, 2021) [27, 64] through enhance plant defence and eco-friendly mechanisms (Hardoi*m et al.*, 2015) [81].

Endophytes are group of different types of microbes that colonize and establish, internal plant tissues (Kobayashi and Palumbo, 2000; Mengistu, 2020) [102, 125] without causing any apparent disease (Carroll, 1986; Petrini, 1991) [40, 155]. They are exhibiting numerous properties and play a crucial role in promoting plant growth, inducing stress tolerance (Shaffique et al., 2022; Ullah et al., 2019) [178, 196], and offering protection against harmful pathogens (Rabiae et al., 2025; Radouane et al., 2024) [163, 164], thereby contributing significantly and perform to enhance plant health and productivity (Tarig *et al.*, 2025; Pandey *et al.*, 2023) [191, 179]. Their internal colonization permits them to interact intimately with the plant's metabolic and defence responses, enabling both localized and systemic benefits (Riseh et al., 2025; Ryan et al., 2008) [167, 171], further enhances their appeal for inclusion in integrated pest and nutrient management practices (Sena et al., 2024) [177], especially under organic or resource-conserving agricultural models (Kandel, 2016; Santoyo et al., 2016) [93, 176].

From a biological control point of view, endophytes have antagonistic biotic effects on plant pathogens through several well-characterized mechanisms (Fite et al., 2023; Doty et al., 2022; Pandit et al., 2022) [68, 60, 145]. For example, the production of antimicrobial compounds e.g., antibiotics, lipopeptides, volatile organic compounds (Nimbulkar et al., 2025; Prabhu et al., 2025; Narayanasamy et al., 2023; Panwar et al., 2021) [138, 158, 130, 147], induction of host defence through systemic resistance competition for ecological niches and nutrients (Gulyamova et al., 2025; Mamarasulov et al., 2025) [120], enzymatic degradation of phytopathogen cell walls (Thomas et al., 2024; Gagné-Bourque, 2015) [192, 70], and siderophores production (Chagas et al., 2018) [42] that sequester essential micronutrients like iron (Compant et al., 2005; Pieterse et al., 2014) [51, 156].

The earth's surface diversity of endophytes is vast yet widely underexplored (Sivalingam *et al.*, 2024; Dos Reis *et al.*, 2022; Aghdam and Brown, 2021; Harrison *et al.*, 2020) [185, 59, 8, 84], as traditional culture-dependent methods often don't cover to their hidden complexity (Mametja *et al.*, 2025) [121]. Chaurasia *et al.* (2018) [46] suggested that *Actinomycetes*: an unexplored microorganism for plant growth promotion and biocontrol in vegetable crop.

According to Kayode *et al.* (2025) ^[98] previous discoveries have denoted that nearly 300,000 species worldwide-harbour one or more endophytic microbial communities. Endophytes live multiple domains of life, including bacteria, fungi, actinomycetes, and even archaea. Among bacterial endophytes, diversity is commonly dominated by members of the phylum *Proteobacteria* (alpha, beta, and gamma classes) (Anand *et al.*, 2023) ^[17], with the most frequently found genera like *Bacillus*, *Pseudomonas*, *Enterobacter*, *Streptomyces*, *Klebsiella*, *Azospirillum*, and *Rhizobium* (Ahmed *et al.*, 2024). These organisms may be Gramnegative rods or Gram-positive spore-formers (Beskrovnaya)

et al., 2021) [30] and exhibit high metabolic mechanism (Soni and Keharia, 2021) [186] and can survive in the dynamic internal environments of plant tissues (Andryukov et al., 2020; Van Elsas et al., 2019) [19, 198]. Fungal endophytes dominated by the genera such as *Penicillium*, *Trichoderma*, *Neotyphodium Fusarium* non-pathogenic strains, *Cladosporium*, and *Aspergillusm* (Grabka et al., 2022; Fontana et al., 2021; Al-Ani et al., 2019) [75, 69, 15], these genera have shown the ability to confer stress tolerance and enhance pathogen resistance in host plants (Deepa et al., 2024; Akram et al., 2023; Arnold et al., 2001) [56, 13, 21].

Tariq et al. (2025) [191] strongly suggested that nowadays advances in Next-Generation Sequencing technologies have revolutionized the identify of these symbiotic endophytic microbes by overcoming the limitations and challenges of traditional culture-dependent techniques. NGS offers a powerful base for exploring the genomic, transcriptomic, and functional potential of endophytic microbes (Shishodia et al., 2025; Verma et al., 2024; Bielecka et al., 2022; Chen et al., 2022; Wani et al., 2022; Bosamia et al., 2020) [18, 203, ^{34, 48, 207, 38]} at unprecedented depth. It has become a powerful platform in modern life sciences, investigating innovations in consumer genomics, translational research, and diagnostics at molecular level (Kaur et al., 2024; Sagita et al., 2021; Kaul et al., 2016) [133, 173, 96]. Platforms such as Illumina NGS enable high-throughput sequencing (Prasanna et al., 2022) [161] that reveals genetic variations, taxonomic diversity, and metabolic pathways within microbial communities (Patel et al., 2025; Deepa et al., 2024) [149, 56]. These tools have analysed that each plant species harbours a unique core microbiome that can be altered by genotype (Maestro-Gaitán et al., 2025: Tabassum et al., 2024: Latz et al., 2021; Mina et al., 2020) [118, 190, 109, 126], developmental stage (Bintarti et al., 2022) [35], environmental conditions (Sahu and Mishra, 2021), and agricultural practices (Berg et al., 2014) [29].

According to Deepa *et al.* (2024) ^[56] through metagenomic, meta-transcriptomic, and functional genomic approaches, NGS enables the discovery of novel genes, biosynthetic pathways, and secondary metabolites with potential biotechnological applications. Additionally, the integration of Next Generation Sequencing technology with microbial biotechnology opens new doors for crop improvement in sustainable agriculture by developing the biofertilizers, biocontrol, and stress-resilient crop varieties presented in Figure 1.

This review therefore focuses the endophytic microbial advancement through NGS technology: unlocking the power of the genome", highlighting on scenario of endophytes, the interconnected with plants surrounds, advantages, their limitations and challenges, how to identified their potential genes or traits or endophytes through Next Generation Sequencing technologies-driven discoveries are shaping their application through case studies in agricultural microbial biotechnology and beyond.

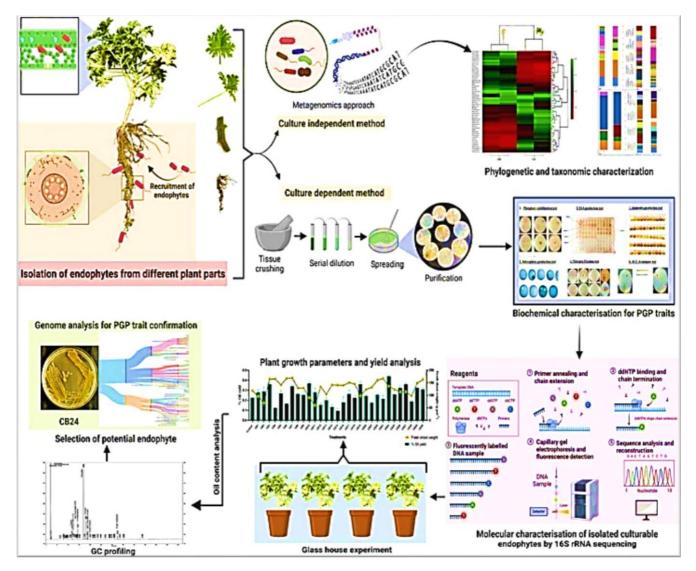


Fig 1: Endophytic Microbial Analysis through Traditional Culture-Dependent Methods and NGS Technology (Adapted from Deepa *et al.* 2024)

Endophytic Microbes

Endophytic microorganisms are having a taxonomically wide range and ecologically diverse group that colonize (Nair et al., 2014) [129] and establish the internal plant tissues without causing any disease symptoms or damages (Harrison and Griffin, 2020) [83]. These endophytes occupy a crucial ecological niche, symbiotic relationship (Kuźniar et al., 2025; Zou et al., 2025) [108, 218] that can wide range from mutualistic to commensal (Bard et al., 2024; El-Metwally et al., 2023) [24, 65] and sometimes opportunistic (Mishra et al., 2021), based on the host physiology, biochemical and environmental conditions (Hajji-Hedfi et al., 2025; Hakansson et al., 2018) [78, 79]. Endophytes are not only widespread across terrestrial and aquatic plant (Lazar et al., 2022) [110] but also are recognized as integral form of the plant holobiont systems (Pramanic et al., 2023; Durand et al., 2021; L'Hoir et al., 2021; Lyu et al., 2021) [160, 62, 111, 116] a functional unit forming the host and its relation with microbial communities (Grzyb et al., 2024; Anand et al., 2023; Dittami et al., 2021; Khare et al., 2018) [76, 17, 57, 101].

According to Vasileva *et al.* (2019) [199] some common endophytic bacterial genera isolated from agronomic plants reported in literature and found that diversity of endophytes in leguminous plants are more as comparison to non-leguminous plants.

Verma *et al.* (2021) ^[202] revealed that potential application of endophytes is stress controller to manangement of abiotic stress from drought and salinity in crop plants, through different mechanism.

Plant-Microbial relationships at phyllo-spheric and rhizospheric zone

Zhao *et al.* (2024) ^[217] reported that endophytic microbes and rhizospheric microbes are efficient to promote growth of the plants directly as well as by indirectly through increasing macronutrient and mineral uptake, plant protection against pathogens respectively presented in Figure 2. Naturally synthesized bioactive compounds with antimicrobial activities can be exploited in various sectors, especially in the Agricultural Biotechnology sectors.

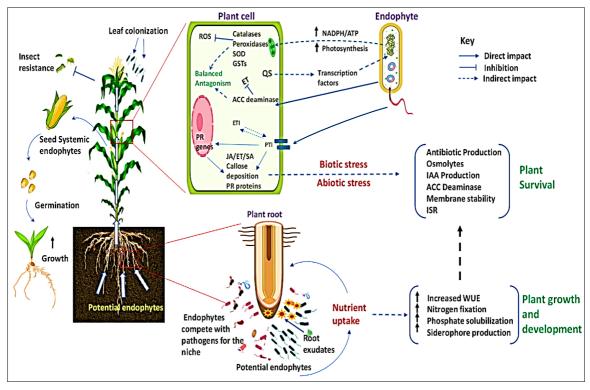


Fig 2: Mechanisms of Plant-Endophytic Microbial-Interaction (Adapted from Zhao et al. 2024)

Timeline of Endophyte Research and identification

Endophytes were first described by J. H. Friedrich Link, the German botanist, in 1809. They were about plant parasitic fungi. the French scientist Béchamp were later coined the term as "microzymas". There was a strongly belief that

plants were healthy under sterile conditions and it was not until 1887 that Victor Galippe identified bacteria normally occurring inside plant tissues (Hardoim *et al.*, 2015) ^[81], history of endophytic microorganism shown in Figure 3.

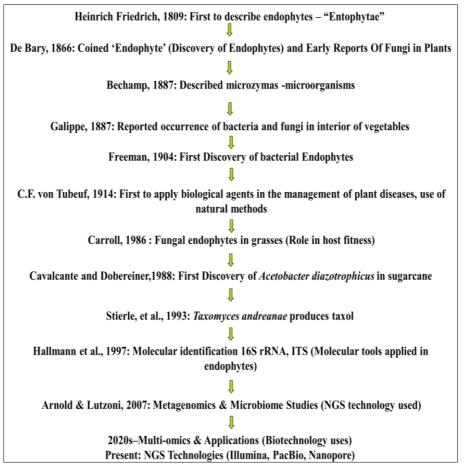


Fig 3: A schematic representation, timeline of endophytic microbial research and identification in 1809 up to present year

Classification of Endophytic Microorganisms

According to Das *et al.* (2025) [159] obligate endophytes rely redefined as entirely on the host plant for growth, development, establishments, and reproduction without host can't survive. These are often associated in long-time, coevolved relationships with specific plant lineages. In case of

facultative endophytes are colonizing plant tissues under favourable conditions and free-living in the soil or rhizosphere under an unfavourable condition. Most bacterial endophytic species belong to this category (Compant *et al.*, 2005; Compant and Vacher, 2019) [51, 52], presented in Table 1

Table 1: Classification of Endophytic Microorganisms in different Categories with examples (Das et al., 2025) [159]

Basis of Classification	Category	Examples
On the Basis of Microbial Type	Bacterial Endophytes	Pseudomonas, Bacillus, Enterobacter, Klebsiella, Streptomyces
	Fungal Endophytes	Epichloë, Trichoderma, Fusarium, Aspergillus, Penicillium
On the Basis of Colonization Pattern	Obligate Endophytes	Epichloë coenophiala, Arbuscular mycorrhizal fungi
	Facultative Endophytes	Pseudomonas fluorescens, Trichoderma harzianum
Transmission Mode	Vertical Transmission	Epichloë spp. in grasses
	Horizontal Transmission	Fusarium, Alternaria, Colletotrichum
Symbiotic Relationship	Mutualistic Endophytes	Trichoderma spp., Azospirillum brasilense
	Commensal Endophytes	Some Penicillium spp.
	Latent/Opportunistic Endophytes	Fusarium oxysporum (non-pathogenic strains)
On the Basis of Fungal Ecological Classes	Class1 (Clavicipitaceous)	Epichloë coenophiala (seed transmitted in tall fescue)
	Class 2 (non-clavicipitaceous)	Trichoderma, Alternaria, Fusarium (broad host range, colonize
		roots/shoots)
	Class 3 (non-clavicipitaceous)	Colletotrichum, Ustilago (above-ground limited colonization)
	Class 4 (non-clavicipitaceous)	Phialocephala, Chloridium (root-restricted, melanized)

Transmission Mode of Endophytic microorganism

Endophytes can enter plants tissue or cell through natural openings like hydathodes or stomata and lenticels, as well as through wounds on the root surface or tips (Mattoo and Nonzom *et al.*, 2020; Kumar *et al.*, 2020) [124, 92]. Once inside, they colonize intercellular spaces, vascular tissues and sometimes even intracellular compartments, depending

on host-microbe mutualistic relation (Khan *et al.*, 2025; Stotz *et al.*, 2022) [100, 195], presented in Figure 4. According to Ahlawat *et al.* (2022) [9] colonization patterns are affected by both microbial traits (e.g., motility, adhesion, quorum sensing) and host factors (*e.g.*, immune responses, secondary metabolite production).

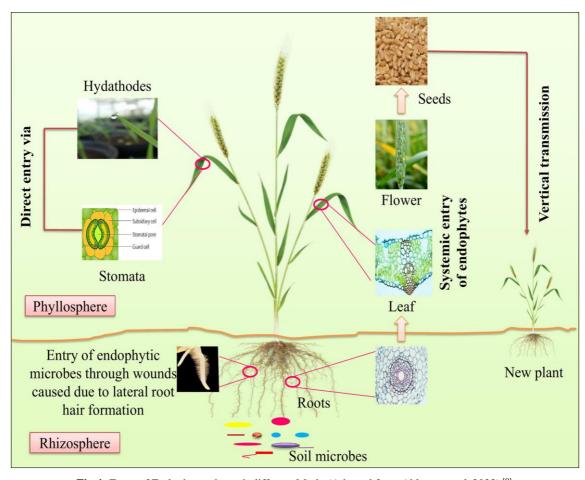


Fig 4: Entry of Endophytes through different Mode (Adapted from Ahlawat et al. 2022) [9]

Diversity Drivers

Anand *et al.* (2023) ^[17] reported the cluster data analysis was performed by "VOS viewer version 1.6.16," 2020 represented in Figure 5. The map highlights the most frequently used bibliographic terms to understand the most active research areas and results can be grouped into five clusters. The first cluster is represented by green balls exhibit 241 items, and devoted to stress factors and adaptation. Keywords are related to abiotic stress (*e.g.*, salinity and drought) and biotic (pathogen). Cluster 2 is represented by blue balls exhibit 224 items, keywords highlight the reactivity of endophytes, the endophytic

production of metabolites, and the antibacterial activity of the obtained bioactive compounds. The third cluster is highlighted by yellow balls (192 items), and mainly concerns colonization mechanisms, with several keywords devoted to culture, and bacterial and fungi growth. Cluster 4 is represented by violet balls, 75 items, and devoted to remediation, with keywords related to contamination and detoxification. The keywords contaminants denote to heavy metals and organic pollutants. Finally, cluster 5 is exhibit 168 items, represented by red balls, mainly devoted to genome and genetic expressions.

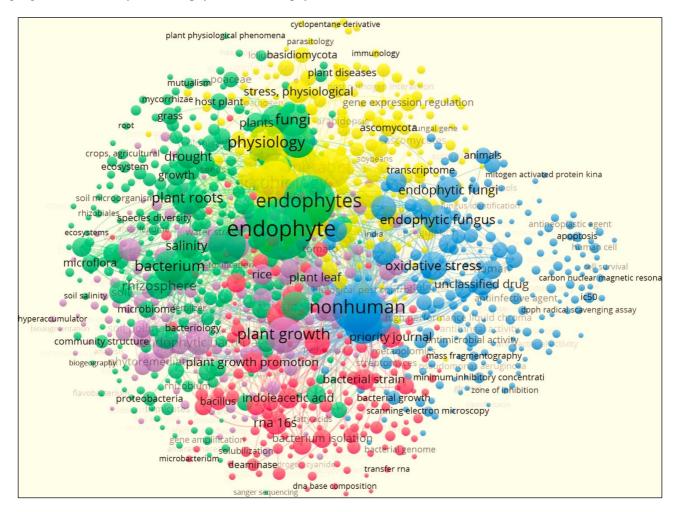


Fig 5: Reports the cluster analysis provided from the co-occurrence network of keywords of the papers extracted from the SCOPUS platform (Adapted from Anand *et al.* 2023) [17]

Endophytic community consisted by a range of biotic and abiotic factors (Kandel, 2016) [93].

Plant genotype and species: The host plant genome makeup affects its root exudates substance and physicochemical process, which in turn modulate according to microbial needs.

Tissue specificity: Endophytic populations vary with its living plant parts like roots, stems, leaves, flowers, and seeds, each offering unique microhabitats.

Developmental stage: The successional dynamics of plant growth affect microbial turnover and taxonomical functional.

Environmental variables: Soil types, pH, EC, temperature, moisture level, and agricultural management practices *e.g.*, chemical pesticide and fertilizer use, crop rotation and patterns significantly influence endophytic functions.

Advantages of using Endophytic microorganism

Endophytic microorganisms exhibit advantages through a complicated and multifaceted array of mechanisms, dealing with direct antagonism, boost of plant immunity, and competitive ecological interactions (Lu *et al.*, 2021) [189]. These mechanisms are the result of millions of years of coevolution between plants and their associated microbiota and reflect both microbial lifestyle and ecological resilience. The studying of these association at a molecular, cellular and physiological level is important for making effective biological control strategies in modern agriculture,

presented in Figure 6 (Tariq *et al.*, 2025) ^[191]. Chaudhary *et al.* (2022) ^[45], and Kumar *et al.* (2022) ^[45] suggested that endophytes are promising biofertilizers in crop improvement for agriculture production. Field-level applications of endophytic biocontrol agents have observed promising results under natural disease pressures. Aarthi *et al.* (2025) ^[1] conducted an experiment in both greenhouse and field

condition, application of a consortium of *Bacillus subtilis* and *Trichoderma viride* significantly minimized the incidence of disease like damping-off, root rot, and leaf spot in vegetable crops. There results strongly suggested that consortium of *Bacillus subtilis* and *Trichoderma viride* acted synergistically, providing broad-spectrum control and promoting plant growth.

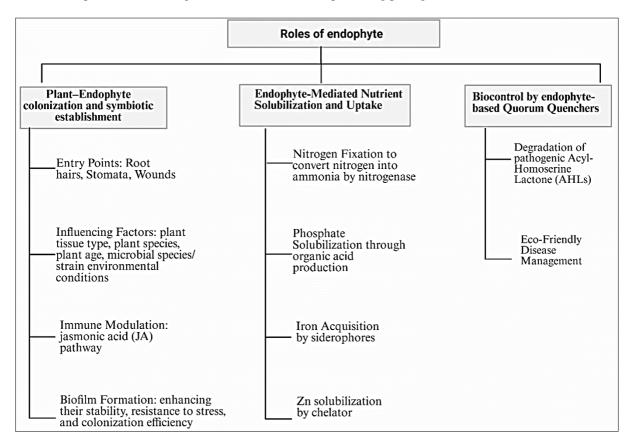


Fig 6: Advantages of Endophytic Microorganism through different Strategies (Adapted by Tariq et al. (2025)

Antibiosis and Secondary Metabolite Production: One of the most effective mechanisms of the plant for the production of bioactive secondary metabolites with antimicrobial compounds (Basit et al., 2021; Jain et al., 2019; Zaynab et al., 2018) [27, 89, 215]. These include antibiotics, cyclic lipopeptides, phenazines, siderophores, terpenoids, and volatile organic compounds VOCs, (Heredia-Bátiz et al., 2025; Navarro et al., 2019) [85, 131] and these compounds directly inhibit the growth or reproduction of pathogenic fungi, bacteria, or nematodes (Jha et al., 2022) [90] by disrupting cell wall integrity (Gogoi et al., 2024) [73], inhibiting essential enzymes, or generating reactive oxygen species (Oyebamiji et al., 2024) [173]. For example, Bacillus subtilis produces lipopeptides degrading substances such as surfactin, fengycin, and iturin, which form pores in fungal membranes, leading to cytoplasmic shrinkage finally death of the cell (Ongena and Jacques, 2008) [141]. Similarly, Pseudomonas fluorescens synthesizes phenazine-1-carboxylic acid and 2,4-diacetylphloroglucinol (DAPG), both of which are efficient antifungal agents (Raaijmakers et al., 2002; Njuguna, 2025) [162, 139]. Endophytic Streptomyces spp. produces a broad spectrum of antibiotics that exhibit antagonism activity toward various phytopathogenic species e.g., Fusarium, Ralstonia, and Pythium spp.

Competition for Nutrients and Ecological Niches: Endophytic microorganisms exhibit most powerful strategy,

suppress the pathogens through various way like niche exclusion (Wallis, 2021) [205], effectively living internal plant parts tissues (Omomowo and Babalola, 2019) [140] and competing with potential pathogens for space and limited nutrients (Liu *et al.*, 2017) [113] such as iron, nitrogen, and carbon sources (Blumenstein *et al.*, 2015) [37]. The ability of endophytes to colonize, stabilize the plant environment before pathogen (Bamisile *et al.*, 2021) [23] and confers a competitive edge, minimizing the chances of pathogen establishment (Gómez-Lama *et al.*, 2025; Khan *et al.*, 2025) [74, 100]. This mechanism is important in root systems where nutrient fluxes are high and microbial interactions are intense (Dutilloy *et al.*, 2022) [63].

According to Neilands, (1995) [136] iron is a critical micronutrient, and many endophytes produces siderophores high-affinity iron-chelating compounds to sequester iron from the rhizosphere to apoplast. By low iron availability, they create a hostile environment for iron-dependent pathogens.

Induced Systemic Resistance (ISR): ISR defines to the activation of plant defence responses triggered by beneficial microorganisms, including endophytes (Oukala *et al.*, 2021; Tonelli *et al.*, 2020; Rashid and Chung, 2017) [142, 193, 166] Unlike systemic acquired resistance (SAR), which is mediated by salicylic acid (SA)-dependent pathway (Sun *et al.*, 2021; Islam *et al.*, 2019) [189, 88] and triggered by

pathogenic infection (Kamle *et al.*, 2020; Roychowdhury *et al.*, 2024) $^{[92,\ 170]}$, ISR is often mediated by jasmonic acid (JA) and ethylene (ET) pathways (Yanti, 2019; Yang *et al.*, 2015) $^{[212,\ 211]}$ and does not necessary the presence of a pathogen (Cho *et al.*, 2008) $^{[49]}$.

Endophytes such as *Pseudomonas fluorescens* and *Trichoderma asperellum* have been observed to prime host plants for a quick and stronger defence response against pathogen attack (Aarthi *et al.*, 2025) ^[1]. This priming involves enhanced expression of pathogenesis-related (PR) genes or protein, production of reactive oxidative enzymes (e.g., peroxidases, polyphenol oxidases), and accumulation of secondary defence metabolites (Adeleke *et al.*, 2022) ^[3].

Production of Lytic Enzymes: Many endophytic microorganism's production of extracellular hydrolytic enzymes for capable of degrading pathogen cell walls (Sahoo *et al.*, 2025; Riseh *et al.*, 2024; Panicker and Sayyed, 2022) [174, 167, 146] and include chitinases, β-1,3-glucanases, proteases, cellulases, and lipases (Chandoliya *et al.*, 2023; Admassie *et al.*, 2022; Lopes *et al.*, 2021) [43, 6, 114]. This enzymatic degradation and ruptured the structural integrity of fungal and bacterial pathogens, leading to lysis finally death (Kandi *et al.*, 2022) [94]. For example, *Trichoderma* spp. has these enzymes in a coordinated manner to parasitize and degrade fungal hyphae a process called as mycoparasitism (Harman *et al.*, 2004) [82]. Similarly, *Streptomyces* and *Bacillus* spp. secretes proteases and chitinases enzymes that lysis pathogen cell walls in situ.

Productions of Volatile Organic Compounds (VOCs): Endophytes produce a wide range of low molecular weight compounds Volatile Organic Compounds (VOCs) with biocontrol activities (Ling *et al.*, 2024) [112] such as antibacterial, nematocidal and antifungal, activities (Karslı and Şahin, 2021; Roy *et al.*, 2019) [95, 169], *viz.*, 2,3-butanediol, hydrogen cyanide (HCN) and acetoin, have been identified and reported to suppress pathogen growth, development and spore germination, and other activities (Naz *et al.*, 2022) [132]. These compounds also have provided beneficial effects on plant physiology, enhancing plant health growth, biotic and abiotic stress tolerance.

Activation of Phytohormone Levels: Most of these beneficial endophytes are produce phytohormones (Younas et al., 2025; Cosoveanu et al., 2021; Xu et al., 2018) [213, 53, 210] such as indole-3-acetic acid (IAA), ethylene, gibberellins, cytokinin, and abscisic acid (ABA). These compounds not only promote plant growth, development and other activities but also mediate stress responses and fortify plant defences (Sharma et al., 2023) [184]. Additionally, hormone mimics or hormone-activating enzymes e.g., ACC deaminase help avoid abiotic stresses and stop pathogen-induced senescence, and enhancing plant vigour (Sadaf et al., 2016) [172].

Biofilm Formation and Colonization Resistance: Biofilm formation defence characteristics by endophytic bacteria creates a physical and biochemical barrier that suppresses pathogen colonization (Ajijah *et al.*, 2023; Pinski *et al.*, 2019) [11, 157]. Biofilms are complex extracellular polymeric substances (EPS) produced by endophytic microbial communities (Velmourougane *et al.*, 2017) [200] and promote adherence to plant tissues, protect against environmental

stresses, and provide nutrient exchange (Gogoi *et al.*, 2021) ^[72]. Endophytic strains of *Bacillus velezensis* and *Pseudomonas putida* have resulted strong biofilm-forming capabilities, contributing to persistent colonization and pathogen exclusion characters (Patil *et al.*, 2022) ^[150].

Quorum Quenching and Signal Interference: Some endophytes have quorum quenching activity, disrupting the communication systems of phytopathogens (Kusari et al., 2015) [107] that rely on quorum sensing (QS) to coordinate virulence (Venkatesh et al., 2019) [201], biofilm formation, and toxin secretion (Joo et al., 2021; Alagarasan et al., 2017) [91, 14]. Endophytes serve as N-acyl homoserine lactone (AHL) lactonases (Paul et al., 2023) [152] and acylases enzymes degrade signalling molecules like N-acyl homoserine lactones (AHLs) (Pellissier et al., 2021; Shastry and Rai, 2017) [153, 181], hindering the pathogens incapable of searching coordinated attacks (Anandan et al., 2019) [18]. These mechanisms enable endophytic microorganisms to function as more effective and multifaceted biocontrol agents (Abid Mehmood et al., 2025) [2]. Their internal stabilization, capacity for symbiotic relationship, and breadth of antagonism activity make them evenly equipped

A broad-level understanding of these interactions provided by multi-omics technologies, synthetic biology, and microbial ecology will be key to utilizing endophyte-based formulations for real-world agricultural.

for deployment in sustainable crop protection and

management strategies (Negi et al., 2024) [133].

Microbial Biotechnology Advancement through NGS

Recent era has supported advancements in Next-Generation Sequencing (NGS) technologies, revolutionizing of endophytic microbiomes and our understanding their functions in plant growth promotion and biocontrol. Culture-dependent methods, still valuable but have some limitations in failing to cover the full spectrum of microbial diversity within plant tissues. NGS technologies enables high-throughput, culture-independent profiling to identify of endophyte communities at an unprecedented resolution, allowing researchers to characterize and analyse microbial composition, function, and interaction, compatibility with high accuracy (Tariq *et al.*, 2025) [191].

Meta transcriptomics and Meta-Proteomics: These advanced omics technologies offer snapshots of endophytic gene expression (RNA-seq) and protein study under *in vivo* plant colonization surrounding. By detecting real-time interactions between endophytes and host plants, scientists and researchers can identify genes involved in immune priming, oxidative stress response mechanism, and signal response pathways. Meta transcriptomic studies offer in *Pseudomonas* and *Streptomyces* endophytic genera have linked expression of defence-mechanism proteins with phytopathogen suppression (Mabaso *et al.*, 2025) [117]

Metagenomics and Functional Annotation: Wholegenome shotgun metagenomics techniques surpass amplicon sequencing by providing dept study into the functional potentialities or capabilities of endophytes (Huang *et al.*, 2023) [86]. Researchers to reconstruct metabolic pathways, detect biosynthetic gene clusters their expression responsible for antibiotics, siderophores, and phytohormones, and study horizontal gene transfer that

transfer to biocontrol traits. For instance, metagenomic analysis has performed that certain *Bacillus* endophytes possess non-ribosomal peptide synthetase (NRPS) and polyketide synthase (PKS) gene clusters responsible for lipopeptide production (Patil *et al.*, 2025) [151].

Genome-Wide Association Studies (GWAS): Integrating NGS with GWAS enables researchers analyse to link between plant genetic traits and endophytic microbiome structure and function (Kumar *et al.*, 2025) [106]. These interactions provide information, how to boast plant immunity, metabolic signalling, and receptor expression affect microbial colonization, establishment, and biocontrol potential (Mani and Kushwaha, 2023) [122].

High-Throughput 16S rRNA and Internal Transcribed Spacer (ITS) Amplicon Sequencing: PCR-based sequencing of conserved genomic regions, such as 16S rRNA for endophytic bacteria (Zhang *et al.*, 2022) [216] and Internal Transcribed Spacer for fungi, facilitates taxonomic analysis into complex endophytic communities (Singh *et al.*, 2023; Błaszczyk *et al.*, 2021) [184, 36]. Through this technology has covered numerous unexplorable and unidentified endophytic microbes in cereals, leguminous and non-leguminous crop plants, such as wheat, maize, rice, and soybean. For their genome sequencing studies using Illumina Mi Seq and Hi Seq platforms have demonstrated (Dong *et al.*, 2023) [58] that each plant species hosts a unique reservoir of microbiome, affected by genotype, development stage, and surrounding (Bulgarelli *et al.*, 2012) [39].

Comparative Genomics and Pan-Genomics: These technologies permit for the comparison of various endophytic microbial strains from the same or different species or genera knowing conserved and unique genomic

sequences (Wang *et al.*, 2025; Mahmoud *et al.*, 2024; Peng *et al.*, 2024; Chen *et al.*, 2007) [214, 119, 156, 47]. Comparative genomic studies provide difference between pathogenic and non-pathogenic endophytic microbial genera, like *Fusarium* (Neik *et al.*, 2020) [135] or *Xanthomonas* strains (Passarelli-Araujo *et al.*, 2020) [148].

Single-Cell Genomics: This emerging technique enables the genomic characterization of individual unculturable and unidentified endophytes isolated from internal plant part tissues (Pradhan *et al.*, 2025; Shishodia *et al.*, 2025; Utturkar *et al.*, 2016) [159, 182, 197]. It holds promise for resolving intragenic variability, strain-specific traits, and efficient endophyte-plant specificity. It has been mainly useful in studying low-diversity microbial taxonomy that could be allows for host defence (Cole *et al.*, 2024) [50].

NGS-Enabled Bioprospecting: NGS technology offers to researchers can be identify microbial genomes for novel antimicrobial peptides, biosynthetic gene clusters, differential gene expression, and enzymes with potential agricultural applications. These studies are vital for developing next generation endophytic bioinoculants with multi-functional efficacy (Xu *et al.*, 2022) [209].

Moreover, the integration of transcriptomics, proteomics, and metabolomics are leveraged to study and offers a framework into real-time microbial metabolism and cross-kingdom signaling, and apply endophytes for improved crop yield and stress resilience.

Manna *et al.* (2025) ^[123] Strongly suggested that omics-based strategies to enhance agricultural productivity through endophyte research. A schematic representation of how genomics, transcriptomics, proteomics, and metabolomics are leveraged to study and apply endophytes for improved crop yield and stress resilience presented in Figure 7.

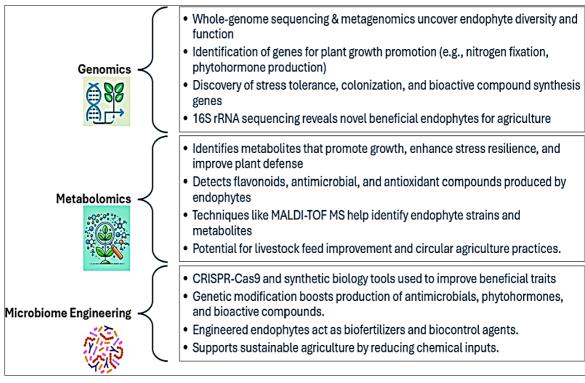


Fig 7: Microbial Biotechnology Advancement through NGS Adapted from Tariq et al. 2025)

Several types of case studies have been reported and explored how Next Generation Sequencing has accelerated the identification, research and application of endophytes in crop achievement, health management, and microbial biotechnology.

Patel *et al.* (2025) [149] reported an endophyte of *Brucella* sp. PM1, from pomegranate, and analysis through wholegenome NGS, technology revealing ~3000 plant growth-promoting trait genes mapped to both direct mechanisms - hormone secretion, nutrient uptake and indirect mechanisms - stress tolerance, colonization, and biocontrol. Their evidence showing capability for secondary metabolite production (ochrobactin), heavy-metal detoxification, and phytohormone biosynthesis pathways beyond IAA. There results concluded that *Brucella* genus is a good PGPR candidates and identifying multifaceted beneficial traits through whole genome sequencing (WGS).

Yu et al. (2025) [214] identified Klebsiella pneumoniae strain YMK25, from maize, and demonstrated multiple in-vitro Plant Growth Promoting traits viz., high IAA, N-fixation, inorganic/organic P solubilization, and siderophore production, sequenced and annotated through whole genome sequencing 5.12 Mb, 4,746 CDS, predicted PGP traits or related genes- nif/ure/gln, trp/ipdC, pho/phn, ent/fep, etc., and showed that YMK25 enhances maize seedling growth and soil nutrients in pot trials. Their results strongly suggested that YMK25 is a promising biofertilizer candidate.

Dong et al. (2023) [58] identified root endophytic microbial communities change across the growth stages of ratooning rice (cv. Jiafuzhan). NGS-based sequencing (16S + ITS) revealed clear shifts in root endophytic diversity across rice life stages. Their finding revealed that bacterial diversity highest at tillering stage, decreased at heading and ratooning and fungal diversity peaked at flowering and filling stages. Dominant bacterial endophytes Phyla: Proteobacteria, Actinobacteria, Firmicutes. Genera: Pseudomonas, Burkholderia, Bacillus and Dominant Fungal Endophytes Phyla: Ascomycota, Basidiomycota. Genera: Fusarium, Trichoderma, Aspergillus. Identify dominant endophytes (bacteria: genes for nitrogen fixation, organic matter decomposition, stress adaptation and fungi: functional guilds for saprotrophs, symbiosis, and plant defence) linked with plant growth, nutrient cycling, and stress tolerance. Their results demonstrated that NGS technology advances biotechnology by decoding endophytic diversity and functions, offering potential for biofertilizer and biocontrol applications in ratooning rice.

used to Next Generation Sequencing technology (Illumina high-throughput sequencing) for identification of the diversity of endophytic fungi in *Camellia reticulata* pedicels. In these studied revealed that *C. reticulata* pedicels host rich and diverse endophytic fungal communities,

dominated by Ascomycota. All the isolates have broad antagonism activities against to nectar yeasts. Alternaria alternata D23 results showed that highest antimicrobial potential and producing multiple antibiotics. NGS technology strongly revealed endophytic fungal diversity and reported 1,034 OTUs (4 phyla, 24 classes, 77 orders, 161 families, 267 genera), while 4,036 metabolites (including amino acids, fatty acids, vitamins, nucleotides) uncovered novel bioactive compounds. Findings suggest that endophytic fungi contribute to nectar stability, plant defence, and reproductive fitness in cross-pollinated plants. Endophytes can synthesize a larger number of secondary metabolites, including antibiotics, and plant growth promotion and disease suppression compound like siderophores, phytohormones, and VOCs etc. The redundancy and multifunctionality of these metabolites underscore the ecological importance of endophytes as keystone members of the plant microbiome. The endophytic lifestyle is spending within plant thought to be an evolutionary prospectus for microbial longevity, enabling colonization of a nutrient-rich and relatively protected environment. In return, plants benefit from enhanced nutrient acquisition, disease resistance, and stress tolerance. Some endophytes even exhibit vertical flow through seeds, indicating long-term host association and better co-evolution (Hardoim et al., 2015) [81].

The advancement of NGS technologies has overview expanded our capacity to find out the complexity of endophytic microbiomes and their ecological and functional roles in plant disease suppression. Future integration of multi-omics technologies data with AI-based analytics and systems. Researchers will further investigate, to optimization, and deployment of endophytic biocontrol agents in sustainable agriculture.

Overall, at functional characterization and taxonomic level diversity of endophytic microorganisms affects their ecology and evolutionary relationship. Identifying the complex interaction between host plant and endophytic microorganism at cellular, molecular, and ecological levels is important for their efficient characterization in biocontrol in sustainable agriculture.

Limitations and Challenges of Next Generation Sequencing Technology

Despite its power, Next Generation Sequencing technology faces numerous challenges and limitations like cost effective, incomplete reference databases, expertise for advanced bioinformatics studies, lack of standardized protocols causes variability across studies and difficulty in distinguishing true endophytic microbes from contaminants. Need of high-level functional predictions for experimental validation presented in Figure 8.

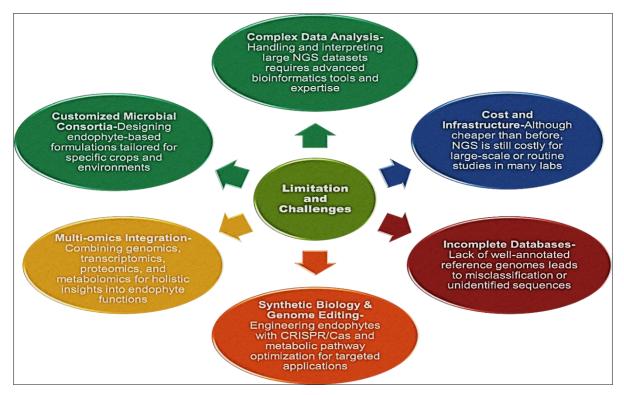


Fig 8: Limitations and Challenges of NGS Technology

Conclusion

Endophytic microorganisms represent powerful antagonistic activity against disease or phytopathogens through defense mechanism pathway and their modes of action, ability to colonize internal plant tissues, and potential to enhance plant growth make them ideal candidates for sustainable agriculture. Future discovery and development should focus on overcoming the current drawback and exploring their full potential for commercial applications in agricultural sector. Endophytic microorganisms hold great potential in agriculture. In advancement of Next-generation sequencing (NGS) technologies has revealing their hidden diversity, functional traits genes, and bio-physio-chemical metabolic pathways beyond conventional methods. These technologies accelerate the discovery of efficient endophytes as biofertilizers, novel metabolites and biocontrol agents, in conclusion Next-generation sequencing (NGS) technologies making endophytes valuable potential tools for sustainable biotechnology. Integrating Nextgeneration sequencing (NGS) technologies with multi-omics and synthetic biology will further unlock their applications sustainable agriculture for crop improvement, environmental sustainability.

Commercial Applications and Future Prospects

The commercialization of endophytic biocontrol agents is maintaining, with several products already available or under process. Advances in molecular biology, genomics, and bioinformatics and nanotechnology are facilitating the identification and characterization of promising potent endophytes. The future holds potential for engineering endophytes with enhanced biocontrol potent traits catch several diseases. Formulation of microbial consortia to facilitate broad level disease resistance, integration into integrated pest management programs. Further using to CRISPR-Cas based gene editing technology to investigate and manipulate plant-microbial interactions.

Conflict of Interest

The Authors declare no conflict of interest

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References

- 1. Aarthi R, Devanathan M, Harish S, Ganesan KN, Manivannan V. Biocontrol potential of endophytes against *Rhizoctonia bataticola*: antifungal activity, metabolite profiling, and plant defence mechanisms. Physiol Mol Plant Pathol. 2025;103:102734.
- 2. Abid Mehmood M, Rauf A, Abd-Elsalam KA, Ashfaq M, Kayani SB, Javeed S, *et al.* The multifaceted biocontrol mechanisms of endophytic fungi. In: Fungal endophytes volume II: applications in agroecosystems and plant protection. Singapore: Springer Nat.; 2025. p. 433-84.
- 3. Adeleke BS, Ayilara MS, Akinola SA, Babalola OO. Biocontrol mechanisms of endophytic fungi. Egypt J Biol Pest Control. 2022;32(1):46-54.
- 4. Adeleke BS, Fadiji AE, Ayilara MS, Igiehon ON, Nwachukwu BC, Babalola OO. Strategies to enhance the use of endophytes as bioinoculants in agriculture. Horticulturae. 2022;8(6):498-507.
- Adetunji CO, Anani OA, Islam S, Thangadurai D. Recent trends in the utilization of endophytic microorganisms and other bio-pesticidal technology for the management of agricultural pests, insects, and diseases. In: Organic farming for sustainable development. Apple Academic Press; 2022. p. 105-32.
- 6. Admassie M, Woldehawariat Y, Alemu T. *In vitro* evaluation of extracellular enzyme activity and its biocontrol efficacy of bacterial isolates from pepper plants for the management of *Phytophthora capsici*. Biomed Res Int. 2022;2022:6778352.

- 7. Afzal I, Shinwari ZK, Sikandar S, Shahzad S. Plant beneficial endophytic bacteria: mechanisms, diversity, host range and genetic determinants. Microbiol Res. 2019;221:36-49.
- 8. Aghdam SA, Brown AM. Deep learning approaches for natural product discovery from plant endophytic microbiomes. Environ Microbiol. 2021;16(1):6-16.
- 9. Ahlawat OP, Yadav D, Kashyap PL, Khippal A, Singh G. Wheat endophytes and their potential role in managing abiotic stress under changing climate. J Appl Microbiol. 2022;132(4):2501-20.
- 10. Ahmed A, He P, He Y, Singh BK, Wu Y, Munir S, *et al.* Biocontrol of plant pathogens in omics era with special focus on endophytic bacilli. Crit Rev Biotechnol. 2024;44(4):562-80.
- 11. Ajijah N, Fiodor A, Pandey AK, Rana A, Pranaw K. Plant growth-promoting bacteria (PGPB) with biofilm-forming ability: a multifaceted agent for sustainable agriculture. Diversity. 2023;15(1):112-25.
- 12. Akram S, Ahmed A, He P, Liu Y, Wu Y, Munir S, *et al.* Uniting the role of endophytic fungi against plant pathogens and their interaction. J Fungi. 2023;9(1):72-84.
- 13. Akram S, Ahmed A, He P, Liu Y, Wu Y, Munir S, *et al.* Uniting the role of endophytic fungi against plant pathogens and their interaction. J Fungi. 2023;9(1):72-84
- 14. Alagarasan G, Aswathy KS, Madhaiyan M. Shoot the message, not the messenger: combating pathogenic virulence in plants by inhibiting quorum sensing mediated signaling molecules. Front Plant Sci. 2017;8:556-65.
- 15. Al-Ani LK. The importance of endophytic fungi from the medicinal plant: diversity, natural bioactive compounds, and control of plant pathogens. In: Medically important plant biomes: source of secondary metabolites. Singapore: Springer; 2019. p. 189-238.
- 16. Ali M, Ali Q, Sohail MA, Ashraf MF, Saleem MH, Hussain S, *et al.* Diversity and taxonomic distribution of endophytic bacterial community in the rice plant and its prospective. Int J Mol Sci. 2021;22(18):10165-77.
- 17. Anand U, Pal T, Yadav N, Singh VK, Tripathi V, Choudhary KK, *et al.* Current scenario and future prospects of endophytic microbes: promising candidates for abiotic and biotic stress management for agricultural and environmental sustainability. Microb Ecol. 2023;86(3):1455-86.
- 18. Anandan K, Vittal RR. Quorum quenching activity of AiiA lactonase KMMI17 from endophytic *Bacillus thuringiensis* KMCL07 on AHL-mediated pathogenic phenotype in *Pseudomonas aeruginosa*. Microb Pathog. 2019;132:230-42.
- 19. Andryukov BG, Karpenko AA, Lyapun IN. Learning from nature: bacterial spores as a target for current technologies in medicine. Sovrem Tekhnol Med. 2020;12(3):105-22.
- 20. Arnold AE, Lutzoni F. Diversity and host range of foliar fungal endophytes: are tropical leaves biodiversity hotspots? Ecology. 2007;88(3):541-9.
- 21. Arnold AE, Maynard Z, Gilbert GS. Fungal endophytes in dicotyledonous neotropical trees: patterns of abundance and diversity. Mycol Res. 2001;105(12):1502-17.

- 22. Bamisile BS, Akutse KS, Siddiqui JA, Xu Y. Model application of entomopathogenic fungi as alternatives to chemical pesticides: prospects, challenges, and insights for next-generation sustainable agriculture. Front Plant Sci. 2021;12:741804.
- 23. Bamisile BS, Siddiqui JA, Akutse KS, Ramos Aguila LC, Xu Y. General limitations to endophytic entomopathogenic fungi use as plant growth promoters, pests and pathogens biocontrol agents. Plants. 2021;10(10):2119-32.
- 24. Bard NW, Cronk QC, Davies TJ. Fungal endophytes can modulate plant invasion. Biol Rev. 2024;99(5):1652-71.
- 25. Baron NC, Rigobelo EC. Endophytic fungi: a tool for plant growth promotion and sustainable agriculture. Mycology. 2022;13(1):39-55.
- 26. Basit A, Shah ST, Ullah I, Mohamed HI. Microbial bioactive compounds produced by endophytes (bacteria and fungi) and their uses in plant health. In: Plant growth-promoting microbes for sustainable biotic and abiotic stress management. Springer Int Publ.; 2021. p. 285-318.
- 27. Basit A, Shah ST, Ullah I, Mohamed HI. Microbial bioactive compounds produced by endophytes (bacteria and fungi) and their uses in plant health. In: Plant growth-promoting microbes for sustainable biotic and abiotic stress management. Cham: Springer Int Publ.; 2021. p. 285-318.
- 28. Béchamp A. Du rôle de la craie dans les fermentations butyrique et lactique, et des organismes actuellement vivants qu'elle contient. CR Hebd Seances Acad Sci. 1866;63:451-6.
- 29. Berg G, Erlacher A, Smalla K, Krause R. Vegetable microbiomes: is there a connection among opportunistic infections, human health and our 'gut feeling'? Microb Biotechnol. 2014;7(6):487-95.
- 30. Beskrovnaya P, Sexton DL, Golmohammadzadeh M, Hashimi A, Tocheva EI. Structural, metabolic and evolutionary comparison of bacterial endospore and exospore formation. Front Microbiol. 2021;12:630573.
- 31. Bhagya M, Nagaraju K, Praveen Biradar B, Santhosh G, Gundappagol R. Isolation and characterization of endophytic bacteria from nodule, root and seeds of greengram (*Vigna radiata* L.). Indian J Pure Appl Biosci. 2019;7:319-28.
- 32. Bhardwaj M, Kailoo S, Khan RT, Khan SS, Rasool S. Harnessing fungal endophytes for natural management: a biocontrol perspective. Front Microbiol. 2023;14:1280258.
- 33. Bharti C, Mishra P, Verma P, Bhattacharya A, Arora P, Arora NK. Microorganisms for sustainable agriculture and food security. In: Environmental sustainability and biotechnology: opportunities and challenges. Springer Nat.; 2025. p. 53-94.
- 34. Bielecka M, Pencakowski B, Nicoletti R. Using nextgeneration sequencing technology to explore genetic pathways in endophytic fungi in the syntheses of plant bioactive metabolites. Agriculture. 2022;12(2):187-201.
- 35. Bintarti AF, Sulesky-Grieb A, Stopnisek N, Shade A. Endophytic microbiome variation among single plant seeds. Phytobiomes J. 2022;6(1):45-55.
- 36. Błaszczyk L, Salamon S, Mikołajczak K. Fungi inhabiting the wheat endosphere. Pathogens. 2021;10(10):1288-96.

- 37. Blumenstein K, Albrectsen BR, Martín JA, Hultberg M, Sieber TN, Helander M, *et al.* Nutritional niche overlap potentiates the use of endophytes in biocontrol of a tree disease. Biocontrol. 2015;60(5):655-67.
- 38. Bosamia TC, Barbadikar KM, Modi A. Genomic insights of plant-endophyte interaction: prospective and impact on plant fitness. In: Microbial endophytes. Woodhead Publishing; 2020. p. 227-49.
- 39. Bulgarelli D, Rott M, Schlaeppi K, Ver Loren van Themaat E, Ahmadinejad N, Assenza F, *et al.* Revealing structure and assembly cues for *Arabidopsis* root-inhabiting bacterial microbiota. Nature. 2012;488(7409):91-5.
- 40. Carroll GC. The biology of endophytism in plants with particular reference to woody perennials. In: Microbiology of the phyllosphere. London: Academic Press; 1986. p. 203-22.
- 41. Cavalcante VA, Dobereiner J. A new acid-tolerant nitrogen-fixing bacterium associated with sugarcane. Plant Soil. 1988;108(1):23-31.
- 42. Chagas FO, de Cassia Pessotti R, Caraballo-Rodríguez AM, Pupo MT. Chemical signaling involved in plantmicrobe interactions. Chem Soc Rev. 2018;47(5):1652-704
- 43. Chandoliya R, Sharma S, Sharma V. Role of microbial oxidative enzymes in biological control of plant pathogens. In: Microbial oxidative enzymes: biotechnological applications. 2023. p. 129-50.
- 44. Chaudhary P, Agri U, Chaudhary A, Kumar A, Kumar G. Endophytes and their potential in biotic stress management and crop production. Front Microbiol. 2022;13:933017.
- 45. Chaudhary P, Agri U, Chaudhary A, Kumar A, Kumar G. Endophytes and their potential in biotic stress management and crop production. Front Microbiol. 2022;13:933017.
- 46. Chaurasia A, Meena BR, Tripathi AN, Pandey KK, Rai AB, Singh B. Actinomycetes: an unexplored microorganism for plant growth promotion and biocontrol in vegetable crops. World J Microbiol Biotechnol. 2018;34(1):6-12.
- 47. Chen XH, Koumoutsi A, Scholz R, Eisenreich A, Schneider K, Heinemeyer I, *et al.* Comparative analysis of the complete genome sequence of the plant growth-promoting bacterium *Bacillus amyloliquefaciens* FZB42. Nat Biotechnol. 2007;25(9):1007-14.
- 48. Chen XL, Sun MC, Chong SL, Si JP, Wu LS. Transcriptomic and metabolomic approaches deepen our knowledge of plant-endophyte interactions. Front Plant Sci. 2022;12:700200.
- 49. Cho SM, Kang BR, Han SH, Anderson AJ, Park JY, Lee YH, *et al.* 2R,3R-butanediol, a bacterial volatile produced by *Pseudomonas chlororaphis* O6, is involved in induction of systemic tolerance to drought in *Arabidopsis thaliana*. Mol Plant Microbe Interact. 2008;21(8):1067-75.
- 50. Cole J, Raguideau S, Abbaszadeh-Dahaji P, Hilton S, Muscatt G, Mushinski RM, *et al.* Comparative genomic analysis of a metagenome-assembled genome reveals distinctive symbiotic traits in a *Mucoromycotina* fine root endophyte arbuscular mycorrhizal fungus. bioRxiv. 2024. Preprint. doi:10.1101/2024.11.
- 51. Compant S, Duffy B, Nowak J, Clément C, Barka EA. Use of plant growth-promoting bacteria for biocontrol

- of plant diseases: principles, mechanisms of action, and future prospects. Appl Environ Microbiol. 2005;71(9):4951-9.
- 52. Compant S, Vacher C. Sources, niches and routes of colonization by beneficial bacterial endophytes. In: Endophyte biotechnology: potential for agriculture and pharmacology. Wallingford: CABI; 2019. p. 32-41.
- 53. Cosoveanu A, Chowdhary K, Cabrera R, Sharma S. Role of phytohormone-producing fungal endophytes in plant-microbial interactions under stress. In: Endophytes: potential source of compounds of commercial and therapeutic applications. 2021. p. 195-223.
- 54. Das D, Sharma PL, Paul P, Baruah NR, Choudhury J, Begum T, *et al.* Harnessing endophytes: innovative strategies for sustainable agricultural practices. Discov Bacteriol. 2025;2(1):1-12.
- 55. de Bary A. Morphologie und physiologie der pilze, flechten und myxomyceten. Leipzig: Engelmann; 1866. p. 1-200.
- 56. Deepa N, Chauhan S, Singh A. Unraveling the functional characteristics of endophytic bacterial diversity for plant growth promotion and enhanced secondary metabolite production in *Pelargonium graveolens*. Microbiol Res. 2024;283:127673.
- 57. Dittami SM, Arboleda E, Auguet JC, Bigalke A, Briand E, Cárdenas P, *et al.* A community perspective on the concept of marine holobionts: current status, challenges, and future directions. PeerJ. 2021;9:e10911.
- 58. Dong M, Shi L, Xie Z, Lian L, Zhang J, Jiang Z, *et al.* Shifts in the diversity of root endophytic microorganisms across the life cycle of the ratooning rice Jiafuzhan. Front Microbiol. 2023;14:1161263.
- 59. Dos Reis JB, do Vale HM, Lorenzi AS. Insights into taxonomic diversity and bioprospecting potential of Cerrado endophytic fungi: a review exploring a unique Brazilian biome and methodological limitations. World J Microbiol Biotechnol. 2022;38(11):202-15.
- Doty SL, Joubert PM, Firrincieli A, Sher AW, Tournay R, Kill C, et al. Potential biocontrol activities of *Populus* endophytes against several plant pathogens using different inhibitory mechanisms. Pathogens. 2022;12(1):13-25.
- 61. Dubey A, Malla MA, Kumar A, Dayanandan S, Khan ML. Plant endophytes: unveiling hidden agenda for bioprospecting toward sustainable agriculture. Crit Rev Biotechnol. 2020;40(8):1210-31.
- 62. Durand A, Leglize P, Benizri E. Are endophytes essential partners for plants and what are the prospects for metal phytoremediation? Plant Soil. 2021;460(1):1-30
- 63. Dutilloy E, Oni FE, Esmaeel Q, Clément C, Barka EA. Plant beneficial bacteria as bioprotectants against wheat and barley diseases. J Fungi. 2022;8(6):632-43.
- 64. Eid AM, Fouda A, Abdel-Rahman MA, Salem SS, Elsaied A, Oelmüller R, *et al.* Harnessing bacterial endophytes for promotion of plant growth and biotechnological applications: an overview. Plants. 2021;10(5):935-50.
- 65. El-Metwally MM, Mekawey AA, El-Halmouch Y, Naga NG. Symbiotic relationships with fungi: from mutualism to parasitism. In: Plant mycobiome: diversity, interactions and uses. Cham: Springer Int Publ.; 2023. p. 375-413.

- 66. El-Saadony MT, Saad AM, Soliman SM, Salem HM, Ahmed AI, Mahmood M, *et al*. Plant growth-promoting microorganisms as biocontrol agents of plant diseases: mechanisms, challenges and future perspectives. Front Plant Sci. 2022;13:923880.
- 67. Emitaro WO, Kawaka F, Musyimi DM, Adienge A. Diversity of endophytic bacteria isolated from leguminous agroforestry trees in western Kenya. AMB Express. 2024;14(1):18-28.
- 68. Fite T, Kebede E, Tefera T, Bekeko Z. Endophytic fungi: versatile partners for pest biocontrol, growth promotion, and climate change resilience in plants. Front Sustain Food Syst. 2023;7:1322861.
- 69. Fontana DC, de Paula S, Torres AG, de Souza VH, Pascholati SF, Schmidt D, *et al.* Endophytic fungi: biological control and induced resistance to phytopathogens and abiotic stresses. Pathogens. 2021;10(5):570-83.
- 70. Gagné-Bourque F. *Bacillus subtilis* B26, a plant growth promoter and drought protector of grasses [dissertation]. Montreal: McGill University; 2015.
- 71. Galippe V. Note sur la présence de micro-organismes dans les tissus végétaux. CR Seances Soc Biol Fil. 1887;39:410-6.
- 72. Gogoi A, Poudel M, Sahu J, Baruah G. Role of microbial biofilms in agriculture: perspectives on plant and soil health. In: Microbial polymers: applications and ecological perspectives. Singapore: Springer; 2021. p. 251-88.
- 73. Gogoi K, Gogoi H, Borgohain M, Saikia R, Chikkaputtaiah C, Hiremath S, *et al*. The molecular dynamics between reactive oxygen species, reactive nitrogen species and phytohormones in plant's response to biotic stress. Plant Cell Rep. 2024;43(11):263-78.
- 74. Gómez-Lama Cabanás C, Mercado-Blanco J. Groundbreaking technologies and the biocontrol of fungal vascular plant pathogens. J Fungi. 2025;11(1):77-90.
- 75. Grabka R, d'Entremont TW, Adams SJ, Walker AK, Tanney JB, Abbasi PA, *et al.* Fungal endophytes and their role in agricultural plant protection against pests and pathogens. Plants. 2022;11(3):384-96.
- 76. Grzyb T, Szulc J. Deciphering molecular mechanisms and diversity of plant holobiont bacteria: microhabitats, community ecology, and nutrient acquisition. Int J Mol Sci. 2024;25(24):13601-18.
- 77. Gulyamova T, Madan A. Modes of action of biocontrol agents from rhizosphere to phyllosphere. In: Biocontrol agents for sustainable agriculture: diversity, mechanisms and applications. Singapore: Springer Nat.; 2025. p. 117-32.
- 78. Hajji-Hedfi L, Wannassi T, El-Maradny YA. Fungal endophytes: insight into evolution, classification, and ecological functions in plants. Microb Biosyst. 2025;10(1):51-67.
- 79. Hakansson AP, Orihuela CJ, Bogaert D. Bacterial-host interactions: physiology and pathophysiology of respiratory infection. Physiol Rev. 2018;98(2):781-811.
- 80. Hallmann J, Quadt-Hallmann A, Mahaffee WF, Kloepper JW. Bacterial endophytes in agricultural crops. Can J Microbiol. 1997;43(10):895-914.
- 81. Hardoim PR, Van Overbeek LS, Berg G, Pirttilä AM, Compant S, Campisano A, *et al.* The hidden world within plants: ecological and evolutionary

- considerations for defining functioning of microbial endophytes. Microbiol Mol Biol Rev. 2015;79(3):293-320.
- 82. Harman GE, Howell CR, Viterbo A, Chet I, Lorito M. *Trichoderma* species opportunistic, avirulent plant symbionts. Nat Rev Microbiol. 2004;2(1):43-56.
- 83. Harrison JG, Griffin EA. The diversity and distribution of endophytes across biomes, plant phylogeny and host tissues: how far have we come and where do we go from here? Environ Microbiol. 2020;22(6):2107-23.
- 84. Harrison JG, Griffin EA. The diversity and distribution of endophytes across biomes, plant phylogeny and host tissues: how far have we come and where do we go from here? Environ Microbiol. 2020;22(6):2107-23.
- 85. Heredia-Bátiz JM, Manjarrez-Quintero JP, Valdez-Baro O, Rivera-Salas MM, Bayardo-Rosales H, Jiménez-Ortega LA, *et al.* Role of bioactive compounds in plant disease management. In: Biotechnological intervention in production of bioactive compounds: biosynthesis, characterization and applications. Cham: Springer Nature; 2025. p. 141-62.
- 86. Huang D, Ren J, Chen X, Akhtar K, Liang Q, Ye C, *et al.* Whole-genome assembly of A02 bacteria involved in nitrogen fixation within cassava leaves. Plant Physiol. 2023;193(2):1479-90.
- 87. Huang R, Meng Q, Xun L, Wu X, Yue D, Zhao W, *et al.* Antagonistic effects of endophytic fungi from *Camellia reticulata* pedicels on yeasts: implications for antimicrobial mechanism of nectar. Front Plant Sci. 2024;15:1494855.
- 88. Islam MN, Ali MS, Choi SJ, Park YI, Baek KH. Salicylic acid-producing endophytic bacteria increase nicotine accumulation and resistance against wildfire disease in tobacco plants. Microorganisms. 2019;8(1):31-45.
- 89. Jain C, Khatana S, Vijayvergia R. Bioactivity of secondary metabolites of various plants: a review. Int J Pharm Sci Res. 2019;10(2):494-504.
- 90. Jha Y, Mohamed HI. Plant secondary metabolites as a tool to investigate biotic stress tolerance in plants: a review. Gesunde Pflanzen. 2022;74(4):771-90.
- 91. Joo HS, Deyrup ST, Shim SH. Endophyte-produced antimicrobials: a review of potential lead compounds with a focus on quorum-sensing disruptors. Phytochem Rev. 2021;20(3):543-68.
- 92. Kamle M, Borah R, Bora H, Jaiswal AK, Singh RK, Kumar P. Systemic acquired resistance and induced systemic resistance: role and mechanism of action against phytopathogens. In: Fungal biotechnology and bioengineering. Cham: Springer Int Publ; 2020. p. 457-70.
- 93. Kandel SL. Salicaceae endophytes: growth promotion potential in rice (*Oryza sativa* L.) and maize (*Zea mays* L.) and bio-control of plant pathogen [dissertation]. Pullman: Washington State University; 2016. p. 1-180.
- 94. Kandi SR, Kunduru S, Guduru KK, Naik PK, Naga RY, Akila CS, *et al.* Endophytic bacteria *Bacillus amyloliquefaciens* (Pj2) imparted growth promotion and enhanced drought stress tolerance of peanut. Curr Trends Biotechnol Pharm. 2022;16(2):165-82.
- 95. Karslı A, Şahin YS. The role of fungal volatile organic compounds in biological control. Turk Biyol Muj Derg. 2021;12(1):79-92.

- 96. Kaul S, Sharma T, Dhar MK. Omics tools for better understanding the plant-endophyte interactions. Front Plant Sci. 2016;7:955-65.
- 97. Kaur R, Kaur C, Kaur G, Kaur J, Rath SK, Dwibedi V. From microscopy to omics: a comprehensive review of tools and techniques in studying endophytic adaptation under abiotic and biotic stress. J Plant Growth Regul. 2024;43(5):1-9.
- 98. Kayode AJ, Igwaran A, Banji-Onisile F, Akwu NA, Unuofin JO, Osunla AC, *et al.* Unveiling the hidden allies in the fight against antimicrobial resistance: medicinal plant endophytes. Bacteria. 2025;4(2):26-40.
- 99. Khan D, Shaw R, Kabiraj A, Paul A, Bandopadhyay R. Microbial inheritance through seed: a clouded area needs to be enlightened. Arch Microbiol. 2025;207(1):23-34.
- 100.Khan D, Shaw R, Kabiraj A, Paul A, Bandopadhyay R. Microbial inheritance through seed: a clouded area needs to be enlightened. Arch Microbiol. 2025;207(1):23-34.
- 101.Khare E, Mishra J, Arora NK. Multifaceted interactions between endophytes and plant: developments and prospects. Front Microbiol. 2018;9:2732.
- 102. Kobayashi DY, Palumbo JD. Bacterial endophytes and their effects on plants and uses in agriculture. In: Bacon CW, White JF, editors. Microbial endophytes. Boca Raton: CRC Press; 2000. p. 213-50.
- 103.Kumar A, Radhakrishnan EK, editors. Microbial endophytes: functional biology and applications. Cambridge: Woodhead Publishing; 2020.
- 104.Kumar J, Murali-Baskaran RK, Jain SK, Sivalingam PN, Mallikarjuna J, Kumar V, *et al.* Emerging and reemerging biotic stresses of agricultural crops in India and novel tools for their better management. Curr Sci. 2021;121(1):26-36.
- 105.Kumar M, Saxena R, Tomar RS. Endophytic microorganisms: promising candidate as biofertilizer.
 In: Meena VS, *et al.*, editors. Microorganisms for Green Revolution. Vol. 1. Singapore: Springer; 2017. p. 77-85
- 106. Kumar N, Sindhu S, Gawaria B, Ansari A, Dhariwal A, Sharma P, *et al.* GWAS approaches in medicinal plants for enhanced secondary metabolite production. In: Kumar V, *et al.*, editors. Biotechnology, multiple omics, and precision breeding in medicinal plants. Boca Raton: CRC Press; 2025. p. 119-34.
- 107. Kusari P, Kusari S, Spiteller M, Kayser O. Implications of endophyte-plant crosstalk in light of quorum responses for plant biotechnology. Appl Microbiol Biotechnol. 2015;99(13):5383-90.
- 108. Kuźniar A, Kruczyńska A, Włodarczyk K, Vangronsveld J, Wolińska A. Endophytes as permanent or temporal inhabitants of different ecological niches in sustainable agriculture. Appl Sci. 2025;15(3):1253.
- 109.Latz MA, Kerrn MH, Sørensen H, Collinge DB, Jensen B, Brown JK, *et al.* Succession of the fungal endophytic microbiome of wheat is dependent on tissue-specific interactions between host genotype and environment. Sci Total Environ. 2021;759:143804.
- 110.Lazar A, Mushinski RM, Bending GD. Landscape scale ecology of *Tetracladium* spp. fungal root endophytes. Environ Microb. 2022;17(1):40.
- 111.L'Hoir M, Duponnois R. Combining the seed endophytic bacteria and the back to the future

- approaches for plant holobiont breeding. Front Agron. 2021;3:724450.
- 112.Ling L, Feng L, Li Y, Yue R, Wang Y, Zhou Y. Endophytic fungi volatile organic compounds as crucial biocontrol agents used for controlling fruit and vegetable postharvest diseases. J Fungi. 2024;10(5):332.
- 113.Liu H, Carvalhais LC, Crawford M, Singh E, Dennis PG, Pieterse CM, *et al.* Inner plant values: diversity, colonization and benefits from endophytic bacteria. Front Microbiol. 2017;8:2552.
- 114.Lopes FC, Martinelli AH, John EB, Ligabue-Braun R. Microbial hydrolytic enzymes: powerful weapons against insect pests. In: Saini RK, *et al.*, editors. Microbes for sustainable insect pest management: hydrolytic enzyme & secondary metabolite. Vol. 2. Cham: Springer; 2021. p. 1-31.
- 115.Lu H, Wei T, Lou H, Shu X, Chen Q. A critical review on communication mechanism within plant-endophytic fungi interactions to cope with biotic and abiotic stresses. J Fungi. 2021;7(9):719.
- 116.Lyu D, Zajonc J, Pagé A, Tanney CA, Shah A, Monjezi N, *et al.* Plant holobiont theory: the phytomicrobiome plays a central role in evolution and success. Microorganisms. 2021;9(4):675.
- 117. Mabaso J, Henema N, Niekerk LA, Holman DE, Keyster M, Klein A, *et al.* Enhancing *Fusarium oxysporum* tolerance in *Phaseolus vulgaris*: isolation and characterization of bacterial endophytes. Physiol Mol Plant Pathol. 2025;138:102688.
- 118.Maestro-Gaitán I, Redondo-Nieto M, González-Bodí S, Rodríguez-Casillas L, Matías J, Bolaños L, *et al.* Insights into quinoa endophytes: core bacterial communities reveal high stability to water stress and genotypic variation. Environ Microb. 2025;20(1):16.
- 119.Mahmoud FM, Pritsch K, Siani R, Benning S, Radl V, Kublik S, *et al.* Comparative genomic analysis of strain *Priestia megaterium* B1 reveals conserved potential for adaptation to endophytism and plant growth promotion. Microbiol Spectr. 2024;12(8):e00422-24.
- 120.Mamarasulov B, Davranov K, Gulyamova T, Madan A, Umruzaqov A, Jabborova D. Modes of action of biocontrol agents from rhizosphere to phyllosphere signaling. In: Guleria S, *et al.*, editors. Bio-control agents for sustainable agriculture: diversity, mechanisms and applications. Singapore: Springer; 2025. p. 117-39.
- 121.Mametja NM, Ramadwa TE, Managa M, Masebe TM. Recent advances and developments in bacterial endophyte identification and application: a 20-year landscape review. Plants. 2025;14(16):2506.
- 122.Mani A, Kushwaha S, editors. Genomics of plant-pathogen interaction and the stress response. Boca Raton: CRC Press; 2023.
- 123.Manna D, Laishram R, Arpitha SR, Rai M, Dahuja A. Omics for thriving plant microbiomes: growing food security for a sustainable future. In: Singh BP, *et al.*, editors. Microbial metabolomics: recent developments, challenges and future opportunities. Singapore: Springer; 2025. p. 315-46.
- 124.Mattoo AJ, Nonzom S. Endophytic fungi: understanding complex cross-talks. Symbiosis. 2021;83(3):237-64.

- 125.Mengistu AA. Endophytes: colonization, behaviour, and their role in defense mechanism. Int J Microbiol. 2020;2020:6927219.
- 126.Mina D, Pereira JA, Lino-Neto T, Baptista P. Impact of plant genotype and plant habitat in shaping bacterial patho-biome: a comparative study in olive tree. Sci Rep. 2020;10(1):3475.
- 127.Mishra S, Bhattacharjee A, Sharma S. An ecological insight into the multifaceted world of plant-endophyte association. Crit Rev Plant Sci. 2021;40(2):127-46.
- 128.Mishra S, Bhattacharjee A, Sharma S. An ecological insight into the multifaceted world of plant-endophyte association. Crit Rev Plant Sci. 2021;40(2):127-46.
- 129. Nair DN, Padmavathy SJ. Impact of endophytic microorganisms on plants, environment and humans. Sci World J. 2014;2014:250693.
- 130.Narayanasamy S, Rajkumar M, Muthuramalingam G, Sudalaimani C, Uthandi S. Microbial metabolites: a potential weapon against phytopathogens. In: Kumar V, *et al.*, editors. Microbial biocontrol: molecular perspective in plant disease management. Singapore: Springer; 2023. p. 1-28.
- 131. Navarro MO, Piva AC, Simionato AS, Spago FR, Modolon F, Emiliano J, *et al.* Bioactive compounds produced by biocontrol agents driving plant health. In: Yadav AN, *et al.*, editors. Microbiome in plant health and disease: challenges and opportunities. Singapore: Springer; 2019. p. 337-74.
- 132.Naz R, Khushhal S, Asif T, Mubeen S, Saranraj P, Sayyed RZ. Inhibition of bacterial and fungal phytopathogens through volatile organic compounds produced by *Pseudomonas* sp. In: Maheshwari DK, *et al.*, editors. Secondary metabolites and volatiles of PGPR in plant-growth promotion. Singapore: Springer; 2022. p. 95-118.
- 133.Negi R, Sharma B, Parastesh F, Kaur S, Khan SS, Kour D, *et al.* Microbial consortia mediated regulation of plant defense: a promising tool for sustaining crops protection. Physiol Mol Plant Pathol. 2024;102393.
- 134.Negi R, Yadav AN. Plant-microbe interaction for environmental and agricultural sustainability: conclusion and future challenges. In: Yadav AN, *et al.*, editors. Plant-microbe interactions for environmental and agricultural sustainability. Singapore: Springer; 2025. p. 695-716.
- 135.Neik TX, Amas J, Barbetti M, Edwards D, Batley J. Understanding host-pathogen interactions in *Brassica napus* in the omics era. Plants. 2020;9(10):1336.
- 136.Neilands JB. Siderophores: structure and function of microbial iron transport compound. J Biol Chem. 1995;270(45):26723-6.
- 137.Ngalimat MS, Mohd Hata E, Zulperi D, Ismail SI, Ismail MR, Mohd Zainudin NA, *et al.* Plant growth-promoting bacteria as an emerging tool to manage bacterial rice pathogens. Microorganisms. 2021;9(4):682.
- 138.Nimbulkar P, Gupta G, Virkhare U, Althubiani AS, Dutta A, Kher D. Bacterial endophytes and their secondary metabolites: mechanisms of biosynthesis and applications in sustainable agriculture. J Umm Al-Qura Univ Appl Sci. 2025;1-3.
- 139. Njuguna DW. Isolation, characterization & identification of cultivable field pea seed bacteria in

- South Dakota [master's thesis]. Brookings: South Dakota State University; 2025.
- 140.Omomowo OI, Babalola OO. Bacterial and fungal endophytes: tiny giants with immense beneficial potential for plant growth and sustainable agricultural productivity. Microorganisms. 2019;7(11):481.
- 141.Ongena M, Jacques P. *Bacillus* lipopeptides: versatile weapons for plant disease biocontrol. Trends Microbiol. 2008;16(3):115-125.
- 142. Oukala N, Aissat K, Pastor V. Bacterial endophytes: the hidden actor in plant immune responses against biotic stress. Plants. 2021;10(5):1012-1022.
- 143.Oyebamiji YO, Yusri NS, Akinola SA, Daba TM, Sinumvayo JP, Adeyemo RO, *et al.* Response of ROS in host plant assaulted by opportunistic fungi and nematodes. In: Opportunistic fungi, nematode and plant interactions: interplay and mechanisms. Cham: Springer Nat.; 2024. p. 69-89.
- 144.Pandey P, Tripathi A, Dwivedi S, Lal K, Jhang T. Deciphering the mechanisms, hormonal signaling, and potential applications of endophytic microbes to mediate stress tolerance in medicinal plants. Front Plant Sci. 2023;14:1250020.
- 145.Pandit MA, Kumar J, Gulati S, Bhandari N, Mehta P, Katyal R, *et al.* Major biological control strategies for plant pathogens. Pathog. 2022;11(2):273-285.
- 146.Panicker S, Sayyed RZ. Hydrolytic enzymes from PGPR against plant fungal pathogens. In: Antifungal metabolites of rhizobacteria for sustainable agriculture. Cham: Springer Int. Publishing; 2022. p. 211-238.
- 147. Panwar JS, Samuel J, Kumar A, editors. Volatiles and metabolites of microbes. Cambridge (MA): Academic Press; 2021.
- 148.Passarelli-Araujo H, Jacobs SH, Franco GR, Venancio TM. Phylogenetic and population structure analyses uncover pervasive misclassification and help assessing the biosafety of *Pseudomonas putida* for biotechnological applications. bioRxiv. 2020;2020-12.
- 149.Patel P, Patel F, Joshi C, Joshi M. Whole genome analysis of endophytic strain PM1 reveals promising plant growth-promoting mechanisms in pomegranate. J Genet Eng Biotechnol. 2025;23(2):100486.
- 150.Patil A, Gondi R, Rale V, Saroj SD. Microbial biofilms in plant disease management. In: Biocontrol mechanisms of endophytic microorganisms. Cambridge (MA): Academic Press; 2022. p. 239-259.
- 151.Patil BL, Gopalkrishna AM, GM SK. Molecular characterization of an endophytic strain of *Bacillus subtilis* with plant growth-promoting properties from a wild relative of papaya. J Appl Microbiol. 2025;136(1):10-22.
- 152.Paul E, Mathur P, Sharma C, Chaturvedi P. Quorum quenching strategies of endophytic bacteria: role in plant protection. In: Plant-microbe interaction: recent advances in molecular and biochemical approaches. Cambridge (MA): Academic Press; 2023. p. 149-166.
- 153.Pellissier L, Leoni S, Marcourt L, Ferreira Queiroz E, Lecoultre N, Quiros-Guerrero LM, *et al.* Characterization of *Pseudomonas aeruginosa* quorum sensing inhibitors from the endophyte *Lasiodiplodia venezuelensis* and evaluation of their antivirulence effects by metabolomics. Microorganisms. 2021;9(9):1807-1824.

- 154.Peng M, Lin W, Zhou A, Jiang Z, Zhou F, Wang Z. High genetic diversity and different type VI secretion systems in *Enterobacter* species revealed by comparative genomics analysis. BMC Microbiol. 2024;24(1):26-40.
- 155.Petrini O. Fungal endophytes of tree leaves. In: Microbial ecology of leaves. New York: Springer; 1991. p. 179-197.
- 156.Pieterse CM, Zamioudis C, Berendsen RL, Weller DM, Van Wees SC, Bakker PA. Induced systemic resistance by beneficial microbes. Annu Rev Phytopathol. 2014;52:347-375.
- 157.Pinski A, Betekhtin A, Hupert-Kocurek K, Mur LA, Hasterok R. Defining the genetic basis of plant-endophytic bacteria interactions. Int J Mol Sci. 2019;20(8):1947-1962.
- 158.Prabhu S, Poorniammal R, Dufossé L. Microbial metabolites: a sustainable approach to combat plant pests. Metabolites. 2025;15(6):418-430.
- 159. Pradhan V, Das P, Deb A, Bhadra P, Hossain A. Single-cell technology for plant biotechnology. In: Guide to plant single-cell technology. Cambridge (MA): Academic Press; 2025. p. 157-188.
- 160. Pramanic A, Sharma S, Dhanorkar M, Prakash O, Singh P. Endophytic microbiota of floating aquatic plants: recent developments and environmental prospects. World J Microbiol Biotechnol. 2023;39(4):96-112.
- 161.Prasanna GL. High throughput sequencing (HTS): an advanced technology used in characterization of the functions of the plant endophytes. Agric Food Enewsletter. 2022;4(2):36540-36548.
- 162.Raaijmakers JM, Vlami M, De Souza JT. Antibiotic production by bacterial biocontrol agents. Antonie Van Leeuwenhoek. 2002;81:537-547.
- 163.Rabiae M, Abdelglil MI, Foda N, Sayedahmed MM, Khedr EH, Abdel-Haleem M. Leveraging endophytic fungi for enhancing plant resilience to abiotic stresses. J Crop Health. 2025;77(5):141-155.
- 164.Radouane N, Lahmamsi H, Kenfaoui J, Legrifi I, Daaboub A, Goura K, *et al.* The role of endophytic bacteria in plant immune responses and plant disease control. In: Microbial biostimulants. Palm Bay (FL): Apple Academic Press; 2024. p. 177-205.
- 165.Rana N, Rathore A, Ghabru A, Chauhan S. Endophytes: role and applications in sustainable agriculture. Pharm Innov Int J. 2023;12(3):139-151.
- 166.Rashid MH, Chung YR. Induction of systemic resistance against insect herbivores in plants by beneficial soil microbes. Front Plant Sci. 2017;8:1816-1827.
- 167.Riseh RS, Fathi F, Vazvani MG, Tarkka MT. Plant colonization by biocontrol bacteria and improved plant health: a review. Front Biosci (Landmark Ed). 2025;30(1):23223-23238.
- 168.Riseh RS, Vatankhah M, Hassanisaadi M, Ait Barka E. Unveiling the role of hydrolytic enzymes from soil biocontrol bacteria in sustainable phytopathogen management. Front Biosci (Landmark Ed). 2024;29(3):105-120.
- 169.Roy S, Banerjee D. Volatile organic compounds from endophytic fungi. In: Gupta VK, Zeilinger S, editors. Recent advancement in white biotechnology through fungi. Volume 2: perspective for value-added products

- and environments. Cham: Springer International Publishing; 2019. p. 149-175.
- 170.Roychowdhury R, Mishra S, Anand G, Dalal D, Gupta R, Kumar A, *et al.* Decoding the molecular mechanism underlying salicylic acid (SA)-mediated plant immunity: an integrated overview from its biosynthesis to the mode of action. Physiol Plant. 2024;176(3):14399-14420.
- 171.Ryan RP, Germaine K, Franks A, Ryan DJ, Dowling DN. Bacterial endophytes: recent developments and applications. FEMS Microbiol Lett. 2008;278(1):1-9.
- 172.Sadaf K, Tushar L, Nirosha P, Podile AR, Sasikala C, Ramana CV. *Paenibacillus arachidis* sp. nov., isolated from groundnut seed. Int J Syst Evol Microbiol. 2016;66(8):2923-2928.
- 173. Sagita R, Quax WJ, Haslinger K. Current state and future directions of genetics and genomics of endophytic fungi for bioprospecting efforts. Front Bioeng Biotechnol. 2021;9:649906-649920.
- 174.Sahoo A, Meher I, Chhotaray SP, Meena M, Swapnil P, Arora J, *et al.* Fungal endophytes: a resource for producing extracellular enzymes. In: Metabolic sustainability of endophytes: current status, challenges and potential. Singapore: Springer Nature; 2025. p. 105-145.
- 175.Sahu PK, Mishra S. Effect of hybridization on endophytes: the endo-microbiome dynamics. Symbiosis. 2021;84(3):369-377.
- 176.Santoyo G, Moreno-Hagelsieb G, del Carmen Orozco-Mosqueda M, Glick BR. Plant growth-promoting bacterial endophytes. Microbiol Res. 2016;183:92-99.
- 177. Sena L, Mica E, Valè G, Vaccino P, Pecchioni N. Exploring the potential of endophyte-plant interactions for improving crop sustainable yields in a changing climate. Front Plant Sci. 2024;15:1349401-1349412.
- 178. Shaffique S, Khan MA, Wani SH, Pande A, Imran M, Kang SM, *et al.* A review on the role of endophytes and plant growth-promoting rhizobacteria in mitigating heat stress in plants. Microorganisms. 2022;10(7):1286-1305.
- 179.Sharma A, Kumar P, Pahal V, Kumar J, Pandey SS. Endophytic phytohormone production and utilization of functional traits in plant growth promotion. In: Plant microbiome for plant productivity and sustainable agriculture. Singapore: Springer Nature; 2023. p. 365-385.
- 180.Sharma V, Kumari A, Meena AK, Sharma AK, Meena PK, Gambhir AYK. Advancing precision farming through microbial genomics and biotechnological tools: innovations for improving crop productivity and soil health. Growth. 2025;112:140-156.
- 181.Shastry RP, Rai VR. Quorum-quenching endophytes: a novel approach for sustainable development of agroecosystem. In: Endophytes. Cham: Springer International Publishing; 2017. p. 41-57.
- 182.Shishodia SK, Thakur R, Shankar J, Singh R, De LosRios-Escalante P. Unlocking endophytic mysteries by leveraging multi-omics technologies. In: Multi-omics approach to investigate endophyte diversity. Singapore: Springer Nature; 2025. p. 125-44.
- 183.Shishodia SK, Thakur R, Shankar J, Singh R, De LosRios-Escalante P. Unlocking endophytic mysteries by leveraging multi-omics technologies. In: Multi-

- omics approach to investigate endophyte diversity. Singapore: Springer Nature; 2025. p. 125-44.
- 184.Singh SK, Kumar D, Shamim M, Sharma R, editors. Applied mycology for agriculture and foods: industrial applications. Boca Raton: CRC Press; 2023.
- 185. Sivalingam P, Easwaran M, Ganapathy D, Basha SF, Poté J. Endophytic *Streptomyces*: an underexplored source with potential for novel natural drug discovery and development. Arch Microbiol. 2024;206(11):442-56.
- 186.Soni R, Keharia H. Phytostimulation and biocontrol potential of Gram-positive endospore-forming *Bacilli*. Planta. 2021;254(3):49-62.
- 187. Srivastava P, Tiwari SP, Srivastava AK, Sharma R. Optimization of sterilization parameters for isolation of endophytes from *Allium sativum* and exploring its antibacterial activity. J Pure Appl Microbiol. 2024;18(2):1081-90.
- 188. Stierle A, Strobel G, Stierle D. Taxol and taxane production by *Taxomyces andreanae*, an endophytic fungus of Pacific yew. Science. 1993;260(5105):214-6.
- 189.Sun K, Xie XG, Lu F, Zhang FM, Zhang W, He W, *et al.* Peanut preinoculation with a root endophyte induces plant resistance to soil-borne pathogen *Fusarium oxysporum* via activation of salicylic acid-dependent signaling. Plant Soil. 2021;460(1):297-312.
- 190. Tabassum N, Ahmed HI, Parween S, Sheikh AH, Saad MM, Krattinger SG, *et al.* Host genotype, soil composition, and geo-climatic factors shape the fonio seed microbiome. Microbiome. 2024;12(1):11-24.
- 191. Tariq A, Tanvir A, Barasarathi J, Alsohim AS, Mastinu A, Sayyed R, *et al.* Endophytes: key role players for sustainable agriculture: mechanisms, omics insights and future prospects. Plant Growth Regul. 2025;101(2):1-22.
- 192. Thomas BO, Lechner SL, Ross HC, Joris BR, Glick BR, Stegelmeier AA. Friends and foes: bacteria of the hydroponic plant microbiome. Plants. 2024;13(21):3069-82.
- 193. Tonelli ML, Figueredo MS, Rodríguez J, Fabra A, Ibañez F. Induced systemic resistance-like responses elicited by rhizobia. Plant Soil. 2020;448(1):1-4.
- 194. Trivedi P, Leach JE, Tringe SG, Sa T, Singh BK. Plant-microbiome interactions: from community assembly to plant health. Nat Rev Microbiol. 2020;18(11):607-21.
- 195.Stotz HU, Brotherton D, Inal J. Communication is key: extracellular vesicles as mediators of infection and defence during host-microbe interactions in animals and plants. FEMS Microbiol Rev. 2022;46(1):044-59.
- 196. Ullah A, Nisar M, Ali H, Hazrat A, Hayat K, Keerio AA, *et al.* Drought tolerance improvement in plants: an endophytic bacterial approach. Appl Microbiol Biotechnol. 2019;103(18):7385-97.
- 197.Utturkar SM, Cude WN, Robeson MS Jr, Yang ZK, Klingeman DM, Land ML, *et al.* Enrichment of root endophytic bacteria from *Populus deltoides* and single-cell genomics analysis. Appl Environ Microbiol. 2016;82(18):5698-708.
- 198. Van Elsas JD, Hartmann A, Schloter M, Trevors JT, Jansson JK. The bacteria and archaea in soil. In: Modern soil microbiology. 3rd ed. Boca Raton: CRC Press; 2019. p. 49-64.
- 199. Vasileva EN, Akhtemova GA, Zhukov VA, Tikhonovich IA. Endophytic microorganisms in

- fundamental research and agriculture. Ecol Genet. 2019;17(1):19-32.
- 200. Velmourougane K, Prasanna R, Saxena AK. Agriculturally important microbial biofilms: present status and future prospects. J Basic Microbiol. 2017;57(7):548-73.
- 201. Venkatesh Kumar R, Singh RP, Mishra P. Endophytes as emphatic communication barriers of quorum sensing in Gram-positive and Gram-negative bacteria—a review. Environ Sustain. 2019;2(4):455-68.
- 202. Verma H, Kumar D, Kumar V, Kumari M, Singh SK, Sharma VK, *et al.* The potential application of endophytes in management of stress from drought and salinity in crop plants. Microorganisms. 2021;9(8):1729-46.
- 203. Verma V, Srivastava A, Garg SK, Singh VP, Arora PK. Incorporating omics-based tools into endophytic fungal research. Biotechnol Notes. 2024;5:1-7.
- 204. Von Tubeuf CF. Biologische Bekämpfung von Pilzkrankheiten der Pflanzen. Naturwiss Z Forst Landwirtsch. 1914;12:11-9.
- 205. Wallis CM. Nutritional niche overlap analysis as a method to identify potential biocontrol fungi against trunk pathogens. BioControl. 2021;66(4):559-71.
- 206. Wang Y, Hao J, Gu J, Wu J, Zhang Y, Liang T, et al. Genome-driven functional validation of *Bacillus amyloliquefaciens* strain MEPW12: a multifunctional endophyte for sustainable sweet potato cultivation. Microorganisms. 2025;13(6):1322-38.
- 207. Wani AK, Akhtar N, Singh R, Chopra C, Kakade P, Borde M, *et al.* Prospects of advanced metagenomics and meta-omics in the investigation of phytomicrobiome to forecast beneficial and pathogenic response. Mol Biol Rep. 2022;49(12):12165-79.
- 208. Wijesekara T, Xu B. Health-promoting effects of bioactive compounds from plant endophytic fungi. J Fungi. 2023;9(10):997-1012.
- 209.Xu H, Gao J, Portieles R, Du L, Gao X, Borras-Hidalgo O. Endophytic bacterium *Bacillus aryabhattai* induces novel transcriptomic changes to stimulate plant growth. PLoS One. 2022;17(8):e0272500.
- 210.Xu L, Wu C, Oelmüller R, Zhang W. Role of phytohormones in *Piriformospora indica*-induced growth promotion and stress tolerance in plants: more questions than answers. Front Microbiol. 2018;9:1646-60.
- 211. Yang YX, Ahammed GJ, Wu C, Fan SY, Zhou YH. Crosstalk among jasmonate, salicylate and ethylene signaling pathways in plant disease and immune responses. Curr Protein Pept Sci. 2015;16(5):450-61.
- 212. Yanti Y. Involvement of jasmonic acid in the induced systemic resistance of tomato against *Ralstonia syzygii* subsp. *indonesiensis* by indigenous endophyte bacteria. IOP Conf Ser Earth Environ Sci. 2019;347(1):012024.
- 213. Younas H, Younas F. Phytohormone production by endophytic fungi. In: Fungal endophytes. Vol. I. Singapore: Springer Nature; 2025. p. 385-413.
- 214. Yu X, Xia J, Bi S, Wang H, Zhao C. Whole-genome analysis and growth-promoting mechanism of *Klebsiella pneumoniae* YMK25 from maize rhizobacteria. Plants. 2025;14(17):2738-51.
- 215. Zaynab M, Fatima M, Abbas S, Sharif Y, Umair M, Zafar MH, *et al.* Role of secondary metabolites in plant

- defense against pathogens. Microb Pathog. 2018;124:198-202.
- 216.Zhang X, Tong J, Dong M, Akhtar K, He B. Isolation, identification and characterization of nitrogen-fixing endophytic bacteria and their effects on cassava production. PeerJ. 2022;10:e12677.
- 217. Zhao C, Onyino J, Gao X. Current advances in the functional diversity and mechanisms underlying endophyte-plant interactions. Microorganisms. 2024;12(4):779-95.
- 218. Zou H, Xu J, An M, Li H, Tian L, Ma Y. Community structure of habitat microorganisms and endophytes of narrow-ranged species *Corybas fanjingshanensis* at different ecological niche specificities. BMC Plant Biol. 2025;25(1):993-1008.