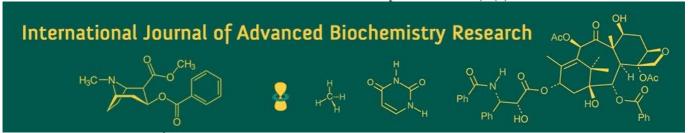
International Journal of Advanced Biochemistry Research 2025; 9(8): 808-811



ISSN Print: 2617-4693 ISSN Online: 2617-4707 NAAS Rating (2025): 5.29 IJABR 2025; 9(8): 808-811 www.biochemjournal.com Received: 08-05-2025 Accepted: 12-06-2025

Tushar Maharishi

Ph.D Scholar, Department of Veterinary Medicine, Rajasthan University of Veterinary and Animal Sciences, Bikaner, Rajasthan, India

Sita Ram Gupta

Assistant Professor,
Department of Veterinary
Medicine, Rajasthan
University of Veterinary and
Animal Sciences, Bikaner,
Rajasthan, India

Manohar Lal Sain

Assistant Professor,
Department of Veterinary
Medicine, Rajasthan
University of Veterinary and
Animal Sciences, Bikaner,
Rajasthan, India

Sunita Choudhary

Assistant Professor, Department of Veterinary Medicine, Rajasthan University of Veterinary and Animal Sciences, Bikaner, Rajasthan, India

JP Kachhawa

Assistant Professor,
Department of Veterinary
Medicine, Rajasthan
University of Veterinary and
Animal Sciences, Bikaner,
Rajasthan, India

Tara Chand Nayak

Assistant Professor,
Department of Veterinary
Medicine, Rajasthan
University of Veterinary and
Animal Sciences, Bikaner,
Rajasthan, India

Corresponding Author: Tushar Maharishi

Ph.D Scholar, Department of Veterinary Medicine, Rajasthan University of Veterinary and Animal Sciences, Bikaner, Rajasthan, India

Prevalence of subclinical mastitis in cattle in Bikaner

Tushar Maharishi, Sita Ram Gupta, Manohar Lal Sain, Sunita Choudhary, JP Kachhawa and Tara Chand Nayak

DOI: https://www.doi.org/10.33545/26174693.2025.v9.i8k.5344

Abstract

The present study was conducted on 472 quarter milk samples from 120 apparently healthy lactating cows to determine the quarter, parity and lactation stage-wise prevalence of sub-clinical mastitis in cattle based on the modified California mastitis test (MCMT). The animal and overall quarter-wise prevalence of sub-clinical mastitis were 37.16 percent and 22.03 percent, respectively. The quarter type-wise highest prevalence of SCM was found in right hind quarters (32.76%), followed by left hind quarters (23.52%), right fore quarters (19.65%) and least prevalence was recorded in left fore quarters (12.50%). The parity-wise highest prevalence of SCM was noticed in IVth parity followed by IIIrd, Vth, VIth, VIIth and above, IInd and least in Is^t parity. Whereas, lactation stage-wise, the highest prevalence of SCM was recorded in early lactating stage 25 (43.86%) followed by mid-lactating stage 17(36.20%) and least in late lactating stage 03 (18.75%) by modified California mastitis test.

Keywords: Lactation stage, modified California mastitis test, parity, subclinical mastitis

Introduction

Bovine mastitis is a multiple etiological disease involving inflammation of the mammary gland parenchyma and characterized by physical, chemical, and bacteriological changes in milk (Constable *et al.*, 2017) ^[8]. Etiologically about 135-150 bacterial species, sub-species and serovariants have been isolated from bovine mammary glands affected with mastitis. Majority of the mastitis cases are caused by staphylococci, streptococci, and *E. coli* bacteria (Bradley, 2002) ^[6]. Subclinical mastitis (SCM) does not cause any visible changes in milk or udder appearance but affects milk quality and quantity causing a reduction in milk yield up to two-third losses of the total milk production (Haque, 2014; Singh *et al.*, 2015) ^[13, 32], altered milk composition, and the presence of inflammatory components and bacteria in milk. The diagnosis of bovine mastitis involves recognition of disease and identification of the causative pathogens (Ashraf and Imran, 2018) ^[2].

To screen the milk for sub-clinical mastitis various direct and indirect methods can be employed for diagnosis such as California mastitis test (CMT), from the milk samples. CMT is an animal-side test and a qualitative measurement of the somatic cells in the mastitic milk. (Duarte *et al.*, 2015; Ashraf and Imran, 2018) [32, 2].

Organism as diverse as bacteria, viruses, mycoplasma, yeast, and algae are involve in the infection. Although the majority of subclinical mastitis is of bacterial origin and *Staphylococcus* spp. *Streptococcus* spp. *Escherichia coli* are the most common bacterial pathogen isolated from sub clinical mastitis in cows by various workers (Kachhawa, 2018; Choudhary, 2018; Savita *et al.*, 2020 and Solanki, 2021) [15, 27, 34].

To manage these infections, farmers and veterinarians frequently rely on antimicrobial drugs. However, the widespread and sometimes indiscriminate use of these drugs has led to the development of antimicrobial resistance (AMR) among the pathogens responsible for mastitis (Kumar *et al.*, 2012; Raorane *et al.*, 2013; Poojitha *et al.*, 2022) [16, 22, 21]. The Bikaner region in Rajasthan, known for its arid climate and traditional dairy practices, is particularly vulnerable to this condition due to environmental stressors and management challenges. Factors such as climate, animal husbandry practices, and breed susceptibility are known to influence of subclinical mastitis (SCM) occurrence in this region, adding complexity to its detection and control (Shabaz *et al.*, 2020; Bhati *et al.*, 2021) [29, 5]. The present study was conducted to detect the prevalence of subclinical mastitis in cattle within the Bikaner region.

Materials and Methods Source of animals

In the present study, 472 functional quarters of 120 apparently healthy lactating cows were screened for subclinical mastitis using modified California mastitis from unorganized dairy farms as well as animals of individual holdings in and around Bikaner. The cows in early lactation (<1 month post-calving) and late lactation were excluded from the study.

Ethical Approval

This study was approved by Institutional Animal Ethics Committee (IAEC) of the College of Veterinary and Animal Science, Rajasthan University of Veterinary and Animal Sciences, Bikaner with approval No. CVAS/IAEC/2024-25/17).

Collection of milk sample

Approximately 30 ml of milk from respective quarters were collected aseptically into sterile test tubes and marked as right fore, right hind, left fore and left hind. Milk samples were immediately transported to the laboratory on ice and maintained at 4 °C until further testing.

Modified California mastitis test

The California mastitis test was developed by Schalm and Noorlander (1957) [28]. The MCMT is a simple, rapid, highly sensitive and inexpensive test that accurately predicts the inflammatory cell counts and infection in milk from individual quarters or pooled milk samples. In the present study, Ezee was used in place of pure laryl sulphates or sulphonates of sodium or potassium as an anionic surface active agentand Bromo thymol blue was replaced by bromocresol purple (Savita *et al.*, 2020) [27].

Procedure

The test was carried out with 3 ml of milk from each quarter into the respective 4 cups in plastic paddle. An equal amount of the above test reagent was added in each cup and gently mixed by circular movement of the paddle in horizontal plane. The total cell count is reflected by the degree of precipitation or gel formation that occurs. The pH change associated with abnormal milk is indicated by a color reaction with bromocresol purple. California mastitis test reactions were scored according to Radostits *et al.* (2007) [23] as follows.

Results and Discussion

In the present study, 472 functional quarters of 120 apparently healthy lactating cattle were screened for subclinical mastitis by the modified California mastitis test (MCMT)

Animal-wise prevalence of subclinical mastitis

In the present study, out of 120 cattle, 45 (37.16%) cattle were found positive for subclinical mastitis based on MCMT. The prevalence obtained in the current study was almost in line with studies by Savita *et al.* (2020) ^[27] Gupta (2021) ^[12] and Sain (2022) ^[25]. Who reported, animal-wise prevalence was 49.78 percent, 38 percent and 38.87 percent, respectively.

Overall quarter-wise prevalence of subclinical mastitis

In the current study, out of 472 quarters, 104 (22.03%) quarters were found positive for subclinical mastitis based

on MCMT. In the present investigation, overall quarter-wise prevalence of subclinical mastitis was recorded 17.99. percent, 26.18 percent, 16.30 percent and 28.06 percent in left side quarters, right side quarters, fore quarters and hind quarters, respectively.

Data of the present study revealed that right-side quarters showed the highest prevalence as compared to left-side quarters.

Quarter wise prevalence of subclinical mastitis

The quarter wise prevalence of subclinical mastitis was recorded as 12.50 percent, 23.52 percent, 19.65 percent and 32.76 percent in left fore, left hind, right fore and right hind quarters, respectively based on MCMT. In the current study, it was observed that the prevalence of subclinical mastitis was higher in the hindquarters as compared to the fore quarters. It was also noticed that the highest prevalence of SCM was recorded in right hindquarters, followed by left hind quarters, right fore quarters and least in left fore quarters.

Findings of the present study are in agreement with the observations of Gupta (2021) [12], Sain (2022) [25] and Gautam (2022) [11] who also observed the highest quarterwise prevalence of subclinical mastitis in right hind quarters followed by left hind quarters, right fore quarters and least in left fore quarters detected by MCMT.

Kachhawa (2018) [15], Choudhary (2018) [7] and Solanki (2021) [34], Kiran (2023) [17] and Sharmila (2023) [30] also revealed that the prevalence of subclinical mastitis was higher in the hind quarters than in the fore quarters. Right hind quarters were more affected than the left fore quarters Gautam *et al.*, 2022 [11] Dangi *et al.* 2025 [9] also observed that right hind quarters were most frequently affected (16.66%) in cattle.

However, Barmendra *et al.* (2011) ^[4], Mustafa *et al.* (2011) ^[20], Shittu *et al.* (2012) ^[31] and Srinivasan *et al.* (2013) ^[35] found higher prevalence in fore quarter than the hind quarter. Due to increased exposure to feces, milkmen's custom of approaching from the side when milking high urine and production capacity, large mass and capacity, increased susceptibility to direct trauma and comparatively closer proximity to the floor than in the forequarters, the higher prevalence of subclinical mastitis was observed in the hind quarters (Akhtar *et al.* 2012) ^[1].

Parity-wise prevalence of subclinical mastitis

In the present study 120 cattle were screened. Out of these, 12, 13, 20, 35, 14, 16 and 10 cattle were in first (Ist), second (IInd), third (IIIrd), forth (IVth), fifth (Vth), sixth (VIth) and seventh (VIIth) and above parity, respectively In the present study based on positive in MCMT, 16.66 percent, 23.80 percent, 40.0 percent, 54.14 percent, 42 percent, 31.25 percent and 20 percent cattle found positive for SCM were in their Ist, IInd, IIIrd, IVth, V th, VIth and VIIth and above parity, respectively. Parity wise highest prevalence of SCM was observed in IVth parity, followed by Vth, IIIrd, VIth, VIIth, IInd and least in Ist parity.

Data analysis revealed that the prevalence of subclinical mastitis in cattle was lowest in the first parity, then increased until the IVth parity, after which it remain almost steady. In agreement with Gupta (2021) [12] and Sain (2022) [25], Gautam (2022) [11] Sharmila (2023) [30] and Kiran (2023) [17] all reported similar findings of a higher prevalence of subclinical mastitis in IVth parity.

Devi *et al.* (1997) [10] reported lowest prevalence of SCM during first parity than increased with subsequent parities which are also in accordance to the present study. This could be due to lowered resistance of the animals as lactation number increased and improper functioning of the teat sphincter results of increased incidence of new intramammary infection (Singh and Baxi, 1980, Devi *et al.* 1997 and Sampimon *et al.*, 2009) [33, 10, 26].

Lactation stage-wise prevalence of subclinical mastitis

In current study, out of 120 screened cows 57, 47 and 16 were in the early, mid and late lactation stage, respectively. The cows in early lactation (less than one month after calving) or late lactation (more than nine months after calving) were not included in the study. On the basis of MCMT, out of 120 cattle, 25(43.86%), 17(36.20%) and 3(18.75%) were found positive from early, mid and late lactation stage, respectively. Highest prevalence of subclinical mastitis was observed in early lactating stage followed by mid lactating stage and least in late lactation stage.

The findings of current study are consistent with those of Choudhary (2018) [7], Mourya *et al.* (2020) [19] and Gupta (2021) [12], Sain (2022) [25], Gautam (2022) [11], Kiran (2023) [17] who also found that the majority of subclinical mastitis occurred in the early stage of lactation as compared to the mid and late stages. The maximum prevalence of SCM was recorded as 30.0% and 47.46%, during the early lactation period by Islam *et al.* (2011) [14] and Maheshwari *et al.* (2016) [18], respectively. Badiuzzamann *et al.* (2015) [3], found that the early lactation stage had the highest prevalence of subclinical mastitis, followed by the mid and late lactation stages.

Negative energy balance, the physiological stress of lactation, changes in homeostasis mechanisms and postpartum rapid physiological changes in the mammary tissue that result in lower or reduced udder resistance could all be contributing factors to the higher prevalence of subclinical mastitis in the early lactation stage, (Rasool *et al.*, 1985 and Mourya *et al.*, 2020) [24, 19]. stated that a cow's body undergoes numerous physiological changes in the early stages of lactation, which results in ongoing stress and weakened immunity.

Conclusions

In the present experiment, animal-wise prevalence of SCM was observed as 37.16 percent and quarter-wise prevalence of SCM was recorded as 22.03 percent by modified California mastitis test. The prevalence of SCM in fore quarters was noted as 16.30 percent and in hindquarters was recorded as 28.06 percent. Right hindquarters were more affected than the left fore quarters. Parity wise highest prevalence of SCM was observed in IVth parity. Lactation stage wise highest prevalence of subclinical mastitis was found in the early lactation stage followed by mid-lactation and least in the late lactation stage.

References

- Akhtar A, Ameer M, Aeshad M. Prevalence of sub clinical mastitis in buffaloes in district Dera Ismail Khan. Pakistan Journal of Science. 2012;64(2):132-138.
- Ashraf A, Imran M. Diagnosis of bovine mastitis: From laboratory to farm. Trop Anim Health Prod. 2018;50:1193-1202.

- 3. Badiuzzaman M, Samad MA, Siddiki SHMF, Islam MT, Saha S. Subclinical mastitis in lactating cows: comparison of four screening tests and effect of animal factors on its occurrence. Bangladesh Journal of Veterinary Medicine. 2015;13(2):41-50.
- 4. Barmendra S, Rahman MSU, Hannan MA, Rahman MM, Bhuiyan MM, Bari FY. Prevalence of mastitis in dairy cows in selected areas of Bangladesh. International Journal of Bioresearch. 2011;1:5-9.
- 5. Bhati T, Chhabra R, Yadav R, Charaya G, Kataria AK. Antimicrobial resistance profiling of *Staphylococcus aureus* isolated from mastitic milk of bovine and dairy environment from arid and semi-arid regions of India. Ruminant Science. 2021;10(1):57-66.
- 6. Bradley AJ. Veterinary drug usage and antimicrobial resistance in bacteria of animal origin. Basic Clin Pharmacol Toxicol. 2002;96:271-281.
- 7. Choudhary S. Studies on diagnosis and therapeutic trial for subclinical mastitis in indigenous cattle [PhD thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2018.
- 8. Constable PD, Hinchcliff KW, Done SH, Grünberg W. Veterinary Medicine. A textbook of diseases of cattle, horses, sheep, pigs and goats. 11th ed. St. Louis (MO): Elsevier; 2017. p.1912.
- 9. Dangi R, Choudhary NS, Mehta HK, Gangil R, Agrawal V, Singh M, *et al.* Prevalence, risk factors, and isolation of primary causative agent of subclinical mastitis in dairy cattle. Journal of Advances in Biology & Biotechnology. 2025;28(3):238-249.
- 10. Devi BK, Shukla PC, Bagherwal RK. Incidence of subclinical mastitis in cows. Indian Journal Dairy Science. 1997;50:477-478.
- 11. Gautam S. Studies on immuno-therapeutic and antioxidative potential of *Phyllanthus emblica* (amla) in subclinical mastitis in cattle [MVSc thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2022.
- 12. Gupta SR. Studies on therapeutic potential of *Nigella sativa* (kalonji) and *Phyllanthus emblica* (amla) in bovine subclinical mastitis [PhD thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2021.
- 13. Haque ME. Rapid detection of subclinical mastitis in dairy cow. J Fisheries Livest Prod. 2014;3:128.
- Islam MA, Islam MZ, Islam MA, Rahman MS, Islam MT. Prevalence of subclinical mastitis in dairy cows in selected areas of Bangladesh. Bangladesh Journal of Veterinary Medicine. 2011;9(1):73-78.
- 15. Kachhawa JP. Studies of therapeutic and antioxidative potential of *Withania somnifera*, *Citrullus colocynthis* and *Piper nigrum* in subclinical mastitis in crossbred cattle [PhD thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2018.
- 16. Kumar V, Patel JS, Patel BR, Mevada VK, Raval AP. Therapeutic efficacy of antimicrobial drugs in clinical mastitis of crossbred cattle. Ruminant Science. 2012;1(2):177-180.
- 17. Kumari K. Studies on therapeutic potential of polyherbal formulation in subclinical mastitis in cattle [MVSc thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2023.
- 18. Maheshwari P, Shukla PC, Rao MLV, Shukla SN. Occurrence of subclinical mastitis in cattle in and

- around Jabalpur, Madhya Pradesh. Haryana Veterinarian. 2016;55(2):160-162.
- 19. Mourya A, Shukla PC, Gupta DK, Sharma RK, Nayak A, Singh B, *et al.* Prevalence of subclinical mastitis in cows in and around Jabalpur, Madhya Pradesh. Journal of Entomology and Zoology Studies. 2020;8(4):40-44.
- 20. Mustafa YS, Awan FN, Zaman T, Chaudhry SR, Zoyfro V. Prevalence and antibacterial susceptibility in mastitis in buffalo and cattle in District Lahore, Pakistan. Journal of Buffalo Science. 2012;1(1):69-72.
- 21. Poojitha R, Shrivastav A, Kumar N, Shrivastav N, Singh SK, Ranjan R. Study of antibiotic resistance pattern in isolated extended spectrum beta-lactamase producing bacteria from milk of healthy cattle. Ruminant Science. 2022;11(1):187-192.
- 22. R A, Chothe S, Dubal ZB, Barbuddhe SB, Karunakaran M, Doijad S, *et al.* Antimicrobial resistance of the pathogens isolated from bovine mastitis in Goa. Ruminant Science. 2013;2(2):139-144.
- 23. Radostits OM, Gay CC, Hinchcliff KW, Constable PD. Mastitis. In: Veterinary Medicine: Textbook of diseases of cattle, sheep, pigs, goats and horses. 10th ed. Philadelphia (PA): Saunders Elsevier; 2007. p.673-748.
- Rasool G, Jabbar MA, Kazmi SE, Ahmad A. Incidence of sub-clinical mastitis in Nili-Ravi buffaloes and Sahiwal cows. Pakistan Veterinary Journal. 1985;5:76-78.
- 25. Sain ML. Studies on therapeutic and antioxidative potential of *Tinospora cardifolia* (Giloy) and *Azadirachta indica* (Neem) in subclinical mastitis in cattle [PhD thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2022.
- 26. Sampimom O, Barkema HW, Berenda I, Sol J, Lam T. Prevalence of intramammary infection in Dutch dairy herds. Journal of Dairy Research. 2009;76:129-136.
- 27. Savita AC, Nayak TC, Marwaha S. Prevalence of subclinical mastitis in cattle using modified California mastitis test. Veterinary Journal. 2020;57:723-729.
- 28. Schalm OW, Noorlander DO. Experiments and observations leading to the development of the California mastitis test. J Am Vet Med Assoc. 1957;130:199-204.
- 29. Shabaz SS, Prameela DR, Sreenivasulu D, Sujatha K. Evaluation of on-farm milk culture system for identification of mastitis pathogens. Ruminant Science. 2020;9(2):363-372.
- 30. Sharmila K. Evaluation of therapeutic potential of kalonji (*Nigella sativa*) in subclinical mastitis in cattle [MVSc thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2023.
- 31. Shittu A, Abdullahi J, Jibril A, Mohammed AA, Fasina FO. Sub-clinical mastitis and associated risk factors on lactating cows in the Savannah Region of Nigeria. BMC Vet Res. 2012;8(1):134.
- 32. Singh D. Diagnostic and therapeutic studies on subclinical mastitis in cows [MVSc thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2015. Duarte CM, Freitas PP, Bexiga R. Technological advances in bovine mastitis diagnosis: an overview. J Vet Diagn Investig. 2015;27:665-672.
- 33. Singh KB, Baxi KK. Studies on the incidence and diagnosis of sub-clinical mastitis in dairy cows. Indian Veterinary Journal. 1980;57:723-729.

- 34. Solanki P. Therapeutic studies of Neem (*Azadirachta indica*) and Giloy (*Tinospora cordifolia*) on subclinical mastitis in cattle [MVSc thesis]. Bikaner: Rajasthan University of Veterinary and Animal Sciences; 2021.
- 35. Srinivasan P, Jagadeswaran D, Manoharan R, Giri T, Balasubramaniam GA, Balachandran P. Prevalence and etiology of subclinical mastitis among buffaloes (*Bubalus bubalus*) in Namakkal, India. Pakistan Journal of Biological Sciences. 2013;16(23):1761-1780.