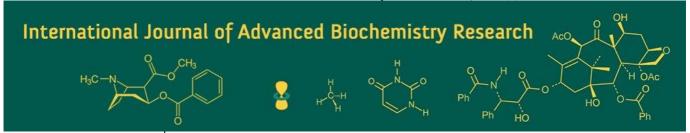
International Journal of Advanced Biochemistry Research 2025; SP-9(7): 199-204



ISSN Print: 2617-4693 ISSN Online: 2617-4707 IJABR 2025; SP-9(7): 199-204 www.biochemjournal.com Received: 25-04-2025 Accepted: 29-05-2025

Sheela P

Assistant Professor, Department of Veterinary Microbiology, KVAFSU, Veterinary College, Hassan, Karnataka, India

Ashwini A

Department of Veterinary Medicine, KVAFSU, Veterinary College, Hassan, Karnataka, India

Bhagya BK

Department of Veterinary Medicine, KVAFSU, Veterinary College, Hassan, Karnataka, India

Pralhad

Department of Veterinary Microbiology, KVAFSU, Veterinary College, Hassan, Karnataka, India

Sundareshan S

Office of the Deputy Director, Department of Animal Husbandry and Veterinary Sciences, Hassan, Karnataka, India

Sathisha KB

Livestock Research and Information Center (Sheep), KVAFSU, Nagamangala, Karnataka, India

Ramesh D

Department of Veterinary Physiology and Biochemistry, KVAFSU, Veterinary College, Hassan, Karnataka, India

Corresponding Author: Sheela P

Assistant Professor, Department of Veterinary Microbiology, KVAFSU, Veterinary College, Hassan, Karnataka, India

A study on *in-vitro* anti microbial sensitivity pattern of the bacterial isolates obtained from bovine mastitis

Sheela P, Ashwini A, Bhagya BK, Pralhad, Sundareshan S, Sathisha KB and Ramesh D

DOI: https://www.doi.org/10.33545/26174693.2025.v9.i7Sc.4783

Abstract

A study was under taken to estimate the prevalence of bacterial agents associated with bovine mastitis in and around Shivamogga and Hassan of Karnataka State, India and to know their invitro susceptibility and resistance patterns to the commonly used antimicrobial drugs in the treatment of mastitis. A total of 117 milk samples were collected and subjected to bacteriological studies and in-vitro antimicrobial sensitivity test. In all, 117 milk samples subjected for isolation of bacteria yielded 132 bacterial isolates. The most predominant bacteria isolated were *Staphylococcus aureus* (27.3%) followed by *Streptococcus* spp. (26.5%), *E. coli* (18.2%), *Klebsiella pneumoniae* (12.1%), *Pseudomonas aeruginosa* (5.3%), *Proteus mirabilis* (0.76%) and *Trueperella pyogenes* (0.76%). The antibiogram studies revealed that most of the bacterial isolates were sensitive to chloramphenicol (56.8%) followed by enrofloxacin (36.36%) and cotrimoxazole (33.3%). However, due to the concerns about drug residues with chloramphenicol causing potential human health risks, the use of drugs like enrofloxacin and cotrimoxazole is proposed for the treatment of bovine mastitis in the study area.

Keywords: Antimicrobial sensitivity, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, drug residues, enrofloxacin

Introduction

Bovine mastitis is an inflammatory condition of the mammary gland in dairy cattle, primarily caused by bacterial infections. The infection develops when the organisms' defense mechanisms are weakened. Mechanical trauma, thermal trauma, and chemical insult also predispose the gland to intramammary infection (IMI). Occurrence of mastitis depends on the interaction of host, agent, and environmental factors. It is an important production related disease of dairy animals as it is directly or indirectly affecting the economy of the farmers and ultimately the country (Miller et al., 1993) [9]. It is estimated that mastitis alone accounts for about 70 percent of all avoidable losses incurred during milk production. At least 137 species of microorganisms from a broad phylogenetic spectrum, including bacteria, yeast, fungi and algae, are associated with bovine mastitis (NAAS, 2013) [11]. In Asia, major mastitis causing organisms are Staphylococcus aureus, Streptococcus species, Escherichia coli, Corynebacterium species and Klebsiella species. Mastitis can be caused by contagious pathogens like Staphylococcus aureus, Streptococcus agalactiae, Mycoplasma bovis and Corynebacterium bovis that reside primarily in the mammary gland of cows and multiply enormously in infected mammary gland and by environmental pathogens like E. coli, Streptococcus uberis, Klebsiella species, Pseudomonas aeruginosa (through contaminated intramam-mary tubes) that are present in the environment which enter the udder through contamination of teat ends (Quin et al., 2011) [13].

A knowledge of the predominant bacterial species associated with bovine mastitis in a particular geographical area is required for its effective treatment and control. To prevent the development of antimicrobial resistance and indiscriminate use of antimicrobial drugs, a knowledge of the antimicrobial profile of the bacterial species associated with mastitis is of paramount importance. Hence a study was under taken to know the etiological agents associated with mastitis and to know their invitro susceptibility and resistance patterns.

Materials and Methods

A total of 117 milk samples collected from clinical and subclinical cases of bovine and bubaline mastitis, presented to the veterinary clinical complexes (VCC) of the veterinary colleges at Shivamogga and Hassan during the period of 2013-2015 and 2021-2023 respectively and maintained in the Department of Veterinary Microbiology were used in the present study.

Culture and biochemical tests

The milk samples were inoculated into brain heart infusion broth (BHI) and tryptose phosphate broth (TPB) and thioglycolate broth (TGB) and incubated at 37° C for 12-18 hours under aerobic conditions and in anaerobic gas jar respectively. After the preliminary identification of the bacterial species by the staining morphology of the broth culture they were plated on to BHI agar, Mannitol salt agar (MSA), Mc-Conkey agar, EMB agar, Edward's agar and blood agar. Identification of bacteria on primary culture was done based on hemolytic characteristics, colony morphology, Gram-stain reaction including shape and arrangements of the bacteria. Staphylococci were identified using growth characteristics, tube coagulase tests, and catalase tests. Identification of Streptococcus isolates was made on the basis of colony morphology, staining morphology and catalase tests. Gram-negative bacteria grown on MacConkey agar were differentiated according to their growth characteristics, catalase test, oxidase reaction, triple sugar-iron agar test, urease test and the IMViC (indole, methyl red, Voges-Proskauer, citrate) test. Gram positive rods (pleomorphic) by their staining morphology, haemolytic pattern and catalase test.

Antibiogram studies

All the bacterial isolates obtained were subjected to antibiogram studies by employing the Kirby-Bauer disk diffusion method. A total of 13 different antimicrobial drugs commonly used in the treatment of mastitis *viz.*, ampicillin cloxacillin (A), amoxycillin with clavulanic acid (AMC), amikacin (AK), co-trimoxazole (sulfamethoxazole with trimethoprim) (COT), ceftriaxone with tazobactum (CIT), cefotaxime (CTX), cefoxitin (CX), ceftriaxone (CTR), cefoperazone (CPZ), chloramphenicol (C), enrofloxacin (EX), gentamicin (GEN) and tetracycline (TE) were used in the present study. The interpretations were carried out as per CLSI standards, 2013.

Results

Out of the 117 milk samples subjected to culture for isolation of bacteria, 11 samples did not yield any growth, 22 samples revealed two bacteria and 4 samples yielded three bacteria (mixed infection) and the remaining 80 samples yielded single bacteria. Seven other isolates (gram positive rods) of the present study could not be assigned any species/genus. The most predominant bacteria isolated were Staphylococcus aureus (27.3%) followed by Streptococcus spp (26.5%), E. coli (18.2%), Klebsiella pneumoniae (12.1%), Pseudomonas aeruginosa (5.3%), Proteus mirabilis (0.76%) and Trueperella pyogenes (0.76%) (Table 1, Fig.1). In studies conducted in Asia, Staphylococcus species have been reported to be the predominant etiological agents of mastitis in cattle and buffaloes capable of causing peracute, acute, subacute, chronic, gangrenous and subclinical types of mastitis (Sharma et al., 2012) [15]. Bhat et al. (2017) [3] reported *Staphylococcus aureus* (61%) as the most predominant bacteria isolated from the mastitic cattle of Jammu and Kashmir, followed by *E. coli* (13%), coagulase-negative staphylococci (13.04%), *Streptococcus uberis* (4.35%), and *Streptococcus dysgalactiae* (8.69%). The etiological differences in mastitis could be attributable to varied topographical and animal management conditions in addition to the usage of varied antimicrobials.

Of the 41 staphylococcal isolates subjected to antibiogram studies using 13 antimicrobial drugs, eight isolates were found to be resistant to all the drugs under study and were categorized as multidrug resistant (MDR). Most of the isolates were found to be sensitive to chloramphenicol (63.4%) followed by enrofloxacin (46.3%), cotrimoxazole (46.3%) and ceftriaxone (39%) (Table.2, Fig.2). Similar sensitivity pattern of staphylococci to chloramphenicol and ceftriaxone has been reported by many workers (Bhati *et al.*, 2013) [4]; Yadav *et al.* (2015b)) [16]. Staphylococci are potent pathogens and control is complicated by high antibiotic resistance (Livermore, 2000) [8] and the carriage of a variety of virulence determinants including several toxins and virulent factors which evade immune responses and biofilm formation (Zecconi and Scali, 2013) [17].

Most of the streptococcal isolates obtained in the present study were also found to be sensitive to chloramphenicol (57.1%) followed by amoxycillin with clavulinic acid (37.1%) and cefotaxime (25.7%) and five of them were resistant to all the drugs under study. However, this is in contrary to the findings of another antibiogram study on bovine mastitis, where the sensitivity pattern of Streptococcus isolates showed highest sensitivity for ceftiofur (68%), followed by enrofloxacin (50%), gentamicin (48%), tetaracycline (46%), ampicillin (40%), and ceftrioxone (40%) (Muley et al., 2018) [10]. None of the streptococcal isolates were sensitive to gentamycin and amikacin in the present study (Table 2), contrary to the findings of another study on bovine mastitis in Jordan where the S. agalactiae isolates have been reported to show highest susceptibility to gentamycin (40%) (Alekish et al., 2013) [1]. This variation in susceptibility patterns might be due varied environmental conditions, animal husbandry practices and use of different drugs in the treatment of mastitis in different geographical areas that can indeed put significant pressure on organisms resulting in evolution of drug-resistant organisms.

E. coli, the third predominant bacteria isolated in the present study, has been reported to be one of the most significant causes of clinical mastitis in dairy animals, affecting high producing cows as wells as cows in the early lactation period with low somatic cell counts and its endotoxin is potential health threat at consumer end (Liu et al., 2018) [7]. The antibiogram studies on Enterobacteriaceae in the present study revealed that 58.5% of the isolates were sensitive to chloramphenicol followed by sensitivity to both ceftriaxone and cotrimoxazole (39%) and three, resistant to all the drugs under study (Fig.2). Least sensitivity was reported to ampicillin cloxacillin followed by cefoxitin. Similar observations were made in a study on antimicrobial resistance of Enterobacteriaceae isolated from healthy, clinical and subclinical mastitis udders of cows (Alves et al., 2024) [2].

All the seven isolates of *Pseudomonas aeruginosa* in the preset study were found to be sensitive to only two to three antimicrobial drugs and were MDR exhibiting resistance to

at least three different classes of antimicrobials with most of them exhibiting sensitivity to enrofloxacin followed by ceftriaxone (Table.2). This bacterial pathogen is reported to be a potential environmental pathogen causing mastitis, frequently associated with wet bedding, water used in milking parlor, intramammary tubes used for the treatment and is extremely resistant to commonly used antimicrobials with the potential to form biofilms (Park *et al.*, 2014) [12].

with the potential to form biofilms (Park *et al.*, 2014) ^[12]. *T. pyogenes*, isolated from a case of mastitis in the present study (based on phenotypic characters) is a rare pathogen reported from cases of mastitis in India (Fig.3, Fig.4). It is a Gram-positive, catalase negative, facultatively anaerobic, fastidious, coryneform bacteria with enhanced growth on blood agar producing pinpoint, convex, slightly translucent, circular colonies surrounded by a zone of β -hemolysis (Quin *et al.*, 2011)) ^[13]. In the present study, the milk sample was obtained from a heifer with swelling of the udder, with the milk emanating a foul smell, akin to the clinical signs of summer mastitis caused by *T. pyogenes* as reported by Kibebew (2017) ^[6]. This bacterial isolate was found to be

resistant to all the antimicrobials used in the present study (Table 2). Antimicrobial resistance, including multidrug resistance among *T. pyogenes* isolates, has been reported by various researchers. Rezanejad *et al.* (2019) [14] reported that *T. pyogenes* bacteria isolated from the mastitic milk samples harbored the highest prevalence of resistance toward gentamicin (100%), penicillin (100%), ampicillin (90.62%), amoxicillin (87.50%), trimethoprim-sulfamethoxazole (87.50%), cefalexin (84.37%) and streptomycin (81.25%) antibiotic agents.

Most of the bacterial isolates in the present study exhibited highest susceptibility to chloramphenicol (56.8%) followed by enrofloxacin (36.36) and cotrimoxazole (33.3%) (Fig.2). This might be because of lesser usage of the drug which is usually used as last alternative in mastitis, because of the concerns about drug residues causing potential human health risks. Based on the findings of the present study, the use of enrofloxacin and cotrimoxazole is proposed in the study area for the treatment of bovine mastitis.

S. No Name of the bacteria		Species	No. of isolates		
1	Stanbulancai	S. aureus	36		
1	Staphylococci	CoNS (Non aureus staphylococci)	5		
2	Streptococci	Streptococcal spp	35		
		K. pneumoniae	16		
3	Enterobacteria	E. coli	24		
		P. mirabilis	1		
4	Pseudomonas	Pseudomonas aeruginosa	7		
5	Trueperella	Trueperella pyogenes	1		
5	Other Gram-positive bacilli		7		
	Total		132		

Table 1: Bacterial isolates obtained from the milk samples

Table 2: Phenotypic pattern of sensitivity of the bacterial isolates to the antimicrobial drugs under study

Species/family assigned (No of isolates)	Ceftriaxone	Cefotaxime	Ampicillin Cloxacillin	Amikacin	Cefoperazone	Cefoxitin	Chloramphenicol	Amoxycillin with clavulanic acid	Cotrimoxazole	Enrofloxacin	Gentamicin	Ceftrioxone with Tazobactum	oxytetracycline
Staphylococcus spp. (41)	16	14	9	15	13	13	26	14	19	19	12	10	4
Streptococcus spp. (35)	1	10	9	-	7	7	20	13	5	7	-	8	1
Enterobacteriaceae (41)		11	1	4	4	2	24	4	16	12	5	7	4
Pseudomonas aeruginosa (7)		1	-	2	-	-	1	1	-	5	1	1	-
Trueperella pyogenes (1)		-	-	-	-	-	-	-	-	-	-	-	-
Others (7)	1	2	2	3	2	4	4	2	4	5	4	3	-
Total (132)	38	38	21	24	26	26	75	34	44	48	22	29	9

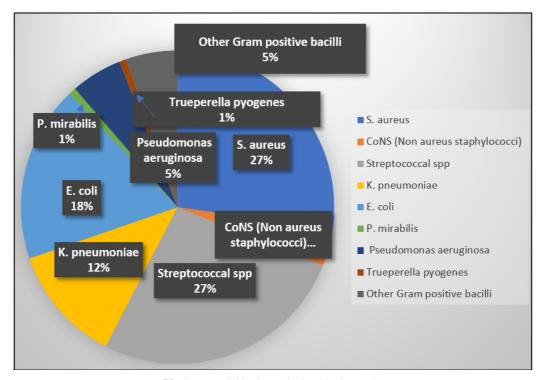


Fig 1: Bacterial isolates obtained in the study

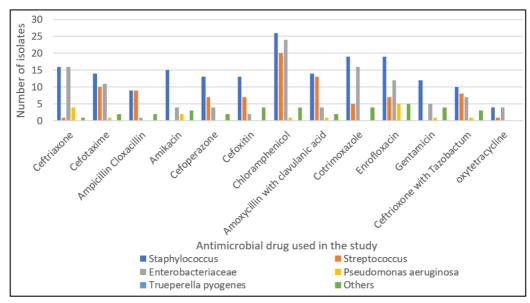


Fig 2: Phenotypic pattern of sensitivity of the bacterial isolates to the antimicrobial drugs under study

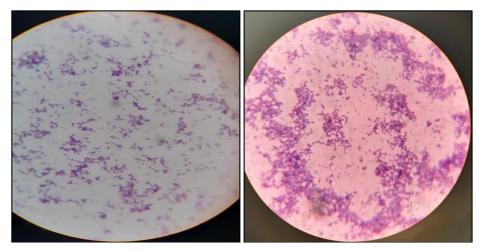


Fig 3: Trueperella pyogenes staining morphology-Gram's stain





Fig 4: Trueperella pyogenes colony morphology-complete haemolysis on blood agar

Conclusion

The study highlights the predominant bacterial species and the antibiogram of the isolates associated with bovine mastitis in the study area. The most predominant bacteria isolated were Staphylococcus aureus followed Streptococcus spp. and E. coli. Highest sensitivity was among observed all the bacterial isolates chloramphenicol, followed by enrofloxacin and cotrimoxazole. 17 bacterial isolates were found to be resistant to all the antimicrobial drugs used and several MDR bacteria were also reported. In the present study, though most of the bacterial isolates were found to be sensitive to chloramphenicol, due to the concerns about drug residues causing potential human health risks the use of drugs like enrofloxacin and cotrimoxazole is recommended for the treatment of bovine mastitis in the study area. In addition, for effective control of mastitis, continuous monitoring of mastitis, early detection, careful management and treatment of the mastitis infection which should be preceded with identification of the causative agent and susceptibility test. Such an initiative would prevent, the development of antimicrobial-resistant bacteria (AMR) with a potential to spread resistance genes to previously susceptible bacteria as AMR is a problem that affects animal, human and environmental health and should be evaluated within the one-health concept.

Acknowledgement

This work was supported by veterinary clinical complexes at veterinary colleges of Shivamogga and Hassan, KVAFSU, Bidar.

References

- Alekish MO, Al-Qudah KM, Al-Saleh A. Prevalence of antimicrobial resistance among bacterial pathogens isolated from bovine mastitis in northern Jordan. Rev Med Vet. 2013;164(6):319-326.
- Alves JS, de Moura Souza R, de Lima Moreira JP, Gonzalez AG. Antimicrobial resistance of Enterobacteriaceae and *Staphylococcus* spp. isolated from raw cow's milk from healthy, clinical and subclinical mastitis udders. Preventive Veterinary Medicine. 2024;227:106205.
- 3. Bhat AM, Soodan JS, Singh R, Dhobi IA, Hussain T, Dar MY, *et al.* Incidence of bovine clinical mastitis in Jammu region and antibiogram of isolated pathogens.

- Vet World. 2017;10(8):984-989. doi:10.14202/vetworld.2017.984-989.
- Bhati T, Kataria AK, Nathawat P, Sharma SK, Mohammed N, Mathur M. Antimicrobial susceptibility profiling of *Staphylococcus aureus* isolates from bovine subclinical mastitis. Veterinary Research. 2013;6(2):39-42.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing: twenty-first informational supplement. CLSI document M100-S23. Wayne, PA: CLSI; 2013. Vol. 33(1).
- 6. Kibebew K. Bovine mastitis: A review of causes and epidemiological point of view. Journal of Biology, Agriculture and Healthcare. 2017;7(2):1-4.
- 7. Liu G, Ding L, Han B, Piepers S, Naqvi SA, Barkema HW, *et al.* Characteristics of *Escherichia coli* isolated from bovine mastitis exposed to subminimum inhibitory concentrations of cefalotin or ceftazidime. BioMed Research International. 2018;2018:4301628.
- 8. Livermore DM. Antibiotic resistance in staphylococci. International Journal of Antimicrobial Agents. 2000:16:3-10.
- 9. Miller GY, Bartlett PC, Lance SE, Anderson J, Heider LE. Costs of clinical mastitis and mastitis prevention in dairy herds. Journal of the American Veterinary Medical Association. 1993;202(8):1230-1236.
- Muley V, Dhanalakshmi K, Reddy YN. Antibiogram of bacterial isolates obtained from milk samples in and around Hyderabad, India. International Journal of Current Microbiology and Applied Sciences. 2018;7(3):3720-3724.
- 11. National Academy of Agricultural Sciences (NAAS). Mastitis management in dairy animals. Policy Paper 61. New Delhi: NAAS; 2013. p. 1-12.
- 12. Park H, Hong M, Hwang S, Park Y, Kwon K, Yoon J, *et al.* Characterisation of *Pseudomonas aeruginosa* related to bovine mastitis. Acta Veterinaria Hungarica. 2014;62(1):1-12.
- 13. Quinn PJ, Markey BK, Leonard FC, Hartigan P, Fanning S, Fitzpatrick E. Veterinary microbiology and microbial disease. 2nd ed. Chichester: John Wiley & Sons; 2011.
- 14. Rezanejad M, Karimi S, Momtaz H. Phenotypic and molecular characterization of antimicrobial resistance in *Trueperella pyogenes* strains isolated from bovine

- mastitis and metritis. BMC Microbiology. 2019;19(1):305.
- 15. Sharma N, Rho GJ, Hong YH, Kang TY, Lee HK, Hur TY, Jeong DK. Bovine mastitis: an Asian perspective. Asian Journal of Animal and Veterinary Advances. 2012;7(6):454-476.
- 16. Yadav R, Sharma SK, Yadav J, Choudhary S, Kataria AK. Profiling of antibiotic resistance of *Staphylococcus aureus* obtained from mastitic milk of cattle and buffalo. Journal of Pure and Applied Microbiology. 2015;9(2):1539-1544.
- 17. Zecconi A, Scali F. *Staphylococcus aureus* virulence factors in evasion from innate immune defenses in human and animal diseases. Immunology Letters. 2013;150(1-2):12-22.