

## International Journal of Advanced Biochemistry Research



ISSN Print: 2617-4693  
 ISSN Online: 2617-4707  
 IJABR 2024; SP-8(8): 16-20  
[www.biochemjournal.com](http://www.biochemjournal.com)  
 Received: 19-05-2024  
 Accepted: 29-06-2024

**Indrapreet Singh Saini**  
 Department of Genetics and  
 Plant Breeding, SOAG, ITM  
 University, Gwalior,  
 Madhya Pradesh, India

**Soni Singh**  
 Department of Genetics and  
 Plant Breeding, SOAG, ITM  
 University, Gwalior,  
 Madhya Pradesh, India

**Akash Barela**  
 Department of Genetics and  
 Plant Breeding, SOAG, ITM  
 University, Gwalior,  
 Madhya Pradesh, India

**Vyenkatesh Charke**  
 Department of Genetics and  
 Plant Breeding, SOAG, ITM  
 University, Gwalior,  
 Madhya Pradesh, India

**Shailendra Sagar Prajapati**  
 Guest Faculty, Department of  
 Plant Breeding and Genetics,  
 Jawaharlal Nehru Krishi  
 Vishwa Vidyalaya, Jabalpur,  
 Madhya Pradesh, India

**Corresponding Author:**  
**Indrapreet Singh Saini**  
 Department of Genetics and  
 Plant Breeding, SOAG, ITM  
 University, Gwalior,  
 Madhya Pradesh, India

## Studies on chlorophyll spectrum and mutation effectiveness in M<sub>2</sub> generation of Sesame (*Sesamum indicum* L.)

**Indrapreet Singh Saini, Soni Singh, Akash Barela, Vyenkatesh Charke and Shailendra Sagar Prajapati**

DOI: <https://doi.org/10.33545/26174693.2024.v8.i8Sa.1692>

### Abstract

Ten desirable macro mutants in TKG-55 sesame (*Sesamum indicum* L.), resulting from induced NaN<sub>3</sub> and combination treatment with EMS chemical mutagenesis, have been identified for: viridis (seedling color as marker, increased seed protein content), Albino (White), Xantha (Yellow), Chlorina (Light green), Albomaculata (White dots on green leaves), Alboviridis (Initially white and later becomes normal plants), Xanthaviridis (Initially yellow and later become normal plants). All mutant traits were recessive compared to normal, with viridis showing digenic inheritance and the others monogenic. Along with macro mutants were initially observed for germination, survival and lethality percentage respectively.

**Keywords:** *Sesamum indicum*, desirable macro mutants, chemical mutagenesis

### Introduction

Sesame (*Sesamum indicum* L.) is considered to be an ancient oil seed crop (Brar and Ahuja, 1979) [31]. According to Oplinger *et al.*, (1990) [7], it is an important oilseed to generate high quality edible oil and protein for low income peasants of major sesame growing countries including Sudan, Ethiopia, Uganda, Nigeria, Mexico, Venezuela, India, China, Pakistan, Turkey and Myanmar. Sesame is a diploid species ( $2n = 2x = 26$ ), with a basic chromosome number of  $X = 8$  or  $13$ , while some are tetraploids and octaploids (Kobayashi 1991) [18], and it belongs to the Pedaliaceae family. It is one of the most ancient oil seeds known to mankind and plays a major role in human nutrition (komivi *et al.*, 2018) [19].

Mutations are the tools at hand exploited by the geneticist to study the nature and function of genes which are the basis of plant growth and development, hence producing raw materials for genetic improvement of economic crops (Adamu *et al.*, 2007) [1]. Mutation breeding is one possible alternative to conventional breeding for crop improvement. Induced mutations can rapidly create variability in quantitatively and qualitatively inherited traits in crops (Maluszynski *et al.*, 1995; Muduli and Mishra, 2007) [21, 24]. Mutagenesis has been successfully used to induce genetic variability in many crops, allowing to isolate mutants with desirable characters such as increased seed yield, earliness (Wongyai *et al.*, 2001) [32], modified plant architecture, closed capsules, disease resistance, seed retention, larger seed size, desirable seed colour and high oil content. Chemical mutagens are agents that change the genetic materials, usually the DNA of an organism by increasing the frequency of mutations above the natural background level. Mutagens have played a pivotal role in creating crop varieties with induced mutations resulting in significant increases in food production. Sodium azide is a powerful mutagen in plant and its efficiency depend on concentration and treatment duration (Al-Qurainy and Khan, 2009) [16]. It has been successfully used in the improvement of rice, barley and oats, (Awan *et al.*, 2011) [4].

### Materials and Methods

Chemical mutagenic treatment hundred well filled seeds of TKG-55 variety were pre-soaked for four hours in distilled water. Seeds were treated with different concentrations of NaN<sub>3</sub> & EMS in double distilled water and pH of the mutagenic solution was adjusted to seven by

using phosphate buffer. The pre-soaked seeds after removal from the water were packed between the folds of blotting paper to remove excess water adhering to the surface. The seeds were immersed for three hours in the required concentrations of the mutagen with intermittent shaking. To ensure uniform absorption of the mutagen, the volume of the mutagen solution was maintained at a proportion of ten times to that of seed volume. The whole treatment was carried out at room temperature. After the treatment, the seeds were thoroughly washed with tap water for ten times. Non-treated dry seeds and pre-soaked seeds in distilled water for seven hours were used as control.

The M<sub>2</sub> generation study was conducted by resorting to methods suggested by Ganguli (1991)<sup>[11]</sup>. The normal good looking plants based on base population randomly selected in each treatment in the M<sub>1</sub> generation were advanced to M<sub>2</sub> generation during Aug - Nov 2023. They were sown in family rows in different blocks with a spacing of 45 cm between rows and 30 cm between plants to study the micro and macro mutations.

The chlorophyll mutants were classified as per the system proposed by Gustafsson (1940)<sup>[14]</sup> and Blixt and Gottscalk (1975)<sup>[6]</sup>. The chlorophyll mutants include the following. Mutant Described as viridis (seedling color as marker, increased seed protein content), Albino (White), Xantha (Yellow), Chlorina (Light green), Albomaculata (White dots on green leaves), Alboviridis (Initially white and later becomes normal plants), Xanthaviridis (Initially yellow and later become normal plants). The colour of the first formed leaf was taken for scoring the chlorophyll mutants. To compute the spectrum (relative percentage of different types) of mutants, the different types of seedling mutants for chlorophyll deficiency were scored separately from seventh to fifteenth day. The data were analysed for statistical parameters such as mutant effectiveness, mutation frequency for individual mutants as well as for different treatments respectively.

## Results and Discussion

The seed germination percentage was reduced more under higher dose of combination treatments than NaN<sub>3</sub> mutagen treatments. Maximum reduction was recorded at 0.4+0.4% dose of NaN<sub>3</sub> + E.M.S. treatments (58.82 per cent reduction over control) and minimum of 0.1+0.1 per cent dosage in NaN<sub>3</sub> + E.M.S. (84 per cent reduction over control). Lethality was observed ranging minimum at 0.1% dose of NaN<sub>3</sub> (causes 17% lethality) to maximum at 0.3+0.3% combination dose of NaN<sub>3</sub> + E.M.S. (causes 47% lethality). This finding was in agreement with the report of Ganesh Kumar *et al.* (2001)<sup>[10]</sup> in sesame. The decrease in germination due to mutagenic treatment observed in the present study was also in conformity with the earlier reports of Anitha Vasline (1998)<sup>[3]</sup>, Radhakrishnan *et al.* (2001)<sup>[26]</sup> in sesame; Jegadeeswaran (1989)<sup>[15]</sup> in groundnut; Shamsi (1981)<sup>[30]</sup> in sunflower; Ahmed John (1996)<sup>[2]</sup>, Barela *et al.*, (2022)<sup>[5]</sup> in green pea and Deepalakshmi (2000)<sup>[9]</sup> in black gram (table 1, fig 1 & 2).

Seedling survival during early phase, the seedlings could adjust or repair themselves to eliminate the dead or unwanted cells. On the other hand, some of the seedlings were not able to overcome the radiation damage and hence, they died before they put forth any side effects. This might be due to inhibition of auxin synthesis, lack of assimilatory mechanism, inhibition of mitosis and chromosomal

damages. An inverse relationship between the dosage of mutagen and survival was observed in both cases. The survival reduction ranged from 53% in NaN<sub>3</sub> + E.M.S. (0.3+0.3%) to 83 per cent in NaN<sub>3</sub> (0.1) in chemical mutagens. Many workers viz., Cagirgan (1996)<sup>[8]</sup>, Govindarasu *et al.* (1998)<sup>[13]</sup>, Sasikala and Kamala (1988)<sup>[29]</sup>, Anitha Vasline (1998)<sup>[3]</sup> and Khin Mar Mar New (2001)<sup>[17]</sup> have reported a dose dependent decrease in plant survival in sesame.

## Effect of Mutagenic Treatments in M<sub>2</sub> Generation Frequency of chlorophyll mutants

The number of plants segregated for chlorophyll deficiency on the basis of M<sub>1</sub> plants and M<sub>2</sub> seedlings were computed and furnished in Table 2 (TKG 55). The chlorophyll mutants occurred in all the treatments, the frequency of chlorophyll mutation varied from 0.49 (NaN<sub>3</sub> 0.1%) to 2.11 per cent (NaN<sub>3</sub> 0.3%) on M<sub>2</sub> plant basis and from 2% (NaN<sub>3</sub> 0.4%) to 4% per cent (NaN<sub>3</sub> 0.3%) on M<sub>1</sub> seedling basis. In combination treatments, the frequency ranged from 0.93% (NaN<sub>3</sub> + E.M.S. (0.2+0.2%)) to 5 per cent (NaN<sub>3</sub> + E.M.S. (0.3+0.3%)) on M<sub>2</sub> plant basis and 2% (NaN<sub>3</sub> + E.M.S. (0.2+0.2%)) dose to 4% per cent (NaN<sub>3</sub> + E.M.S. (0.2+0.2%)) on M<sub>1</sub> seedling basis. The treatment (NaN<sub>3</sub> + E.M.S. (0.3+0.3%)) exhibited the maximum frequency of chlorophyll mutation for both M<sub>1</sub> plant basis (4%) and M<sub>2</sub> seedling basis (5%) (Table 2). The frequency of chlorophyll mutations, in general, were low in this crop thus it may be attributed to the fact that oil seed crops are resistant to induced chlorophyll mutations as reported by Rajan (1969)<sup>[27]</sup> and Rangaswamy (1982)<sup>[28]</sup>.

## Spectrum of chlorophyll mutation

The spectrum of chlorophyll mutations and the relative frequencies of different types of chlorophyll mutants are presented in Table 2 for TGM 55. Seven types of chlorophyll mutants viz., Albina, Xantha, Chlorotica, Viridis, Variegata, Marginata and Terminalis were observed in M<sub>2</sub> seedlings. Maximum number of chlorophyll were observed in xantha mutants except in NaN<sub>3</sub> @ 0.3% while albina and terminalis are intermediate in its occurrence. EMS & NaN<sub>3</sub> exposed treatments showed the maximum xantha albina mutants. The frequency ranged from 0.04 per cent of viridis to 0.13 per cent of xantha (Table 2). The presence of occurrence of chlorophyll mutants was higher in individual treatments (from 2 per cent in NaN<sub>3</sub> @ 0.1% to 4.0 per cent in NaN<sub>3</sub> @ 0.3% while compared to combination treatments NaN<sub>3</sub> & EMS (from NaN<sub>3</sub> + E.M.S. (0.3+0.3%) and NaN<sub>3</sub> + E.M.S. (0.4+0.4%). The order of frequency of mutants occurrence was chlorina < Vario-maculata < Auera < Viridis < Marginata < Albina < Chlorotica < Variegata < Terminalis < Xantha on over all basis. The reason for the appearance of greater number of Xantha viridis type may be attributed to involvement of polygenes in chlorophyll formation. According to the greater efficiency of low dose of mutagens appeared in relation to the fact that lethality and injury increased with increase in dose at faster rate than the useful mutations (table 2, Fig. 3).

## Chlorophyll mutants Efficiency

The efficiency was calculated on lethality, injury and sterility basis. In TGM 55, the efficiency on lethality basis showed a declining trend as dose increase up to 0.1% in NaN<sub>3</sub> treated dose and the maximum value obtained on

lethality basis was 34 per cent in  $\text{NaN}_3$  @ 0.4% (Table1). The seedling injury causes lethality varied from 3%  $\text{NaN}_3$  @ 0.1% to 7.8% per cent  $\text{NaN}_3$  @ 0.4% for sodium azide and from 3.4% ( $\text{NaN}_3$  + E.M.S. (0.2+0.2)) to 11.23 per cent

( $\text{NaN}_3$  + E.M.S. (0.3+0.3)) for combination treatment of  $\text{NaN}_3$  & EMS in TGM 55. Among all chemical treatments, the treatment  $\text{NaN}_3$  @ 0.4 with the value of 1.03 in TGM 55 was found to be efficient. (table 1, fig.1&2).

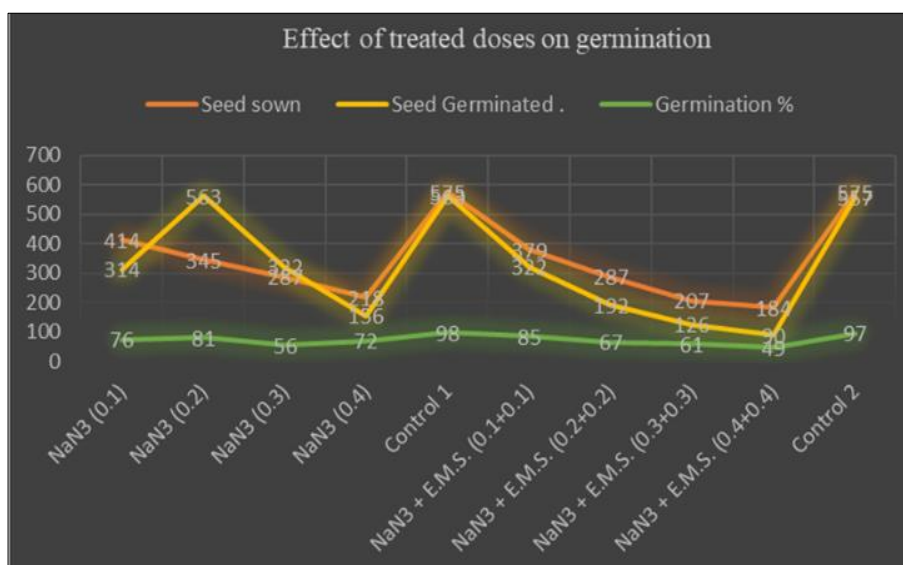
**Table 1:** Effect of mutagens on single plant yield in  $M_2$  generation

S. No.	Treated Dose (in%)	Mutant population	Germination (%)	Lethality (%)	Injury (%)	Survival (%)
1	$\text{NaN}_3$ (0.1)	333	74	17	3	83
2	$\text{NaN}_3$ (0.2)	243	79	30	4.7	70
3	$\text{NaN}_3$ (0.3)	234	54	25	5	75
4	$\text{NaN}_3$ (0.4)	153	70	34	3.4	66
5	Control 1	450	98	0	0	100
6	$\text{NaN}_3$ + E.M.S. (0.1+0.1)	279	84	32	2.8	68
7	$\text{NaN}_3$ + E.M.S. (0.2+0.2)	207	65	20	3.4	80
8	$\text{NaN}_3$ + E.M.S. (0.3+0.3)	126	59	47	11.23	53
9	$\text{NaN}_3$ + E.M.S. (0.4+0.4)	207	52	35	8.3	65
10	Control 2	450	95	0	0	100

**Table 2:** Chlorophyll mutation spectrum in  $M_2$  generation

S. No.	Mutant Characters type	Treated dose (in%)										Total Mutant	$M_2$ pop.	Mutation Frequency	
		T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	T <sub>8</sub>	T <sub>9</sub>	T <sub>10</sub>			$M_1$ Plant basis	$M_2$ Plant basis
1	Albina	0	0	0	0	0	1	0	0	1	0	2	205	4	0.08
2	Xantha	0	1	0	1	0	0	0	1	0	0	3	134	6	0.13
3	Chlorina	0	0	0	0	0	0	0	0	0	0	0	95	0	0
4	Chlorotica	1	0	0	0	0	0	0	1	0	0	2	71	4	0.08
5	Viridis	0	0	0	0	0	0	1	0	0	0	1	441	2	0.04
6	Auera	0	0	0	0	0	0	0	0	0	0	0	160	0	0
7	Variegata	0	0	2	0	0	0	0	0	0	0	2	108	4	0.08
8	Marginata	0	0	0	0	0	0	0	0	1	0	1	40	2	0.04
9	Vario-maculata	0	0	0	0	0	0	0	0	0	0	0	70	0	0
10	Terminalis	0	1	0	0	0	1	0	0	0	0	2	427	4	0.08
	Total	1	2	2	1	0	2	1	2	2	0	13	1751	26	0.56
	$M_2$ population	205	134	95	71	441	160	108	40	70	427	1751			
Mutation Frequency	$M_1$ Plant basis	2	4	4	2	0	4	2	4	4	0	26			
	$M_2$ Plant basis	0.49	1.49	2.11	1.41	0.00	1.25	0.93	5.00	2.86	0.00	0.74			

Here, T<sub>1</sub>- ( $\text{NaN}_3$  (0.1)), T<sub>2</sub>-  $\text{NaN}_3$  (0.2), T<sub>3</sub>-  $\text{NaN}_3$  (0.3), T<sub>4</sub> -  $\text{NaN}_3$  (0.4), T<sub>5</sub>- Control 1, T<sub>6</sub>-  $\text{NaN}_3$  + E.M.S. (0.1+0.1), T<sub>7</sub>-  $\text{NaN}_3$  + E.M.S. (0.2+0.2), T<sub>8</sub>-  $\text{NaN}_3$  + E.M.S. (0.3+0.3), T<sub>9</sub>-  $\text{NaN}_3$  + E.M.S. (0.4+0.4), T<sub>10</sub>- Control 2



**Fig 1:** Effect of treated doses on seed germination

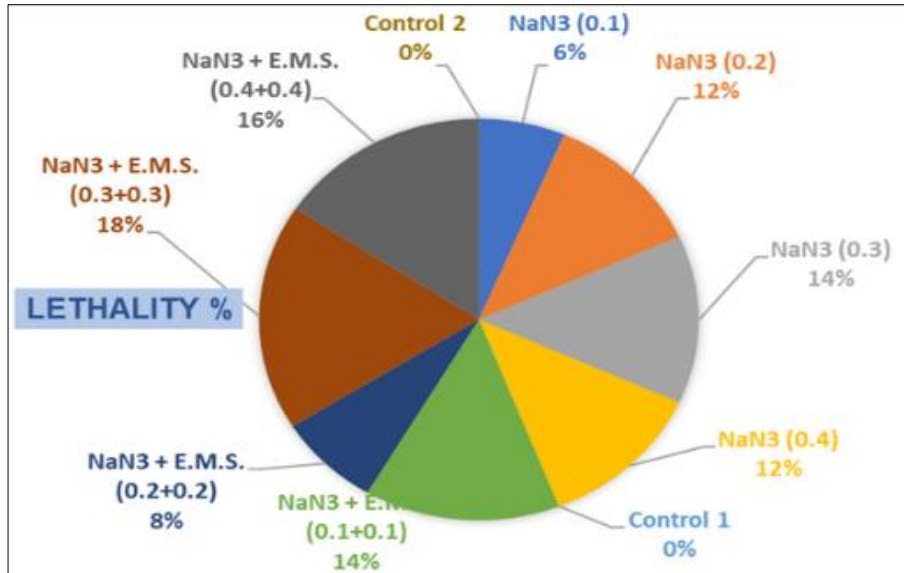


Fig 2: Effect of treated doses on population lethality

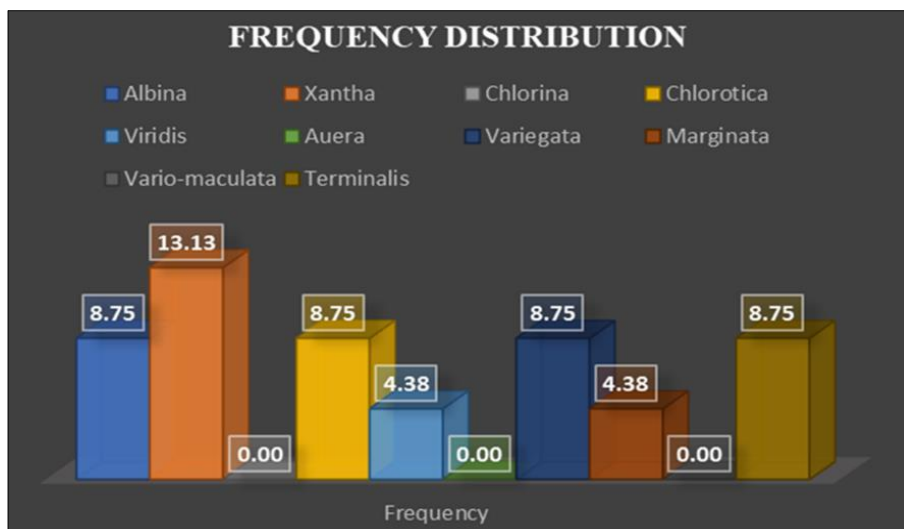


Fig 3: Spectrum of chlorophyll mutation in M<sub>2</sub> generation

**Conclusion**

In M<sub>2</sub> generation, the frequency of chlorophyll and viable mutants were calculated as percentage of M<sub>1</sub> plant and M<sub>2</sub> seedling basis. No clear trend could be observed for number of chlorophyll mutants in NaN<sub>3</sub> and combination treatment of NaN<sub>3</sub> & EMS in TGM 55. Among the treatments, TGM 55 registered more frequency than rest of the treatments. The spectrum of chlorophyll mutants comprised of xantha, albina and viridis. NaN<sub>3</sub> exhibited maximum number of xantha mutants except in NaN<sub>3</sub> @0.1% whereas combination treatment of NaN<sub>3</sub> & EMS exposed treatments showed the maximum marginata mutants. The proportion of one type of mutant was more than those segregating for either two or three types. Combination dose were found to be more efficient and effective than NaN<sub>3</sub> for chlorophyll mutants. The effectiveness and efficiency of both the mutagens were more in TGM 55 treatment dose.

**References**

1. Adamu AK, Aliyu H. Morphological effects of sodium azide on tomato (*Lycopersicon esculentum* Mill). *Sci World J.* 2007;2(4):1-6.
2. Ahmed JS. Impact of gamma irradiation on survival of blackgram. *Madras Agric J.* 1996;83(9):592-593.

3. Anitha Vasline Y. Studies on induced mutagenesis in *Sesamum indicum* L. Ph.D. Thesis, Annamalai University; c1998.
4. Awan H, Awan I, Mansoor M, Khan EA, Khan MA. Effect of sowing time and plant spacing on fiber quality and seed cotton yield. *Sarhad J Agric.* 2011;27(3):411-413.
5. Barela A, Jain S, Khandalkar VS, Singh P, Rahangdale S, Behra K, *et al.* Ameliorations of gamma radiations on chlorophyll spectrum and morphological characteristics of peas. *Int J Environ Climate Change.* 2022;12(12):1724-1731.
6. Blixt S, Gottschalk W. Mutation in the leguminosae. *Agric Hort Genet.* 1975;33:33-85.
7. Buhler DD, Oplinger ES. Influence of tillage systems on annual weed densities and control in solid-seeded soybean (*Glycine max*). *Weed Sci.* 1990;38(2):158-165.
8. Cagirgan MI. Radiosensitivity of Turkish sesame cultivars to gamma rays. *Turk J Field Crops.* 1996;1(2):39-43.
9. Deepalakshmi AJ. Mutagenesis in blackgram (*Vigna mungo* (L.) Hepper). M.Sc. Thesis, TNAU, Coimbatore; c2000.

10. Ganesh Kumar N, Kannan K, Ganesan J. Response of sesame seed coat colour to induced chemical mutation. Natl Seminar on Sesame Crop Improvement and Future Prospects, 28-1 February, Dept Agrl Bot, Annamalai University; c2001. p. 18.
11. Ganguli PK. Induced mutation as a method. In: Crop Improv Adv Plant Breed. 1991;1:228-240.
12. Gaul H. Mutagens effects in first generation after seed treatment: plant injury, lethality, cytological effects, sterility. In: Manual on Mutation Breeding. Tech Plant Ser No. 119. IAEA, Vienna; c1977. p. 87-88.
13. Govindarasu R, Natarajan M. Transgressive variation following irradiation of heterozygous genotype in sesame for morphological and agronomic traits. J Nucl Agric Biol. 1998;27(3):172-180.
14. Gustafsson A. The mutation system of the chlorophyll apparatus. Lunds Univ Arsskv. 1940;36:1-40.
15. Jegadeeswaran G. Studies on induced mutagenesis in groundnut. M.Sc. Thesis, TNAU, Coimbatore; c1989.
16. Khan S, Al-Qurainy F, Anwar F. Sodium azide: a chemical mutagen for enhancement of agronomic traits of crop plants. Environ We Int J Sci Tech. 2009;4:1-21.
17. Khin Mark Mar New, Ganesh Kumar N, Ganesan J. Induced mutagenesis on seedlings characters in sesame (*Sesamum indicum* L.). Natl Seminar on Sesame Crop Improvement and Future Prospects, 28-1 February, Dept Agrl Bot, Annamalai University; c2001. p. 18.
18. Kobayashi K. On generalized gamma functions occurring in diffraction theory. J Phys Soc Jpn. 1991;60(5):1501-1512.
19. Komivi D, Marie AM, Rong Z, Qi Z, Mei Y, Ndiaga C, et al. The contrasting response to drought and waterlogging is underpinned by divergent DNA methylation programs associated with transcript accumulation in sesame. Plant Sci. 2018;277:207-217.
20. Konzak CF, Nilan RA, Wagner J, Faster RJ. Efficient chemical mutagenesis: the use of induced mutations in plant breeding. Radiat Bot. 1965;5:49-70.
21. Maluszynski M, Ahloowalia BS, Sigurbjörnsson B. Application of in vivo and in vitro mutation techniques for crop improvement. Euphytica. 1995;85:303-315.
22. Mensah JK, Obadoni BO, Akomeah PA, Ikhajiagbe, Ajibolu. The effect of sodium azide and colchicine treatments on morphological and yield traits of sesame seed (*Sesamum indicum* L.). Afr J Biotechnol. 2007;6(5):534-538.
23. Mohammed R. Removal of fluoride from Shatt al-Arab drinking water using a novel low cost material. Kufa J Eng. 2018;9(3):1-16.
24. Muduli KC, Misra RC. Efficacy of mutagenic treatments in producing useful mutants in finger millet (*Eleusine coracana* Gaertn.). Indian J Genet Plant Breed. 2007;67(3):232-237.
25. Prabakaran AJ. Identification of male sterile sources through wide hybridization and induced mutagenesis in sesame (*Sesamum indicum* L.). Ph.D. Thesis, TNAU, Coimbatore; c1992.
26. Radhakrishnan V, Prabhu S, Sivaji, Gogineni RS, Suresh Y, Anbuselvam Y, Ganesan J. Effect of DES as mutagen on germination and seedling behaviour in sesame (*Sesamum indicum* L.). Natl Seminar on Sesame Crop Improvement and Future Prospects, 28-1 February, Dept Agrl Bot, Annamalai University; c2001. p. 19.
27. Rajan SS. Relative biological effectiveness of monoenergetic fast neutrons on oil seeds. Proc Sym Radiations and Radioactive Substances in Mutation Breeding. FAO Dept Atom Ener, Govt India; c1969. p. 79-98.
28. Rangaswamy M, Rathinam M. Mutagen induced male sterile lines in *Sesamum indicum*. Indian J Genet. 1982;42(2):142-143.
29. Sasikala S, Kamala T. Mutagenic effectiveness and efficiency of gamma rays on four gingelly cultivars. Indian J Bot. 1988;11(2):118-122.
30. Shamsi SRA, Rochi R, Nisa A. Radiosensitivity studies in oil seed crops. I. Sunflower (*Helianthus annuus* L.). Biologia, Pakistan. 1981;27(1):115-125.
31. Suri SK, Brar AS, Ahuja LD. Adsorption from binary solutions of benzene and cyclohexane on cobalt-chalcogenide surfaces. J Colloid Interface Sci. 1979;69(2):347-351.
32. Wongyai W, Saengkaewsook W, Veerawudh J. Sesame mutation induction: improvement of non-shattering capsule by using gamma rays and EMS. [No journal information provided; ensure to add if available].