

## International Journal of Advanced Biochemistry Research



ISSN Print: 2617-4693  
 ISSN Online: 2617-4707  
 IJABR 2024; 8(8): 987-992  
[www.biochemjournal.com](http://www.biochemjournal.com)  
 Received: 10-06-2024  
 Accepted: 14-07-2024

**Sakshi Tomar**  
 Department of Silviculture and  
 Agroforestry, COF, Navsari  
 Agricultural University,  
 Gujarat, India

**Prabhat Tiwari**  
 College of Horticulture and  
 Forestry, RLBCAU, Jhansi,  
 Uttar Pradesh, India

**Indu Kale**  
 Department of Forest Biology  
 and Tree Improvement, COF,  
 Navsari Agricultural  
 University, Gujarat, India

**Ashok Kumar**  
 Department of Silviculture and  
 Agroforestry, Dr. YSPUHF,  
 Solan, Himachal Pradesh,  
 India

**Pooja**  
 Department of Plant  
 Pathology, CCSHAU, Hisar,  
 Haryana, India

**Sumit**  
 Department of Forestry,  
 CCSHAU, Hisar, Haryana,  
 India

**Corresponding Author:**  
**Indu Kale**  
 Department of Forest Biology  
 and Tree Improvement, COF,  
 Navsari Agricultural  
 University, Gujarat, India

## Revitalizing Tulsi growth: Impact of pre-germination treatments on seed quality and seedling performance in Uttar Pradesh's semi-arid zones

**Sakshi Tomar, Prabhat Tiwari, Indu Kale, Ashok Kumar, Pooja and Sumit**

DOI: <https://doi.org/10.33545/26174693.2024.v8.i8m.1912>

### Abstract

Tulsi revered across cultures for its medicinal, culinary, and spiritual significance, is a plant of considerable importance. This study, conducted in 2020 at the RLBCAU nursery in Jhansi, Uttar Pradesh, sought to enhance the germination process and seed quality of Tulsi through various pre-germination treatments. The treatments tested included gibberellic acid (GA<sub>3</sub>) at 0.05% and 0.1%, potassium nitrate (KNO<sub>3</sub>) at 0.05% and 0.1%, indole-3-butyric acid (IBA) at 0.05% and 0.1%, hot water treatment, and an untreated control. Among these, the application of 0.1% GA<sub>3</sub> yielded the highest germination percentage at 32.1%, along with a significantly enhanced seedling vigour index of 20.21. The study underscores the substantial impact of pre-germination treatments on seed germination rates and seedling traits. Seeds treated with 0.1% GA<sub>3</sub> demonstrated notable improvements in both germination rate and overall seedling quality, highlighting the effectiveness of GA<sub>3</sub> in promoting robust seedling development. These findings suggest that GA<sub>3</sub> treatment at 0.1% concentration is particularly beneficial for improving the germination and early growth of Tulsi seedlings, offering a promising approach for optimizing cultivation practices in semi-arid regions like Jhansi.

**Keywords:** Dormancy, germination, *Ocimum tenuiflorum*, GA<sub>3</sub>, KNO<sub>3</sub>, IBA, seedling vigour index

### Introduction

Tulsi (*Ocimum tenuiflorum* L.), an aromatic shrub from the basil family *Lamiaceae*, is believed to have originated in north-central India. Highly revered in Ayurveda, Tulsi is often referred to as "The Incomparable One," "Mother Medicine of Nature," and "The Queen of Herbs." It is celebrated as an "elixir of life" for its profound medicinal and spiritual significance. Beyond its extensive therapeutic properties, Tulsi plays a crucial role in spiritual and cultural practices, making it a cherished herb in both health and religious contexts. The essential oil extracted from Tulsi is highly valued in various industries, including food, pharmaceuticals, cosmetics, and aromatherapy, owing to its distinctive flavor, aroma, and recognized antimicrobial and insecticidal properties.

The rising demand in the pharmacological industry and the need for efficient cultivation of medicinal plants has spurred significant interest in the study of seed germination for species like Tulsi. High seed quality and successful seedling establishment are essential for sustainable and profitable crop production. However, seed dormancy—defined as the failure of an intact, viable seed to complete germination under favorable conditions—often poses a significant challenge. Environmental factors such as light, temperature, and seed storage duration influence dormancy. To overcome this barrier, various methods, including scarification, pre-treatment with plant growth regulators (PGRs), and temperature shocks, are employed depending on the species and type of dormancy.

*Ocimum tenuiflorum* is known for its high yields of raw materials and active ingredients, including total phenols, flavones, chlorophyll, and L-ascorbic acid. In Western Europe, herb growers have extensively used plant growth regulators for over a decade. These regulators promote metabolic processes; enhance seed and fruit production, and increase resistance to fungal and bacterial diseases. PGRs are commonly employed to improve seed germination capacity, increase biomass yield, and enhance resistance to diseases and adverse growth conditions.

Gibberellins (GAs), a type of PGR, are naturally synthesized by seeds and play a crucial role in germination by hydrolyzing storage nutrients and directly influencing embryo growth. The external application of PGRs to seeds has been shown to break seed dormancy and enhance seedling establishment in many aromatic and medicinal plants.

Among the various PGRs, Gibberellic Acid (GA<sub>3</sub>) is particularly notable for its broad physiological effects. GA<sub>3</sub> has been shown to positively influence various aspects of plant life, including promoting plant vigour and stimulating the biosynthesis of secondary metabolites. Its application can lead to improved growth, increased biomass, and enhanced production of compounds essential for plant health and agricultural productivity.

This study focused on investigating the seed germination and seedling growth of the medicinal plant species *Ocimum tenuiflorum* L., given the limited research on the seed germination of this species using PGRs and inorganic pre-treatments. To address this gap, we explored various pre-sowing treatments, including different concentrations of GA<sub>3</sub>, IBA, KNO<sub>3</sub>, and hot water treatments, to enhance seed germination and seedling growth in this medicinally significant plant species. The findings from this research are expected to provide valuable insights into optimizing the cultivation practices of Tulsi, ensuring improved germination rates and healthier seedlings, which are crucial for meeting the growing demand for this revered herb across various industries.

## Materials and Methods

### Seed Collection and Surface Sterilization

The experiment was conducted in 2020 at the nursery of Rani Lakshmi Bai Central Agricultural University, Jhansi, Uttar Pradesh. Seeds were collected from the nursery of Dr. Y.S. Parmar University of Horticulture and Forestry, Nauni,

Solan district, Himachal Pradesh. The seeds underwent surface sterilization by immersing them in 70% ethanol for 1 minute, followed by a 10-minute treatment with 5% hypochlorite. Afterward, the seeds were thoroughly rinsed with sterile distilled water to eliminate any contaminants. The sterilized seeds were then stored for 24 hours before commencing the experimental procedures. Evaluation showed that the seeds averaged 100,000 per kilogram, with the weight of the sown seeds ranging between 50 and 60 grams.

### Treatment for Seed Germination

The seeds were carefully selected using the sinker and floater method, as described by Mandal (2000) [41]. In this process, seeds that sank in water were chosen for the experiment, as they are typically more viable. The study aimed to evaluate the effects of different pre-sowing treatments on Tulsi seed germination, specifically using Gibberellic Acid (GA<sub>3</sub>), Indole-3-butyric Acid (IBA), and Potassium nitrate (KNO<sub>3</sub>) at concentrations of 0.1%, 0.05%, as well as hot water soaking. Uniform-sized seeds were immersed in these plant growth regulator (PGR) solutions for 24 hours to assess their impact on germination. Distilled water was used as the control treatment to provide a baseline for comparison.

### Observation of Seed Germination

Seed germination was monitored daily for 30 days. Germination was defined as the point when the tip of the radicle had emerged from the seed coat. Germination parameters like Germination percentage were recorded as per Esehie (1994) [13]. The following observations viz. the imbibition period, speed of germination, mean daily germination (MDG), and the peak value of germination (PV) and germination value (GV) were also made as suggested by Czabator (1962) [10].

**Table 1:** Different pre-sowing treatments of *Ocimum tenuiflorum* L

Treatment code	Treatment	Duration and Concentration of pre-sowing parameter
T <sub>1</sub>	GA <sub>3</sub>	0.1% for 45 min
T <sub>2</sub>	GA <sub>3</sub>	0.05% for 45 min
T <sub>3</sub>	KNO <sub>3</sub>	0.1% for 45 min
T <sub>4</sub>	KNO <sub>3</sub>	0.05% for 45 min
T <sub>5</sub>	IBA	0.1% for 45 min
T <sub>6</sub>	IBA	0.05% for 45 min
T <sub>7</sub>	Hot water soaking	10-15 min
T <sub>8</sub>	Control	24 hrs

**Table 2:** Various parameters studied under the experiment

S. No.	Parameter	Formula
1	Imbibition Period	The number of days from sowing to the commencement of germination.
2	Speed of Germination:	Speed of Germination= $n1/d1+n2/d2+n3/d3+\dots$ Where n represents the number of germinated seeds and d represents the number of days.
3	Mean Daily Germination (MDG)	MDG=Total number of germinated seeds/ Total number of days
4	Peak Value (PV)	PV=Highest number of seeds germinated/ Number of days
5	Germination Value (GV)	GV=PV×MDG
6	Germination Percentage	Germination Percentage=Number of germinated seeds/ Total number of seeds×100

## Attributes of Seedling Vigour

### Seedling length

To determine the average seedling length, ten normal seedlings were randomly selected from each of the three replications. The seedling length was carefully measured in centimeters, starting from the root tip and extending to the tip of the primary leaf. This measurement provided a quantitative assessment of seedling growth performance

under various experimental conditions. Each seedling was evaluated based on specific parameters, including shoot length, root length, and total height, following the guidelines outlined in the Seedling Evaluation Handbook (reference). Seedlings that exhibited greater shoot and root lengths, as well as overall height, were considered to demonstrate superior vigor. This superior vigor is indicative of better overall growth performance, reflecting the effectiveness of

the different treatments applied during the experiment.

### Allometric index

The shoot length and root length recorded 30 days after sowing were used to calculate the allometric index using the formula described by Hosseini *et al.* (2013) [17].

Allometric index = Root length/ Shootlength

### Seedling dry weight (mg)

To measure seedling dry weight, the same ten seedlings used for length measurements were carefully dried in a hot air oven at 80 °C ± 2 °C for 24 hours. Once fully dried, the seedlings were allowed to cool in a desiccator for one hour to stabilize their weight. After cooling, the seedlings were weighed using an electronic balance, and the average seedling dry weight was recorded in milligrams, providing valuable data on the biomass accumulation of the seedlings under different treatments.

### Vigour indices

Vigour indices were determined using the formulas established by Abdul-Baki and Anderson for all substrates.

Vigour Index = Germination Percentage (%) × Average Seedling Dry Weight (mg)

### Statistical analysis

The experimental data were analyzed using a completely randomized design, as outlined by Panse and Sukhatme. The results are presented as the mean values of the replicates, with statistical significance determined by calculating the critical difference (CD) at a 5% significance level. The analysis was conducted using R software, ensuring rigorous statistical evaluation of the experimental outcomes.

## Results and Discussion

### Influence of Treatments on Germination Parameters

Various pre-sowing treatments were applied to assess their impact on germination parameters and seedling vigor. The

treatments significantly influenced these parameters, as shown in Table 3. Germination of Tulsi seeds began four days after the experiment's initiation. The imbibition period, indicating the time taken for seeds to absorb water and swell, varied notably based on the pre-treatments applied. The shortest imbibition period of five days was observed in seeds soaked in 0.1% GA<sub>3</sub>, indicating faster germination. In contrast, the control group, with no treatment, exhibited the longest imbibition period of 12 days.

The germination percentage was highest in seeds treated with GA<sub>3</sub> at both 0.1% and 0.05% concentrations, yielding 32% and 28% germination, respectively. These findings are consistent with Dzayi (2010) [11], who highlighted the role of GA<sub>3</sub> in promoting cell growth by facilitating calcium release from the cell wall into the cytoplasm. This process enhances water absorption and cell expansion, which are crucial for germination. After this growth phase, GA<sub>3</sub> is deactivated, and calcium is reabsorbed into the cell wall, leading to cell stiffening. During water absorption, the embryo generates GA<sub>3</sub>, triggering the production of hydrolytic enzymes like α- and β-amylase in the aleurone cells. These enzymes break down starch into glucose, which the embryo absorbs for nourishment. Additionally, GA<sub>3</sub> influences protein synthesis for mRNA production, aiding DNA replication and the breakdown of endosperm materials in the seed, as described by Lahuti *et al.* (2003) [23].

Similar effects of GA<sub>3</sub> on germination have been reported in *Atropa belladonna* (Genova *et al.* 1997) [16] and *Pimpinella anisum* (Shahrajabian *et al.*, 2019) [34]. The speed of germination was enhanced across all pre-sowing treatments compared to the control. The highest Peak Value (PV) was observed in seeds treated with 0.1% GA<sub>3</sub>, followed by hot water treatment, while the lowest PV and Germination Value (GV) were recorded in the control group, at 0.13 and 0.09, respectively. The highest GV was noted in seeds treated with 0.1% GA<sub>3</sub> (3.033), followed by 0.05% GA<sub>3</sub> (2.33). IBA treatments at both 0.1% and 0.05% concentrations showed similar effects. Prabhat and Tiwari (2019) reported comparable results, further supporting the findings of this study.

**Table 3:** The variations in germination parameters of *Ocimum tenuiflorum* L. due to pre-sowing treatments

Treatments	Germination Parameters					
	Germination percentage	Speed of germination	Imbibition period	MDG	PV	GV
GA <sub>3</sub> 0.1% (T <sub>1</sub> )	32.10a	15.23a	5g	4.10a	0.74a	3.03a
GA <sub>3</sub> 0.05% (T <sub>2</sub> )	28.31b	14.40b	6f	3.70b	0.63c	2.33b
KNO <sub>3</sub> 0.1% (T <sub>3</sub> )	26.41c	13.13c	7e	3.20c	0.33f	1.05c
KNO <sub>3</sub> 0.05% (T <sub>4</sub> )	25.31c	12.50d	7e	2.90d	0.27g	0.78d
IBA 0.1% (T <sub>5</sub> )	21.44d	10.00e	8d	2.00e	0.54d	1.08c
IBA 0.05% (T <sub>6</sub> )	18.63e	9.77e	9c	1.6f	0.44e	0.70d
Hot water soaking (T <sub>7</sub> )	15.31f	8.20f	11b	1.10g	0.70b	0.76d
Control (T <sub>8</sub> )	11.01g	5.70g	12a	0.70h	0.13h	0.09e
LSD <sub>p&lt;0.05</sub>	1.13	0.54	0.32	0.15	0.57	0.09

**MDG:** Mean daily germination, **PV:** Peak Value, **GV:** Germination Value

**Note:** Mean Values followed by different alphabets (a,b,c...) in a column varied significantly ( $p<0.05$ )

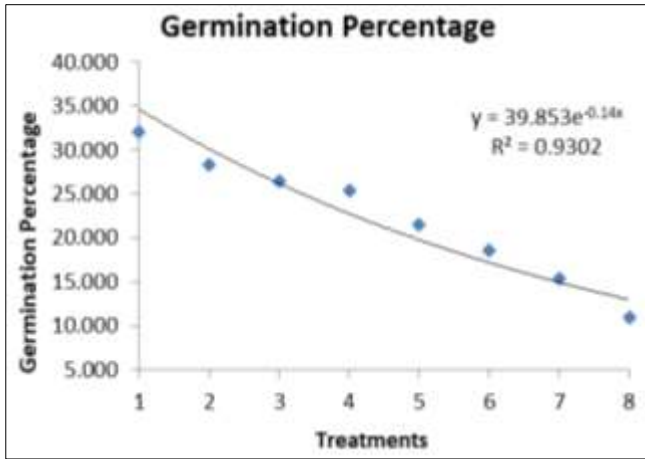


Fig 1: Relationship between different pre-sowing treatment with germination percentage and speed of germination

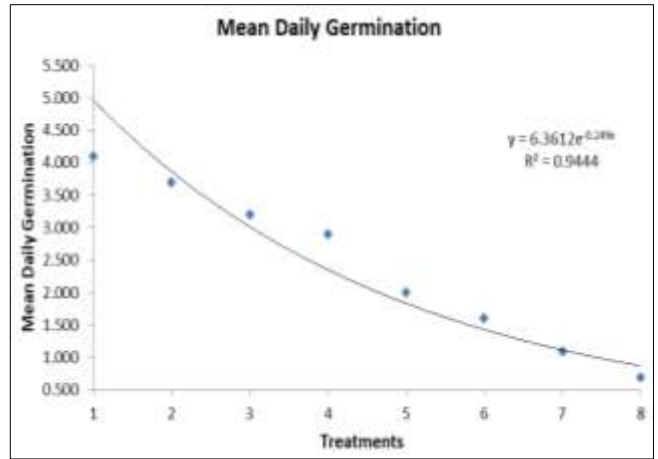


Fig 3: Relationship between different pre-sowing treatment with imbibition period

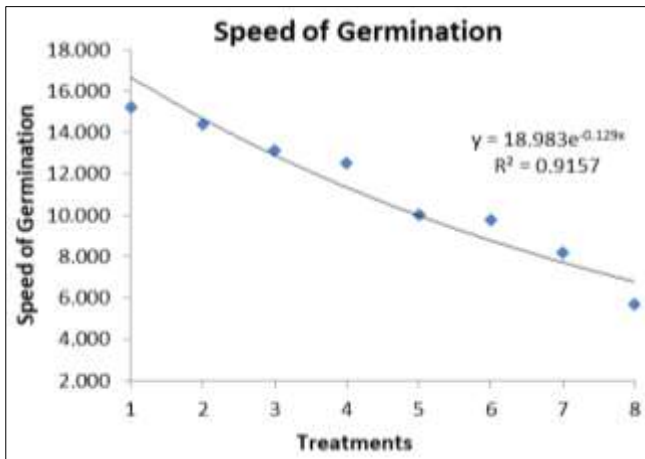


Fig 2: Relationship between speed of germination and treatment

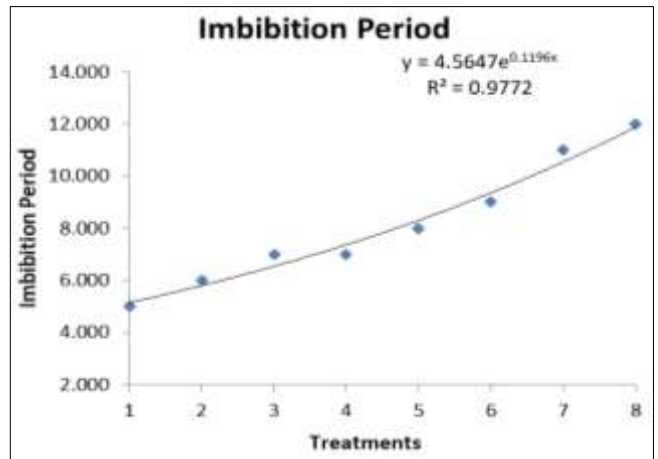


Fig 4: Relationship between different pre-sowing treatment with mean daily germination

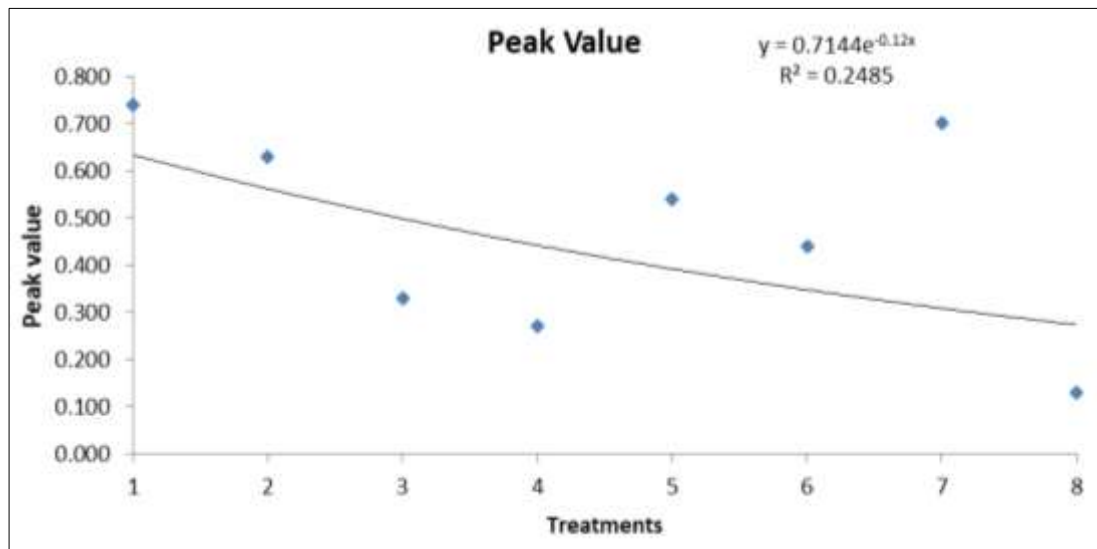


Fig 5: Relationship between different pre-sowing treatment with Peak value

**Seedling Vigour**

The study investigated the effects of various concentrations of Gibberellic Acid (GA3) on seedling growth characteristics and biomass, as shown in Table [reference]. The data reveal that different GA3 concentrations significantly influenced both shoot and root development. The highest shoot length of 15.31 cm was achieved with 0.1% GA3 treatment, while the shortest shoot length of 7.40

cm was recorded in the 0.05% GA3 treatment. Similarly, the longest root length of 9.40 cm was observed with 0.1% GA3, whereas the control group exhibited the shortest root length of 3.32 cm. This increase in root length, particularly with the 0.1% GA3 treatment, correlates with the enhanced above-ground growth, indicating a parallel improvement in root development.



Seedling dry weight was also affected by the treatments, with GA<sub>3</sub> and Potassium Nitrate (KNO<sub>3</sub>) showing superior performance compared to the control, which had the lowest dry weight of 0.25 g. This suggests that plant growth regulators (PGRs) such as GA<sub>3</sub> and KNO<sub>3</sub> can mitigate adverse effects on growth, potentially due to their influence on endogenous growth regulators (Thomas *et al.*, 2005) [39]. Previous research supports that GA<sub>3</sub> treatment promotes growth by activating physiological processes involved in seedling development (Taiz and Zeiger, 2010) [38].

Among all dormancy-breaking treatments, the longest seedling length was achieved with 0.1% GA<sub>3</sub>, measuring 24.71 cm, followed closely by 0.05% GA<sub>3</sub> at 23.20 cm. The control group had the shortest seedling length of 10.72 cm, consistent with findings by Singh *et al.* (2002) [35]. The enhanced performance of GA<sub>3</sub>-treated seeds is likely due to its role in breaking seed dormancy, which facilitates earlier and more vigorous germination by promoting the diffusion of endogenous auxins and gibberellin-like substances

(Gurung *et al.*, 2014) [19]. All GA<sub>3</sub> treatments effectively enhanced seedling growth compared to the control, likely due to the activation of amylase, which converts carbohydrates into simpler sugars, thus providing energy and nutrients for rapid seedling growth. Previous studies have also observed increased plant growth resulting from GA<sub>3</sub> treatment (Lee *et al.*, 1999; Rout *et al.*, 2016) [25, 32].

The allometric index, a measure of growth efficiency, was highest with 0.05% GA<sub>3</sub> (0.646), statistically comparable to 0.1% GA<sub>3</sub> (0.615), and followed by KNO<sub>3</sub> 0.05% (0.661). GA<sub>3</sub> 0.05% and IBA 0.1% treatments were statistically similar. Among all treatments, the highest vigor index of 20.218 was recorded with GA<sub>3</sub> 0.1%, followed by 15.33 with GA<sub>3</sub> 0.05%. The control had the lowest vigor index of 2.753. These results align with Singh *et al.* and are consistent with findings by Fazeli Kakhki *et al.*, who reported that gibberellins significantly enhance both germination and vigor index in scallion (*Allium fistulosum*) seeds.

**Table 4:** The variations in seedling vigor of *Ocimum tenuiflorum* L. due to pre-sowing treatments

Treatments	Seedling Vigor					
	Shoot length (cm)	Root length (cm)	seedling length (cm)	seedling weight (mg)	Vigor index	Allometric index
GA <sub>3</sub> 0.1% (T <sub>1</sub> )	15.3a	9.40a	24.71a	0.63a	20.21a	0.61c
GA <sub>3</sub> 0.05% (T <sub>2</sub> )	14.10b	9.10a	23.20b	0.58b	16.40b	0.64bc
KNO <sub>3</sub> 0.1% (T <sub>3</sub> )	12.60c	7.13d	19.73c	0.51c	13.47c	0.56d
KNO <sub>3</sub> 0.05% (T <sub>4</sub> )	11.45d	7.56c	19.01d	0.46d	11.64d	0.66bc
IBA 0.1% (T <sub>5</sub> )	10.00e	8.33b	18.33e	0.38e	8.14e	0.83a
IBA 0.05% (T <sub>6</sub> )	9.20f	6.37e	15.57f	0.33f	6.15f	0.69b
Hot water soaking (T <sub>7</sub> )	8.60g	4.60f	13.20g	0.29g	4.43g	0.53d
Control (T <sub>8</sub> )	7.40h	3.32g	10.72h	0.25h	2.75h	0.44e
LSD <sub>p&lt;0.05</sub>	0.56	0.32	0.53	0.02	0.57	0.04

**Note:** Mean Values followed by different alphabets (a,b,c...) in a column varied significantly ( $p<0.05$ )

## Conclusion

The findings of the study indicate that Gibberellic Acid (GA<sub>3</sub>) has a substantial effect on both seed germination and seedling growth in *Ocimum tenuiflorum* L. The results demonstrate that a 0.1% GA<sub>3</sub> pre-treatment significantly enhances germination rates, accelerates seedling growth, and improves overall seedling quality. Given these outcomes, it is recommended to use a 0.1% GA<sub>3</sub> solution for pre-treating seeds of *Ocimum tenuiflorum* L. This treatment optimizes germination, promotes robust seedling development, and ensures better quality seedlings, thereby benefiting cultivation practices for this important medicinal plant species.

## References

- Abdul-Baki AA, Anderson JD. Vigor determination in soybean seed by multiple criteria. *Crop Sci.* 1973;13(6):630-633.
- Ali T, Hossein P, Asghar F, Salman Z, Ali ZCM. The effect of different treatments on improving seed germination characteristics in medicinal species of *Descurainia sophia* and *Plantago ovata*. *Afr J Biotechnol.* 2010;9(39):6588-6593.
- Aslan I, Ozbek H, Çalmaşur O, Şahin F. Toxicity of essential oil vapours to two greenhouse pests, *Tetranychus urticae* Koch and *Bemisia tabaci* Genn. *Ind Crops Prod.* 2004;19(2):167-173.
- Association of Official Seed Analysts. Rules for testing seeds. The Association. 1978;12:18-19.
- Berbec S, Andruszczak S, Łusiak J, Sapko A. Effect of foliar application of Atonik and Ekolist on yield and quality of common thyme. *Acta Agrophys.* 2021;2003(85):305-311.
- Bowers WS, Nishida R. Juvocimenes: potent juvenile hormone mimics from sweet basil. *Science.* 1980;208:1030-1032.
- Bozin B, Mimica-Dukic N, Simin N, Anackov G. Characterization of the volatile composition of essential oils of some *Lamiaceae* spices and the antimicrobial and antioxidant activities of the entire oils. *J Agric Food Chem.* 2006;54(5):1822-1828.
- Carvajal M, Alcaraz CF. Why titanium is a beneficial element for plants. *J Plant Nutr.* 1998;21(4):655-664.
- Copeland LO, McDonald MF. Principles of seed science and technology. Dordrecht: Kluwer Academic Publishers; c2012.
- Czabator FJ. Germination value: an index combining speed and completeness of Pine seed germination. *Seed Sci Technol.* 1962;8:386-396.
- Dzayi FH, Rahman. Effect of GA<sub>3</sub> and Soaking Time on Seed Germination and Seedling Growth Lemon (*Citrus lemon* L.) High Diploma. Thesis. Univ. of Salahaddin – Erbil; c2010.
- Elgayyar M, Draughon FA, Golden DA, Mount JR. Antimicrobial activity of essential oils from plants against selected pathogenic and saprophytic microorganisms. *J Food Prot.* 2001;64(7):1019-1024.

13. Esechie HA. Interaction of salinity and temperature on the germination of sorghum. *J Agron Crop Sci.* 1994;172(3):194-199.
14. Fazeli Kakhki SF, Beikzadeh N. Effect of gibberellin and indole-3-butyric acid on germination indices and vigor of scallion (*Allium fistulosum*) seeds. *Iran Agric Res.* 2022;41(1):39-47.
15. Finch-Savage WE. Influence of seed quality on crop establishment, growth, and yield. In: *Seed Quality.* CRC Press; c2020. p. 361-384.
16. Genova E, Komitska G, Beeva Y. Study on the germination of *Atropa belladonna* L. seeds. *Bulg J Plant Physiol.* 1997;23(1-2):61-66.
17. Gholami H, Farhadi R, Rahimi M, Zeinalikharaji A, Askari A. Effect of growth hormones on physiology characteristics and essential oil of basil under drought stress condition. *J Am Sci.* 2013;9:61-63.
18. Gupta V. Seed germination and dormancy breaking techniques for indigenous medicinal and aromatic plants. *J Med Aromatic Plants Sci.* 2003;25:402-407.
19. Gurung N, Swamy GSK, Sarkar SK, Ubale NB. Effect of chemicals and growth regulators on germination, vigour and growth of passion fruit (*Passiflora edulis* Sims.). *The Bioscan.* 2014;9(1):155-157.
20. Hidayati SN, Walck JL, Merritt DJ, Turner SR, Turner DW, Dixon KW. Sympatric species of *Hibbertia* (Dilleniaceae) vary in dormancy break and germination requirements: implications for classifying morpho-physiological dormancy in Mediterranean biomes. *Ann Bot.* 2012;109(6):1111-1123.
21. Kandari LS, Rao KS, Maikhuri RK, Chauhan K. Effect of pre-sowing, temperature and light on the seed germination of *Arnebia benthamii* (Wall. ex G. Don): An endangered medicinal plant of Central Himalaya, India. *Afr J Plant Sci.* 2008;2(1):5-11.
22. Kristinsson KG, Magnúsdóttir AB, Petersen H, Hermansson A. Effective treatment of experimental acute otitis media by application of volatile fluids into the ear canal. *J Infect Dis.* 2005;191(11):1876-1880.
23. Lahuti M, Zare-hasanabadi M, Ahmadian R. Biochemistry and physiology of vegetable hormones. Ferdosi University Mashhad; c2003.
24. Lecat S, Corbineau F, Come D. Effects of gibberellic acid on the germination of dormant oat (*Avena sativa* L.) seeds as related to temperature, oxygen, and energy metabolism. *Seed Sci Technol.* 1992;20:421-433.
25. Lee J, Joung KT, Hayain KH, Hee LS. Effect of chilling and growth regulators in seedling stage on flowering of *Lilium formolongi*. *Hangut Wanyae Hakcheochi.* 1999;40(2):248-252.
26. Macchia M, Angelini LG, Ceccarini L. Methods to overcome seed dormancy in *Echinacea angustifolia* DC. *Sci Hort.* 2001;89(4):317-324.
27. Panse VS, Sukhatme PV. Statistical methods for Agricultural workers. 4<sup>th</sup> ed. New Delhi: ICAR Publication; c1985.
28. Papadopoulou AP, Saha U, Hao X, Khosla S. Response of rockwool-grown greenhouse cucumber, tomato, and pepper to kinetin foliar sprays. *Hort Technology.* 2006;16:32-35.
29. Paton A, Harley RM, Harley MM. An overview of relationships and classification. In: Holm Y, Hiltunen R, editors. *Ocimum. Medicinal and Aromatic Plants- Industrial Profiles.* Amsterdam: Harwood Academic; c1999.
30. Pereira TS. Germination of seeds of *Bauhinia forficata* Link. (Leguminosae-Caesalpinoideae). *J Seeds.* 1992;14:77-82.
31. Rasool Hassan BA. Medicinal plants (importance and uses). *Pharmaceut Anal Acta.* 2012;3(10):2153-2155.
32. Rout S, Beura S, Khare N. Effect of GA3 on seed germination of *Delonix regia*. *Res J Recent Sci.* 2016;(ISC-2015, Special issue):5-7.
33. Sajadi S. Analysis of the essential oils of two cultivated basil (*Ocimum basilicum* L.) from Iran. *Daru.* 2006;14:128-130.
34. Shahrajabian MH, Sun W, Cheng Q. Clinical aspects and health benefits of ginger (*Zingiber officinale*) in both traditional Chinese medicine and modern industry. *Acta Agric Scand B Soil Plant Sci.* 2019;69(6):546-556.
35. Singh N, Hoette Y, Miller DR. Tulsi: The mother medicine of nature. *Int Inst Herbal Med.* 2002;51(1):43-49.
36. Sivakumar V, Anandalakshmi R, Warriar RR, Tigabu M, Ode PC, Vijayachandran SN, *et al.* Effects of pre-sowing treatments, desiccation and storage conditions on germination of *Strychnos nux-vomica* seeds, a valuable medicinal plant. *New For.* 2006;32:121-131.
37. Suppakul P, Miltz J, Sonneveld K, Bigger SW. Antimicrobial properties of basil and its possible application in food packaging. *J Agric Food Chem.* 2003;51(11):3197-3207.
38. Taiz L, Zeiger E. *Plant Physiology.* Sinauer Associates Inc.; c2010.
39. Thomas SG, Rieu I, Steber CM. Gibberellin metabolism and signaling. *Vitam Horm.* 2005;72:289-338.
40. Zare AR, Solouki M, Omidi M, Irvani N, Abasabadi AO, Nejad NM. Effect of various treatments on seed germination and dormancy breaking in *Ferula assa foetida* L. (Asafetida), a threatened medicinal herb. *Trakia J Sci.* 2011;9(2):57-61.
41. Mandal A, Patra AK, Singh D, Swarup A, Masto RE. Effect of long-term application of manure and fertilizer on biological and biochemical activities in soil during crop development stages. *Bioresource technology.* 2007 Dec 1;98(18):3585-3592.