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**Pawan Kumar**  
 Research Scholar, Department  
 of Horticulture, NAI,  
 SHUATS, Prayagraj,  
 Uttar Pradesh, India

**Vijay Bahadur**  
 Professor, Department of  
 Horticulture, NAI, SHUATS,  
 Prayagraj, Uttar Pradesh,  
 India

**Annjoe V Joseph**  
 Assistant Professor,  
 Department of Horticulture,  
 NAI, SHUATS, Prayagraj,  
 Uttar Pradesh, India

**Arun A David**  
 Associate Professor and Head,  
 Department of Soil Science and  
 Agricultural Chemistry, NAI,  
 SHUATS, Prayagraj,  
 Uttar Pradesh, India

**Joy Dawson**  
 Professor and Head,  
 Department of Agronomy,  
 NAI, SHUATS, Prayagraj,  
 Uttar Pradesh, India

**Corresponding Author:**  
**Pawan Kumar**  
 Research Scholar, Department  
 of Horticulture, NAI,  
 SHUATS, Prayagraj,  
 Uttar Pradesh, India

## The impact of various soilless substrates and chitosan application on flowering characteristics of *Lilium cv.* break out grown under shade net condition

**Pawan Kumar, Vijay Bahadur, Annjoe V Joseph, Arun A David and Joy Dawson**

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### Abstract

The current study, titled “The impact of various soilless substrates and chitosan application on flowering characteristics of *Lilium cv.* Break Out grown under shade net conditions,” was conducted at the Department of Horticulture, Sam Higginbottom University of Agriculture, Technology and Sciences, during the years 2022-23 and 2023-24. Throughout both years, as well as in the aggregated data, significant differences were noted in the interaction effects of various soilless substrates and the foliar application of different concentrations of chitosan on flowering and bulb yield. The findings revealed that the earliest flowering times, specifically the days required for bud initiation (44.63), the days from bud initiation to bud color appearance (19.17), and the days until flower opening (67.71), were significantly reduced in the treatment T<sub>11</sub> (M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1) + C<sub>2</sub>: 250 ppm Chitosan). In contrast, the treatment T<sub>11</sub> exhibited significantly higher values for flower characteristics, including the average number of buds per plant (3.25) and the flower diameter (14.53 cm).

**Keywords:** Soilless substrates, chitosan, flowering, bulb and *Lilium*

### Introduction

*Lilium* stands out as one of the most captivating ornamental plants, renowned for its aesthetic appeal, diverse forms, and a wide spectrum of colors. It is characterized as a low-volume, high-value crop and is often regarded as a symbol of purity and nobility. Its versatility in the floral industry is evident, as it is utilized both as cut flowers and potted plants. Native to the Northern Hemisphere, *Lilium* is extensively found in regions such as China, Japan, southern Canada, Siberia, and even extends into Florida in the United States. It ranks among the six principal genera of flower bulbs cultivated globally. The economic significance of *Lilium* is notable, particularly in the production and commercialization of cut flowers in the international market (Jimenez *et al.*, 2012) <sup>[4]</sup>. Its high market value and popularity stem from its rich variety of colors and the abundance of blooms it produces (Chaudhari *et al.*, 2024) <sup>[2]</sup>. *Lilium* is one of the most prevalent bulbous ornamental plants. Horticulturists highly value the cultivars of the *Lilium* genus for their exceptional color range, fragrance, and adaptability to various environmental conditions (Bahr and Compton, 2004) <sup>[1]</sup>. The implementation of shade nets is intended to enhance favorable physiological responses while also providing physical protection, significantly influencing shoot elongation, branching, and flowering in ornamental crops (Oren-Shamir *et al.*, 2001) <sup>[7]</sup>. The physical and chemical properties of soilless substrates are vital for ensuring adequate support and nutrient supply to plants; these substrates must be lightweight, porous, and well-drained (Noguera *et al.*, 2003) <sup>[6]</sup>. The choice of potting media is crucial for the quality and production of flowering plants, with natural soil and peat being the most commonly used growing substrates for the container cultivation of both annual and perennial ornamental plants (Tariq *et al.*, 2012) <sup>[11]</sup>. The use of peat in potting media is commercially costly and cannot be reused; however, the development of affordable alternatives utilizing nutrient-rich organic materials may lead to reduced fertilization and irrigation needs, as well as lower nursery expenses, as noted by Wilson and Stoffella (2002) <sup>[13]</sup>. Peat remains the predominant substrate for the cultivation of potted plants in nurseries, constituting a substantial portion of the materials

employed in this sector, according to Ribeiro *et al.* (2007) [8]. Consequently, there is an increasing demand for lightweight growing media, which are favored for their ease of transport and presentation at exhibitions and flower shows, as highlighted by Dubey *et al.* (2013) [13]. Various lightweight media, such as perlite and leaf mold, have been shown to modify the physical and chemical properties of growing mixtures, thereby influencing the growth of potted ornamental plants, as reported by Vendrame *et al.* (2005) [12] and Kumar *et al.* (2022) [5]. Chitosan, a significant biopolymer, enhances plant growth characteristics, flowering behavior, and yield traits, as indicated by Sultana *et al.* (2019) [10]. This carbohydrate, derived from a glucose framework, features a linear chain structure and includes free amino groups, resembling cellulose. Chemically, chitosan is a linear polymer made up of two subunits, D-glucosamine and N-acetyl-D-glucosamine, which are connected by 1,4-glycosidic bonds, as described by Rinaudo *et al.* (2003) [9]. Chitosan can be sourced from the exoskeletons of insects or extracted from marine crustaceans, including crabs and prawns.

### Materials and Methods

The present investigation 'Influence of different soilless substrates and chitosan treatment on flowering and bulb yield of *Lilium* cv. Break Out grown under shade net condition' was conducted at Department of Horticulture, Sam Higginbottom University of Agriculture, Technology and Sciences, during the year 2022-23 and 2023-24. The experimental material for the research investigation was Asiatic *Lilium* cv. Break out. For the first and second year research trails, Bulbs were obtained from Holland. For the experiment, evenly sized *Lilium* bulbs (12/14 size) were selected. Bulbs were planted at a depth of 5-7 inches. The pots were maintained in shade net house. Different soilless substrates were prepared using peat, perlite, leaf mould and coconut fiber in different proportions. The constituents of growing media were first solar sterilized for a week before planting of the bulbs. Thorough cleaning of the media constituents was done by removing the stones, pebbles and unwanted materials present in it. Different growing media were prepared by mixing peat, perlite, leaf mould and coconut fiber in required ratios volume by volume. The respective media were then filled into earthen pots provided with drainage holes for aeration and drainage of excess water. The experiment comprised of different soilless substrates growing media viz., (Peat, perlite, leaf mould and coconut fiber) and foliar application of different concentration of chitosan viz., (0.00, 100, 250 and 400 ppm) having 16 treatment combinations. The experimental was laid out in Factorial Complete Randomized Design (FCRBD) with three replications and the data were analyzed accordingly. The treatments in each replication were allotted randomly. Sixteen factor combinations having one cultivar were tried in the experimental design. Five plants were randomly chosen from each treatment group and labeled for the purpose of recording observations. Each repetition of the different treatments yielded the necessary data, from which the average values were then computed. The data generated from present investigations were subjected to the statistical analysis in accordance with the procedure outlined by Panse and Sukhatme (1985) [30]. The significance of differences among growing conditions, foliar application treatments and their combinations means were tested by F-test.

### Results and Discussion

The data appraisal to various flowering parameters were influenced by different soilless substrates growing media viz., (Peat, perlite, leaf mould and coconut fiber) and foliar application of different concentration of chitosan in pot *Lilium* were presented in tables 1 and graph 2 and 3 during both the year of research investigations. During both the years as well as in the pooled values, Significantly minimum days taken to bud initiation (45.47), days taken to bud initiation to bud color shown (19.95) and days taken to flower opening (67.00) was recorded in the plants growing in M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1), which was statistically at par with treatment M<sub>1</sub>: Peat + Perlite (1:1) and M<sub>3</sub>: Peat + Perlite + Coconut fiber (1:1:1). However, maximum days taken to bud initiation (48.00), days taken to bud initiation to bud color shown (23.99) and days taken to flower opening (69.77) was reported in the plant growing in control M<sub>0</sub>: Soil. During the first and second year (2022-23 & 2023-24) research experiment with the pooled data respectively. During both the years as well as in the pooled values, flower attributes was found to be significantly affected by the soilless substrates as shown in Table 1 and Fig. 1 & 2. The observed phenomenon may be a result of the plant's vigorous growth within the media, which significantly influences early production through the rapid absorption of nutrients and water. Furthermore, the buildup of photosynthetic substances within this medium may have prompted the onset of early flowering. This phenomenon is probably associated with a heightened synthesis of proteins originating from the carbohydrates accumulated in the vegetative structures, along with improved protoplast development. A comparable outcome was observed by Swarup (1972) [18] in *Gerbera*, as well as by Jawaharlal *et al.* (2001) [17] in *Anthurium* and *Rose*, and by Barreto *et al.* (2006) [23] in *Gerbera*. Significantly maximum number of buds per plant (2.74) was recorded in the plants growing in M<sub>3</sub>: Peat + Perlite + Coconut fiber (1:1:1), which was statistically at par with treatment M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1). However, minimum number of buds per plant (2.27) was reported in the plant growing in control M<sub>0</sub>: Soil. During the first and second year (2022-23 & 2023-24) research experiment with the pooled data respectively. Significantly maximum flower diameter (cm) (13.99) was recorded in the plants growing in M<sub>3</sub>: Peat + Perlite + Leaf Mould (1:1:1), which was statistically at par with treatment M<sub>3</sub>: Peat + Perlite + Coconut fiber (1:1:1). However, minimum flower diameter (12.96 cm) was reported in the plant growing in control M<sub>0</sub>: Soil, during the first and second year (2022-23 & 2023-24) research experiment with the pooled data respectively. The observed increase in size may result from the combined effects of various growth parameters. Enhanced yield parameters within the growing medium contributed to the enlargement of the flowers, facilitated by the availability of moisture and nutrients. The observed enlargement of flower size might be correlated with the increased area of leaves, potentially arising from the augmented production and storage of photosynthetic substances that are conveyed from the leaves to the flowers. The application of potting materials such as cocopeat and leaf mold is likely a factor in the elevated potassium concentration, as well as in the advantageous physicochemical traits, which include high porosity, remarkable water retention capacity, and improved moisture conservation. Collectively, these factors play a significant

role in enhancing petal length, petal width, the number of petals per flower, and the overall diameter of rose flowers. This assertion is supported by the studies of Ysmeen *et al.* (2012) <sup>[24]</sup> on carnations, along with the investigations by Barreto and Jagtap (2006) <sup>[14]</sup> and Barman *et al.* (2006) <sup>[15]</sup> focusing on both roses and carnations. The plants cultivated in leaf mould demonstrate their potential under optimal conditions of bulk density, electrical conductivity (EC), and pH, resulting in enhanced flower production. As substances resembling humus, leaf mould exhibited similar characteristics that facilitated nutrient absorption and positively influenced protein synthesis and vegetative growth, thereby leading to an increase in flower yield. Similar results were obtained by Ysmeen *et al.* (2012) <sup>[24]</sup> in rose, Anthurium in Jawaharlal *et al.* (2001) <sup>[17]</sup>, Carnation in Bhatia *et al.* (2004) <sup>[21]</sup>, Gerbera Gupta *et al.* (2004) <sup>[22]</sup> and Carnation in Ysmeen *et al.* (2012) <sup>[24]</sup>. Lopez *et al.* (2008) <sup>[20]</sup> indicated that various growing media had a substantial impact on the growth and flowering of gladiolus (*Gladiolus tristis* subsp. Concolor). They found that plants cultivated individually in peat exhibited greater stem length, spike length, and number of florets compared to those grown in perlite. Furthermore, peat moss-based substrates produced higher yields and superior flower quality when compared to single substrate options, as noted by Ümmü Özgül Karagüzel (2023) <sup>[19]</sup>. During both the years as well as in the pooled values, flowering and bulb yield was found to be significantly affected by the foliar application of different concentration of chitosan as shown in Table 1 and Fig. 1 & 2. Among the foliar application of various concentration of chitosan, Significantly lowest days taken to bud initiation (45.80), days taken to bud initiation to bud color shown (20.66) and days taken to flower opening (69.294) was recorded in C<sub>2</sub> which includes foliar application of 250 ppm Chitosan which was statistically at par with treatment C<sub>3</sub>:400 ppm Chitosan and C<sub>1</sub>:100 ppm Chitosan. However, highest days taken to bud initiation (47.04), days taken to bud initiation to bud color shown (23.15) and days taken to flower opening (70.20) was recorded in C<sub>0</sub> (foliar application of water). During both the years as well as in the pooled values, Among the foliar application of various concentration of chitosan, Significantly highest number of buds per plant (2.88) and flower diameter (cm) (13.64) was recorded in C<sub>2</sub> which includes foliar application of 250 ppm Chitosan which was statically at par with treatment C<sub>3</sub>:400 ppm Chitosan and C<sub>1</sub>:100 ppm Chitosan. However, lowest number of buds per plant 2.32) and flower diameter (cm) (13.26) was recorded in C<sub>0</sub> (foliar application of water). The application of a chitosan solution to *Ornithogalum*

*saunderiae* bulbs prior to planting resulted in taller plants that flowered earlier (Salachna *et al.*, 2015) <sup>[26]</sup>. Additionally, the use of chitosan improved the inflorescence length and width of pineapple lilies, leading to earlier flowering compared to the control group (Byczyńska, 2018) <sup>[27]</sup>. In addition, research conducted by Ramos-Garcia *et al.* (2009) <sup>[31]</sup> demonstrated that the application of chitosan as a natural stimulant significantly improved the production of gladiolus corms. Notably, the most pronounced increase in corm weight was recorded in freesia plants subjected to chitosan treatment (Salachna and Zawadzińska, 2014) <sup>[25]</sup>. Conversely, Rady *et al.* (2018) <sup>[32]</sup> suggested that soaking garlic bulbs in a seaweed extract at a concentration of 3 ml/l is effective for promoting the growth of larger bulbs. Furthermore, *Dracaena surculosa* plants that received chitosan treatment showed the highest levels of total nitrogen and chlorophyll a in their leaves, with these parameters rising in correlation with increased chitosan concentrations (El-Khateeb *et al.*, 2018; Abd-El-Hady, 2020) <sup>[28, 29]</sup>. Significant differences were noted in the interaction effects of various soilless substrates and the foliar application of different concentrations of chitosan on both flowering and bulb yield, across both years and in the pooled data. The interaction results showed that, minimum days taken to bud initiation, days taken to bud initiation to bud color shown and days taken to flower opening was recorded in treatment T<sub>11</sub>: (M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1) + C<sub>2</sub>: 250 ppm Chitosan) which was statistically at par with treatment T<sub>14</sub>: (M<sub>3</sub>: Peat + Perlite + Coconut fiber (1:1:1) + C<sub>3</sub>: 100 ppm Chitosan), T<sub>10</sub> (M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1) + C<sub>1</sub>: 100 ppm Chitosan) and T<sub>9</sub>: (M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1) + C<sub>0</sub>: Water). Whereas the maximum days taken to bud initiation, days taken to bud initiation to bud color shown and days taken to flower opening was recorded in control T<sub>3</sub>: (M<sub>0</sub>: Soil + C<sub>0</sub>: Water). Where the interaction results showed that, maximum number of buds per plant, bud length (cm), bud diameter (cm), length of floret (cm), width of floret (cm), flower diameter (cm), spike length (cm), vase life (days), number of bulbs per plant, number of bulblets per mother bulb, weight of bulbs (gm) and bulb diameter (cm) was recorded in treatment T<sub>11</sub>: (M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1) + C<sub>2</sub>: 250 ppm Chitosan) which was statistically at par with treatment T<sub>14</sub> (M<sub>3</sub>: Peat + Perlite + Coconut fiber (1:1:1) + C<sub>1</sub> 100 ppm Chitosan), T<sub>10</sub> (M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1) + C<sub>1</sub>: 100 ppm Chitosan) and T<sub>9</sub> (M<sub>2</sub>: Peat + Perlite + Leaf Mould (1:1:1) + C<sub>0</sub> Water). Whereas the minimum number of buds per plant and flower diameter (cm) was recorded in control T<sub>1</sub> (M<sub>0</sub> Soil + C<sub>0</sub> Water).

**Table 1:** Effect of soilless substrates and chitosan treatment on flowering bulb yield parameters of *Lilium* cv. Break Out grown under shade net condition

| Effect of Growing Media (v/v)  | Days taken to bud initiation |         |        | Days taken to bud initiation to bud color shown |         |        | Days taken to flower opening |         |        |
|--|------------------------------|---------|--------|---|---------|--------|------------------------------|---------|--------|
|  | 2022-23                      | 2023-24 | Pooled | 2022-23   | 2023-24 | Pooled | 2022-23                      | 2023-24 | Pooled |
| M <sub>0</sub> : Soil  | 48.25                        | 47.75   | 48.00  | 20.77   | 27.21   | 23.99  | 67.88                        | 71.65   | 69.77  |
| M <sub>1</sub> : Peat + Perlite (1:1)  | 46.25                        | 47.48   | 46.87  | 20.15   | 22.73   | 21.44  | 67.71                        | 71.64   | 69.68  |
| M <sub>2</sub> : Peat + Perlite + Leaf Mould (1:1:1)   | 45.35                        | 45.58   | 45.47  | 18.81   | 21.08   | 19.95  | 63.56                        | 70.44   | 67.00  |
| M <sub>3</sub> : Peat + Perlite + Coconut fiber (1:1:1)  | 46.25                        | 46.25   | 46.25  | 19.73   | 21.67   | 20.70  | 67.25                        | 71.17   | 69.21  |
| F-Test   | S                            | S       | S      | S   | S       | S      | S                            | S       | S      |
| S.Ed. (±)  | 0.447                        | 0.427   | 0.2704 | 0.193   | 0.214   | 0.1385 | 0.469                        | 0.689   | 0.4193 |
| CD (5%)  | 0.910                        | 0.869   | 0.5508 | 0.393   | 0.435   | 0.2821 | 0.956                        | 1.404   | 0.8541 |
| Effect of Chitosan   |                              |         |        |   |         |        |                              |         |        |
| C <sub>0</sub> -Water  | 47.23                        | 46.85   | 47.04  | 20.25   | 26.04   | 23.15  | 68.604                       | 71.791  | 70.20  |
| C <sub>1</sub> :100 ppm Chitosan   | 47.21                        | 46.83   | 47.02  | 20.23   | 22.27   | 21.25  | 68.25                        | 71.042  | 69.65  |
| C <sub>2</sub> :250 ppm Chitosan   | 45.6                         | 46.00   | 45.80  | 19.25   | 22.06   | 20.66  | 67.771                       | 70.813  | 69.29  |
| C <sub>3</sub> :400 ppm Chitosan   | 46.06                        | 47.38   | 46.72  | 19.73   | 22.31   | 21.02  | 67.791                       | 71.249  | 69.52  |
| F-Test   | S                            | S       | S      | S   | S       | S      | S                            | S       | S      |
| S.Ed. (±)  | 0.447                        | 0.427   | 0.2704 | 0.193   | 0.214   | 0.1385 | 0.469                        | 0.689   | 0.4193 |
| CD (5%)  | 0.910                        | 0.869   | 0.5508 | 0.393   | 0.435   | 0.2821 | 0.956                        | 1.404   | 0.8541 |
| Treatment combinations   |                              |         |        |   |         |        |                              |         |        |
| T <sub>1</sub> (M <sub>0</sub> : Soil + C <sub>0</sub> :Water )  | 49.58                        | 48.92   | 49.25  | 22.75   | 24.27   | 23.51  | 71.333                       | 72.80   | 72.83  |
| T <sub>2</sub> (M <sub>0</sub> : Soil + C <sub>1</sub> :100 ppm Chitosan)                                    | 49.33                        | 48.17   | 48.75  | 20.42   | 23.33   | 21.875 | 69.83                        | 71.33   | 70.58  |
| T <sub>3</sub> (M <sub>0</sub> : Soil + C <sub>2</sub> :250 ppm Chitosan)                                    | 48.34                        | 46.5    | 47.42  | 20.58   | 23.83   | 22.205 | 69.000                       | 71.333  | 70.17  |
| T <sub>4</sub> (M <sub>0</sub> : Soil + C <sub>3</sub> :400 ppm Chitosan)                                    | 45.75                        | 46.59   | 46.17  | 19.33   | 21.67   | 20.50  | 68.083                       | 71.667  | 69.88  |
| T <sub>5</sub> (M <sub>1</sub> : Peat + Perlite (1:1) + C <sub>0</sub> :Water)                               | 47.83                        | 47.92   | 47.88  | 19.50   | 23.50   | 21.50  | 69.167                       | 70.667  | 69.92  |
| T <sub>6</sub> (M <sub>1</sub> : Peat + Perlite (1:1) + C <sub>1</sub> :100 ppm Chitosan)                    | 45.67                        | 47.58   | 46.63  | 21.25   | 22.08   | 21.665 | 67.75                        | 70.587  | 69.17  |
| T <sub>7</sub> (M <sub>1</sub> : Peat + Perlite (1:1) + C <sub>2</sub> :250 ppm Chitosan)                    | 44.92                        | 46.59   | 45.76  | 20.83   | 22.66   | 21.745 | 66.583                       | 71.000  | 68.79  |
| T <sub>8</sub> (M <sub>1</sub> : Peat + Perlite (1:1) + C <sub>3</sub> :400 ppm Chitosan)                    | 46.58                        | 48.67   | 47.63  | 19.00   | 24.00   | 21.50  | 68.083                       | 72.247  | 70.17  |
| T <sub>9</sub> (M <sub>2</sub> : Peat + Perlite + Leaf Mould (1:1:1) + C <sub>0</sub> :Water)                | 44.92                        | 44.92   | 44.92  | 19.00   | 20.75   | 19.875 | 69.000                       | 71.747  | 70.37  |
| T <sub>10</sub> (M <sub>2</sub> : Peat + Perlite + Leaf Mould (1:1:1) + C <sub>1</sub> :100 ppm Chitosan)    | 46.17                        | 47.09   | 46.63  | 18.83   | 21.92   | 20.375 | 68.42                        | 70.667  | 69.54  |
| T <sub>11</sub> (M <sub>2</sub> : Peat + Perlite + Leaf Mould (1:1:1) + C <sub>2</sub> :250 ppm Chitosan)    | 44.83                        | 44.42   | 44.63  | 18.17   | 20.17   | 19.17  | 66.167                       | 69.253  | 67.71  |
| T <sub>12</sub> (M <sub>2</sub> : Peat + Perlite + Leaf Mould (1:1:1) + C <sub>3</sub> :400 ppm Chitosan)    | 45.17                        | 45.92   | 45.55  | 19.25   | 21.50   | 20.375 | 67.247                       | 70.08   | 68.66  |
| T <sub>13</sub> (M <sub>3</sub> : Peat + Perlite + Coconut fiber (1:1:1) + C <sub>0</sub> :Water)            | 46.67                        | 45.92   | 46.30  | 19.67   | 22.75   | 21.21  | 67.75                        | 70.917  | 69.33  |
| T <sub>14</sub> (M <sub>3</sub> : Peat + Perlite + Coconut fiber (1:1:1) + C <sub>1</sub> :100 ppm Chitosan) | 47.67                        | 44.5    | 46.09  | 21.17   | 20.25   | 20.71  | 67.00                        | 70.583  | 68.79  |
| T <sub>15</sub> (M <sub>3</sub> : Peat + Perlite + Coconut fiber (1:1:1) + C <sub>2</sub> :250 ppm Chitosan) | 45.5                         | 46.5    | 46.00  | 18.67   | 22.58   | 20.625 | 66.5                         | 71.667  | 69.08  |
| T <sub>16</sub> (M <sub>3</sub> : Peat + Perlite + Coconut fiber (1:1:1) + C <sub>3</sub> :400 ppm Chitosan) | 45.5                         | 48.09   | 46.80  | 19.42   | 21.09   | 20.255 | 67.75                        | 70.503  | 69.13  |
| F-Test   | S                            | S       | S      | S   | S       | S      | S                            | S       | S      |
| S.Ed. (±)  | 0.893                        | 0.853   | 0.5408 | 0.386   | 0.427   | 0.277  | 0.939                        | 1.378   | 0.8386 |
| CD (5%)  | 1.819                        | 1.738   | 1.1017 | 0.787   | 0.870   | 0.5643 | 1.912                        | 2.808   | 1.7082 |

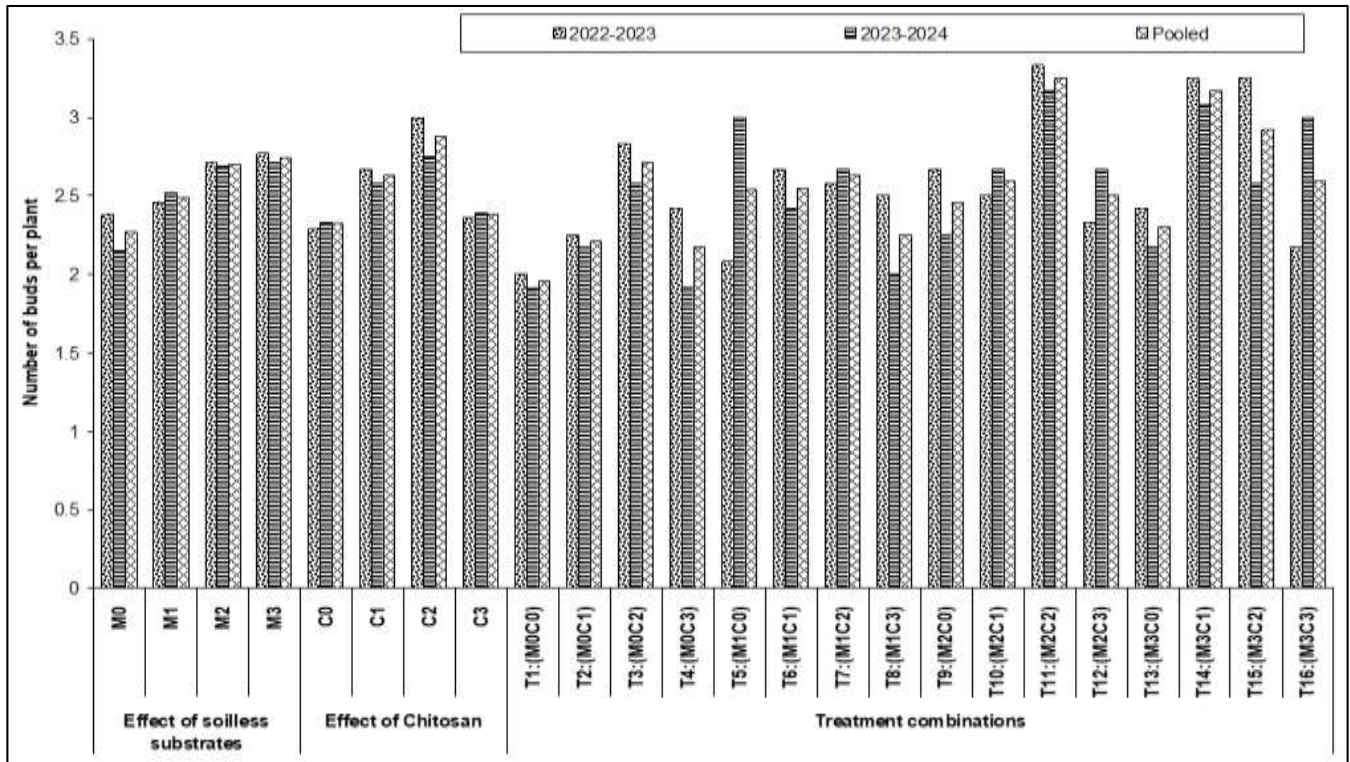


Fig 1: Effect of soilless substrates and chitosan treatment on number of buds per plant of *Lilium* cv. Break Out

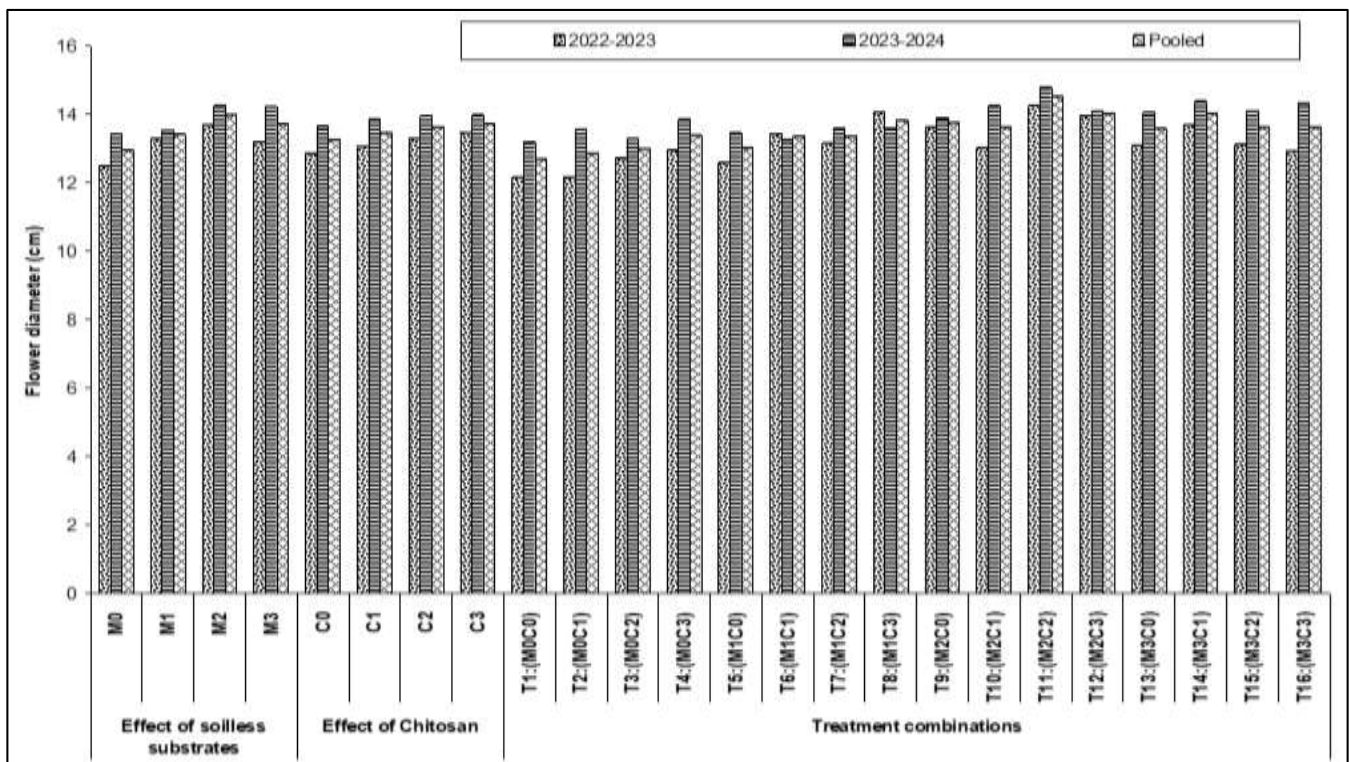


Fig 2: Effect of soilless substrates and chitosan treatment on flower diameter (cm) of *Lilium* cv. Break Out

**Conclusion**

The findings from the current study indicate that the treatment T<sub>11</sub>, which consists of a mixture of Peat, Perlite, and Leaf Mould in a 1:1:1 ratio combined with 250 ppm Chitosan, resulted in the shortest duration for bud initiation (44.63 days), the time from bud initiation to bud color appearance (19.17 days), and the time until flower opening (67.71 days). Additionally, this treatment also yielded the highest number of buds per plant (3.25) and the largest flower diameter (14.53 cm).

**References**

1. Bahr LR, Compton ME. Competence for in vitro bulb regeneration among eight *Lilium* genotypes. HortScience. 2004;39(1):127-129.
2. Chaudhari UC, Shah NI, Makwana RJ, Maheriya PA, Rathva VD. Effect of different growing conditions with growing media on growth, flowering and yield of Asiatic *Lilium*. Int J Adv Biochem Res. 2024;8(8):764-770.

3. Dubey RK, Simrat-Singh, Kukal SS, Kalsi HS. Evaluation of different organic growing media for growth and flowering of *Petunia*. *Commun Soil Sci Plant Anal*. 2013;44:1777–1785.
4. Jimenez S, Plaza BM, Segura ML, Contreras JI, Lao TM. Peat substrate reuse in *Lilium* "Haveltia" crop. *Commun Soil Sci Plant Anal*. 2012;43:243-250.
5. Kumar R, Singh AK, Tomar KS, Gupta A. Effects of different media on growth and flowering traits of *Calendula officinalis* L. *Bangladesh J Bot*. 2022;51:417–422.
6. Noguera P, Abad M, Puchades R, Maquieria A, Noguera V. Influence of particle size on physical and chemical properties of coconut coir dust as container medium. *Commun Soil Sci Plant Anal*. 2003;34:593-605.
7. Oren-Shamir M, Gussakovsky E, Eugene E, Nissim-Levi A, Ratner K, Ovadia R, *et al*. Coloured shade nets can improve the yield and quality of green decorative branches of *Pittosporum variegatum*. *J Hortic Sci Biotechnol*. 2001;76(3):311-318.
8. Ribeiro HM, Romero AM, Pereira H, Borges P, Cabral F, Vaconcelos E. Evaluation of a compost obtained from forestry wastes and solid phase of pig slurry as a substrate for seedlings production. *Bioresour Technol*. 2007;98:3294-3297.
9. Rinaudo M. Chitin and chitosan: properties and applications. *Prog Polym Sci*. 2003;31(7):603–632.
10. Sultana N, Zakir HM, Parvin MA, *et al*. Physiological responses and nutritional qualities of tomato fruits to chitosan coating during postharvest storage. *Asian J Adv Agric Res*. 2019;10(2):1–11.
11. Tariq U, Rehman S, Aslam MK, Younis A, Yaseen M, Ahsan M. Agricultural and municipal waste as potting media components for the growth and flowering of *Dahlia hortensis* 'Figaro'. *Turk J Bot*. 2012;36:378-383.
12. Vendrame AW, Maguire I, Moore KK. Growth of selected breeding plants as affected by different compost percentages. *Proc Fla State Hortic Soc*. 2005;118:368–371.
13. Wilson SB, Stoffella PJ, Graetz DA. Development of compost-based media for containerized perennials. *Sci Hortic*. 2002;93:311-320.
14. Barreto MS, Jagtap KB. Assessment of substrates for economical production of *Gerbera* (*Gerbera jamesonii* Bolus ex. Hooker F.) flowers under protected cultivation. *J Ornamental Horticult*. 2006;9(2):136-138.
15. Barman D, Rajni K, Upadhyaya RC, Singh DK. Effect of horticultural practices for sustainable production of rose in partially modified greenhouse. *Indian J Hortic*. 2006;63(4):415-418.
16. Gupta YC, Le Quec D, Dhiman SR, Jain R. Standardization of growing media under protected environment for *Gerbera* in the mid-hills of Himachal Pradesh. *J Ornamental Horticult*. 2004;7(1):99-102.
17. Jawaharlal M, Joshua PJ, Arumugam J, Arumugam T, Subramanian S, Vijaykumar M. Standardization of growing media for *Anthurium* (*Anthurium andreaeanum*) cv. Temptation under shade net house. *South Indian Hortic*. 2001;46:323-325.
18. Swarup V. Delhi garden magazine. Flower show number. 1972;38-41.
19. Özgül Karagüzel Ü. Assessment of different growing media on cut flower performance of two gladiolus (*Gladiolus grandiflorus*) cultivars. *HortiScience*. 2023;40(2):36-42.
20. Lopez JA, González JE, Cos L, Fernández JA. Influence of different types of substratum on growth and flowering of *Gladiolus tristis* subsp. *concolor*. *Acta Hortic*. 2008;779:513-520.
21. Bhatia S, Gupta YC, Dhiman SR. Effect of growing media and fertilizers on growth and flowering of carnation under protected cultivation. *J Ornamental Horticult*. 2004;7(2):174-178.
22. Gupta YC, Le Quec D, Dhiman SR, Jain R. Standardization of growing media under protected environment for *Gerbera* in mid-hill of Himachal Pradesh. *J Ornamental Horticult*. 2004;7(1):99-102.
23. Barreto MS, Jagtap KB. Assessment of substrates for economical production of *Gerbera* (*Gerbera jamesonii* Bolus ex. Hooker F.) flowers under protected cultivation. *J Ornamental Horticult*. 2006;9(2):136-138.
24. Ysmeen S, Younis A, Rayit A, Raiz A. Effect of different substrate growth and flowering of carnation cv. Cauband Mixed. *Am-Eurasian J Agric Environ Sci*. 2012;12(2):249-258.
25. Salachna P, Zawadzińska A. Effect of chitosan on plant growth, flowering, and corm yield of potted freesia. *J Ecol Eng*. 2014;15(3):97-102.
26. Salachna P, Wilas J, Zawadzińska A. The effect of chitosan coating of bulbs on the growth and flowering of *Ornithogalum saundersiae*. *Acta Hortic*. 2015;1104:115-118.
27. Byczyńska A. Chitosan improves growth and bulb yield of pineapple lily (*Eucomis bicolor* Baker), an ornamental and medicinal plant. *World Sci News*. 2018;110:159-171.
28. El-Khateeb MA, Nasr AAM, Hassan NAA. Growth and quality improvement of *Dracaena surculosa*, Lindl by the foliar application of some bio-stimulants. *Int J Environ*. 2018;7(2):53-64.
29. Abd-El-Hady WMF. Response of tuberose (*Polianthes tuberosa* L.) plants to chitosan and seaweed foliar application. *Sci J Flowers Ornamental Plants*. 2020;7(2):153-161.
30. Panse VG, Sukhatme PV. *Statistical Methods for Agricultural Workers*. Indian Council of Agricultural Research, New Delhi; 1985.
31. Ramos-García SL, Roberson RW, Freitag M, Bartnicki-García S, Mouriño-Pérez RR. Cytoplasmic bulk flow propels nuclei in mature hyphae of *Neurospora crassa*. *Eukaryotic cell*. 2009 Dec;8(12):1880-1890.
32. Rady I, Bloch MB, Chamcheu RC, Banang Mbeumi S, Anwar MR, Mohamed H, *et al*. Anticancer properties of graviola (*Annona muricata*): A comprehensive mechanistic review. *Oxidative medicine and cellular longevity*. 2018;2018(1):1826170.