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## Studies on the impact of organic and inorganic stimulants on growth, flowering and corm production in gladiolus (*Gladiolus grandiflorus* L.)

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### Abstract

An experiment entitled “Studies on the impact of Organic and Inorganic Stimulants on growth, flowering and corm production in *Gladiolus grandiflorus* L.” was carried out at the Horticultural Research cum Instructional farm, College of Agriculture, Indira Gandhi Krishi Vishwavidyalaya, Raipur (C.G.) during the year 2023-24. The treatment contained GA<sub>3</sub> (100 and 200 ppm), Thiourea (2% and 3%), BA (50 and 100 ppm), KNO<sub>3</sub> (250 and 500 ppm), Cow urine (25 and 50%), Cow dung slurry (25 and 50%) and Water (Control). The experiment was performed in a Randomized Block Design with three replications. The application of GA<sub>3</sub> at 200 ppm proved to be the most effective in increasing days taken to sprouting (5.38 days), plant height (129.07 cm), number of days taken to spike initiation (59.00 days), spike length (76.13 cm), number of florets per spike (14.73), vase life of cut spike days (10.20), weight of corms per plant (63.30 g) and weight of cormels per plant (23.86 g). BA increased number of sprouts per corm (3.30), number of spike (3.30) and number of corms per plant (3.06). Whereas among the organic stimulants cow dung slurry at 50% performed best in improving spike length (72.80 cm).

**Keywords:** *Gladiolus*, GA<sub>3</sub>, BA, thiourea, KNO<sub>3</sub>, cow urine, cow dung slurry, randomized block design (RBD)

### Introduction

*Gladiolus* is a prominent cut flower in India and globally valued for its striking spikes adorned with elegant delicate florets that bloom sequentially, offering excellent keeping quality. These flowers ranging from white to deep crimson are primarily used for garden and interior decoration as well as in bouquet. Belonging to the Iridaceae family, *gladiolus* is native to Africa and Asia Minor and was introduced to India by the British in the 16<sup>th</sup> and 17<sup>th</sup> centuries. Initially believed to thrive only in hilly regions, advancements in agricultural practices have demonstrated that *gladiolus* can also flourish in various climates and soil types including the plains of Chhattisgarh. This has led to increased cultivation as a cash crop, particularly in districts like Raipur, Durg and Bilaspur. Currently, around 500 to 1,000 hectares are dedicated to *gladiolus* cultivation in the State.

Plant Growth Regulators (PGRs) are bioactive compounds that modulate specific growth processes in plants including root and shoot development, flowering time and overall yield and quality. Key PGRs like auxins, gibberellins, and cytokinins regulate cell division, elongation and flowering. In *gladiolus* cultivation, thiourea enhances growth, stress resilience, flowering and yield making it a valuable bioactive tool in ornamental horticulture. Additionally, Potassium Nitrate (KNO<sub>3</sub>) is essential for nutrient supply, flowering enhancement, root growth, stress tolerance and quality improvement. Organic inputs such as cow dung slurry and cow urine contribute to *gladiolus* cultivation by providing nutrients, improving soil health, boosting growth, increasing pest resistance, supporting sustainability, and enhancing flower quality and yield. Benzyl Aminopurine (BA) a growth regulator can enhance sprouting, boost the number of sprouts per plant and increase corm production.

### Materials and Methods

A field experiment was conducted at the Horticultural Research cum Instructional Farm, College of Agriculture, Indira Gandhi Krishi Vishwavidyalaya, Raipur (C.G.) during the

2023-24 season to assess the effects of GA<sub>3</sub>, thiourea, BA (benzyl adenine), KNO<sub>3</sub>, cow urine, and cow dung slurry on sprouting, flowering, and corm production. Medium-sized gladiolus corms (4-6 cm in diameter) were soaked in a 0.2% Carbendazim fungicide solution for 30 minutes, then dried in the shade for 3 to 4 hours. The corms were subsequently soaked for 24 hours in the various treatments and a control (water), then dried for another 24 hours before planting. They were planted randomly in plots spaced 30×20 cm apart, with irrigation applied the day before to maintain moisture. Planting done on November 2023, using a Randomized Block Design (RBD).

## Results and Discussion

### Vegetative parameters

#### Number of days taken to sprouting (days)

GA<sub>3</sub> significantly promoted the sprouting of corms. The shortest time to sprouting (5.38 days) was observed in Treatment T<sub>2</sub> - GA<sub>3</sub> at 200 ppm, which was statistically similar to Treatment T<sub>1</sub> - GA<sub>3</sub> at 100 ppm (5.68 days). Among the organic stimulants, T<sub>9</sub> (cow urine) required the least time to sprout, taking 7.60 days. Conversely, the longest duration to sprouting (15.73 days) was recorded in T<sub>0</sub> (Control). The observed decrease in number of days taken to sprouting from applying GA<sub>3</sub> may be attributed to the promoting effects of gibberellic acid on the dormancy of gladiolus corms, leading to enhanced cell division in the shoot tip and cell elongation. These findings align with earlier studies on the sprouting of gladiolus corms on cormels. Additional support for these results comes from Suresh *et al.* (2009) [41], Khan *et al.* (2013) [20], Amin *et al.* (2013) [2], and Neetu *et al.* (2013) [31]. Similar outcomes were also reported by Kumar and Singh (2005) [24] and Baskaran *et al.* (2014) [9] in gladiolus.

#### Number of sprouts per corm

The highest sprouts count per corm (3.30) was noted in Treatment T<sub>5</sub> (BA at 50 ppm), which was statistically similar to Treatment T<sub>6</sub> - BA at 100 ppm (3.20). Relatively, cow urine (T<sub>9</sub>) at 25% generated the maximum number of corms (1.33) among the organic stimulants. In comparison, Treatment T<sub>0</sub> (Control) had the minimum sprout count per corm (1.00). Axillary bud break and subsequent shoot growth have been reported to be stimulated by the external application of cytokinins to a number of plants. This may have been caused by BA breaking dormancy, which promoted sprouting. There are more shoots per corm as a result of BA's increase of cell expansion and shoot differentiation. This is in accordance with the results obtained by Abou-El-Ella (2007) [3], Anju Pal *et al.* (2003) [4] and Baskaran and Misra (2007) [7].

#### Plant Height (cm)

At 30 DAP, clearly showed that the maximum plant height (47.75 cm), was obtained in treatment T<sub>1</sub> (GA<sub>3</sub> at 100 ppm), which was also comparable to T<sub>2</sub> - GA<sub>3</sub> at 200 ppm (44.62 cm). Among the organic stimulants cow dung slurry at 25% (T<sub>11</sub>) obtained maximum height (43.80). However, minimum plant height (35.28 cm) was obtained in T<sub>0</sub> (Control).

At 60 DAP (88.34 cm) is obtained in treatment T<sub>2</sub> (GA<sub>3</sub> at 200 ppm) which was statistically similar to treatment T<sub>1</sub> - GA<sub>3</sub> at 100 ppm (87.79 cm). In case of organic stimulants cow dung slurry at 50% (T<sub>12</sub>) obtained maximum height

(84.46 cm). However, minimum plant height (69.47 cm) was obtained in treatment T<sub>0</sub> (Control).

At 90 DAP the maximum plant height (129.07 cm) was obtained in treatment T<sub>2</sub> (GA<sub>3</sub> at 200 ppm), which was statistically similar to treatment T<sub>1</sub> - GA<sub>3</sub> at 100 ppm (125.73 cm). In Case of organic stimulants cow dung slurry at 50% (T<sub>12</sub>) observed maximum height (123.47 cm). However, minimum plant height (103.8 cm) was obtained in T<sub>0</sub> (Control). GA<sub>3</sub> increases plant cell division and elongation, which results in an increase in the number of cells and lengthening of the cells, and both of which have an impact on plant growth. This is in accordance with the findings of Sharma *et al.* (2004) [39], Rana *et al.* (2005) [36], Bhalla and Kumar (2008) [10], Kumar *et al.* (2008) [23], Awasthi *et al.* (2012) [5], Chopde *et al.* (2012) [14], Dogra *et al.* (2012) [17] and Sudhakar and Kumar (2012) [40].

### Floral and corms Parameters

#### Number of days to spike initiation

The minimum number of days (59.00 days) to spike initiation was obtained in treatment T<sub>2</sub> (GA<sub>3</sub> at 200 ppm) which was statistically similar to treatment T<sub>1</sub> (GA<sub>3</sub> at 100 ppm) that is 59.13 days. Among the organic stimulants cow urine at 25% (T<sub>9</sub>) took minimum days (65.40) to spike initiation. However, Maximum number of days (73.06 days) to spike initiation was obtained in treatment T<sub>5</sub> (BA at 50 ppm). Early spike initiation (59 days) was recorded in GA<sub>3</sub> at 200 ppm, this may be the result of early corm emergence, early vegetative phase competition as seen by rapid cell division and elongation, and the fact that GA<sub>3</sub> is highly effective in shortening the plant juvenile phases (Wagh *et al.* 2012) [46].

#### Spike length (cm)

The maximum spike length (76.13 cm) was obtained with the treatment T<sub>2</sub> (GA<sub>3</sub> at 200 ppm) which was statistically similar to treatment T<sub>1</sub> - GA<sub>3</sub> at 100 ppm (73.26 cm). Among the organic stimulants cow dung slurry at 50% (T<sub>12</sub>) obtained maximum spike length which is (72.80 cm). Whereas, minimum spike length (63.00 cm) was obtained in treatment T<sub>0</sub> (Control). The increased spike length with the application of GA<sub>3</sub> might be due to rapid internodal elongation, rapid cell division and cell elongation in the intercalary meristem (Ashwini *et al.* 2019) [6].

#### Number of spike/plant

The maximum number of spike (3.30) was obtained with the treatment T<sub>5</sub> (BA at 50 ppm) which was statistically similar to treatment T<sub>6</sub> - BA at 100 ppm (3.13). As an organic stimulants cow urine at 25% and cow dung slurry at 50% both obtained maximum number of spike per plant (2.06). Whereas, minimum number of spike per plant (1.53) was obtained in treatment T<sub>0</sub> (Control). A possible explanation for an increase in spike yield is that cytokinin promotes lateral bud growth and cell division, both of which result in multiple shooting. This report is in conformity with Murti and Upreti (1995) [30] and Baskaran and Misra (2007) [7].

#### Number of florets/spike

The maximum number of florets per spike (14.73) was obtained with the treatment T<sub>2</sub> (GA<sub>3</sub> at 200 ppm) which was statistically similar to treatment T<sub>9</sub> - Cow urine at 25% (13.80). Among the organic stimulants cow urine at 25% (T<sub>9</sub>) obtained maximum number of florets per spike (13.80).

Whereas, minimum number of florets per spike (12.07) was obtained in treatment T<sub>0</sub> (Control). Gibberellic acid promotes the growth of auxiliary buds and their flowering. This might be resulted from the optimal amount of GA<sub>3</sub> being available, which enhanced the length of the rachis and

spikes, both of which are positively linked with the number of florets per spike. These observations and findings in the present investigation are in conformity with earlier reports by Sarkar *et al.* (2009) [38] in tuberose.

**Table 1:** Effect of Organic and inorganic stimulants on growth and flowering parameters

Treatments	Number of days taken to sprouting	Number of sprouts per corm	Plant height (cm)			Number of days to spike initiation	Spike length (cm)	Number of spike
Control (T <sub>0</sub> )	15.73 <sup>a</sup>	1.00 <sup>e</sup>	35.28 <sup>e</sup>	69.47 <sup>g</sup>	103.80 <sup>d</sup>	71.60 <sup>b</sup>	63.00 <sup>e</sup>	1.53 <sup>b</sup>
GA <sub>3</sub> 100 ppm (T <sub>1</sub> )	5.68 <sup>i</sup>	1.30 <sup>cd</sup>	47.75 <sup>a</sup>	87.79 <sup>a</sup>	125.73 <sup>ab</sup>	59.13 <sup>i</sup>	73.26 <sup>ab</sup>	1.66 <sup>b</sup>
GA <sub>3</sub> 200 ppm (T <sub>2</sub> )	5.38 <sup>i</sup>	1.60 <sup>b</sup>	44.62 <sup>b</sup>	88.34 <sup>a</sup>	129.07 <sup>a</sup>	59.00 <sup>j</sup>	76.13 <sup>a</sup>	2.00 <sup>b</sup>
Thiourea 2% (T <sub>3</sub> )	7.80 <sup>gh</sup>	1.30 <sup>cd</sup>	37.00 <sup>e</sup>	72.01 <sup>f</sup>	114.80 <sup>bcd</sup>	61.13 <sup>h</sup>	63.70 <sup>e</sup>	1.80 <sup>b</sup>
Thiourea 3% (T <sub>4</sub> )	9.00 <sup>e</sup>	1.50 <sup>bc</sup>	40.61 <sup>cd</sup>	71.99 <sup>f</sup>	113.60 <sup>cd</sup>	62.93 <sup>fg</sup>	63.20 <sup>e</sup>	2.13 <sup>b</sup>
BA 50 ppm (T <sub>5</sub> )	13.40 <sup>e</sup>	3.30 <sup>a</sup>	40.16 <sup>d</sup>	77.79 <sup>cd</sup>	118.13 <sup>abc</sup>	73.06 <sup>a</sup>	67.53 <sup>de</sup>	3.30 <sup>a</sup>
BA 100 ppm (T <sub>6</sub> )	14.13 <sup>b</sup>	3.20 <sup>a</sup>	36.64 <sup>e</sup>	78.86 <sup>c</sup>	119.80 <sup>abc</sup>	71.66 <sup>b</sup>	62.86 <sup>e</sup>	3.13 <sup>a</sup>
KNO <sub>3</sub> 250 ppm (T <sub>7</sub> )	11.86 <sup>d</sup>	1.30 <sup>cd</sup>	36.32 <sup>e</sup>	74.55 <sup>e</sup>	116.67 <sup>bc</sup>	63.26 <sup>f</sup>	66.60 <sup>de</sup>	1.60 <sup>b</sup>
KNO <sub>3</sub> 500 ppm (T <sub>8</sub> )	12.26 <sup>d</sup>	1.70 <sup>b</sup>	42.76 <sup>bcd</sup>	76.99 <sup>d</sup>	117.47 <sup>bc</sup>	62.46 <sup>g</sup>	66.267 <sup>de</sup>	1.73 <sup>b</sup>
Cowurine 25% (T <sub>9</sub> )	7.60 <sup>h</sup>	1.33 <sup>cd</sup>	43.08 <sup>bcd</sup>	83.51 <sup>b</sup>	120.47 <sup>abc</sup>	65.40 <sup>e</sup>	69.80 <sup>bcd</sup>	2.06 <sup>b</sup>
Cowurine 50% (T <sub>10</sub> )	7.80 <sup>gh</sup>	1.26 <sup>cde</sup>	41.98 <sup>bcd</sup>	84.35 <sup>b</sup>	121.93 <sup>abc</sup>	66.73 <sup>d</sup>	71.13 <sup>abcd</sup>	2.00 <sup>b</sup>
Cow Dung slurry 25% (T <sub>11</sub> )	8.06 <sup>g</sup>	1.06 <sup>de</sup>	43.80 <sup>bc</sup>	83.28 <sup>b</sup>	119.47 <sup>abc</sup>	66.80 <sup>d</sup>	67.83 <sup>cde</sup>	2.06 <sup>b</sup>
Cow dung slurry 50% (T <sub>12</sub> )	8.53 <sup>f</sup>	1.20 <sup>de</sup>	41.08 <sup>cd</sup>	84.46 <sup>b</sup>	123.47 <sup>abc</sup>	67.53 <sup>c</sup>	72.80 <sup>abc</sup>	1.80 <sup>b</sup>
S.Em±	0.14	0.084	1.01	0.44	3.32	0.214	1.612	0.216
CD at 5%	0.41	0.247	2.96	1.29	9.71	0.627	4.707	0.631

### Vase life of cut spike (days)

The maximum vase life of cut flower (10.2 days) was obtained with the treatment T<sub>2</sub> (GA<sub>3</sub> at 200 ppm) which was statistically similar to treatment T<sub>1</sub>- GA<sub>3</sub> at 100 ppm (8.80 days). Among the organic stimulants cow dung slurry at 25% (T<sub>11</sub>) obtained maximum vase life (7.86 days) of cut flower. Whereas, minimum vase life of cut flower (5.53 days) was obtained in treatment T<sub>0</sub> (Control). Maximum

vase life of cut spike was obtained maybe due to the superior action of auxin, which has been shown to delay senescence and promote metabolite translocation and may be helpful in extending the vase life of cut spike, may be the cause of the greater efficiency of the optimal dose of GA<sub>3</sub>. The findings are in agreement with Tawar *et al.* (2002) [43], Umrao *et al.* (2007) [45] and Chopde *et al.* (2013) [15] in gladiolus.

**Table 2:** Effect of Organic and inorganic stimulants on flowering and corm parameters

Treatment	Number of florets	Vase life of cut spike (days)	Number of corms per plant	Weight of corms per plant (g)	Weight of corms per plant (g)
Control (T <sub>0</sub> )	12.07 <sup>bc</sup>	5.33 <sup>g</sup>	1.86 <sup>c</sup>	32.70 <sup>f</sup>	11.93 <sup>g</sup>
GA <sub>3</sub> 100 ppm (T <sub>1</sub> )	13.27 <sup>abc</sup>	8.80 <sup>b</sup>	2.6 <sup>ab</sup>	61.90 <sup>a</sup>	20.40 <sup>b</sup>
GA <sub>3</sub> 200 ppm (T <sub>2</sub> )	14.73 <sup>a</sup>	10.20 <sup>a</sup>	2 <sup>c</sup>	63.30 <sup>a</sup>	23.86 <sup>a</sup>
Thiourea 2% (T <sub>3</sub> )	12.73 <sup>bc</sup>	8.20 <sup>c</sup>	2.26 <sup>bc</sup>	37.80 <sup>e</sup>	15.26 <sup>f</sup>
Thiourea 3% (T <sub>4</sub> )	12.73 <sup>bc</sup>	8.66 <sup>b</sup>	2.13 <sup>bc</sup>	39.00 <sup>e</sup>	16.06 <sup>def</sup>
BA 50 ppm (T <sub>5</sub> )	12.53 <sup>bc</sup>	7.40 <sup>de</sup>	3.06 <sup>a</sup>	52.60 <sup>cd</sup>	15.60 <sup>ef</sup>
BA 100 ppm (T <sub>6</sub> )	11.87 <sup>c</sup>	7.53 <sup>de</sup>	2.8 <sup>a</sup>	51.10 <sup>d</sup>	17.93 <sup>cd</sup>
KNO <sub>3</sub> 250 ppm (T <sub>7</sub> )	12.20 <sup>bc</sup>	7.20 <sup>e</sup>	2.2 <sup>bc</sup>	39.20 <sup>e</sup>	12.93 <sup>g</sup>
KNO <sub>3</sub> 500 ppm (T <sub>8</sub> )	13.20 <sup>abc</sup>	6.20 <sup>f</sup>	2.06 <sup>bc</sup>	50.00 <sup>d</sup>	15.53 <sup>f</sup>
Cowurine 25% (T <sub>9</sub> )	13.80 <sup>ab</sup>	7.60 <sup>de</sup>	2.13 <sup>bc</sup>	50.26 <sup>d</sup>	17.80 <sup>cde</sup>
Cowurine 50% (T <sub>10</sub> )	13.60 <sup>abc</sup>	7.73 <sup>d</sup>	1.90 <sup>c</sup>	57.10 <sup>b</sup>	17.20 <sup>cdef</sup>
Cow Dung slurry 25% (T <sub>11</sub> )	12.67 <sup>bc</sup>	7.86 <sup>cd</sup>	2.06 <sup>bc</sup>	54.70 <sup>bc</sup>	18.66 <sup>bc</sup>
Cow dung slurry 50% (T <sub>12</sub> )	13.40 <sup>abc</sup>	7.73 <sup>d</sup>	1.89 <sup>c</sup>	51.60 <sup>d</sup>	16.06 <sup>def</sup>
S.Em±	0.51	0.19	0.17	0.889	0.68
CD at 5%	1.50	0.14	0.51	2.594	1.99

### Number of corms/plant

The maximum number of corms/plants (3.06) was obtained with that of treatment T<sub>5</sub> (BA at 50 ppm) which was statistically similar to treatment T<sub>6</sub> - BA at 100 ppm (2.80). Among the organic stimulants cow urine at 25% (T<sub>9</sub>) obtained maximum number of corms per plant (2.13). Whereas, minimum number of corms per plant (1.86) was obtained in treatment T<sub>0</sub> (Control). Maximum number of corms per plant was obtained in BA at 50 ppm which might be due to, BA typically results in greater cell division and splitting than in gladiolus corm size growth. Khan *et al.* (2013) [20] found that the concentration of BA enhanced

multiple shooting and accelerated corm production in gladiolus. The result is in conformity with Raju (2000) [34] in lillies and Rajaram *et al.* (2002) [35].

### Weight of corms per plant (g)

The maximum weight of corms per plant (63.30 g) was obtained with the treatment T<sub>2</sub> (GA<sub>3</sub> at 200 ppm), which was statistically similar to treatment T<sub>1</sub> - GA<sub>3</sub> at 100 ppm (61.90). Among the organic stimulants treatment T<sub>10</sub> (Cow urine at 50%) obtained maximum weight of corms per plant (57.10 g). Whereas, minimum weight of corms per plant (32.7 g) was obtained in treatment T<sub>0</sub> (Control). GA<sub>3</sub>



obtained maximum weight of corms per plant maybe due to its ability to increase the number of leaves which in turn increased the photosynthesis and photosynthetic assimilates. These assimilates were transported to the daughter corms, thereby increasing size and weight. Supportively, lowest values for corm weight were recorded in control (Reddy *et al.*, 2013) [37].

#### Weight of cormels/Plant (g)

The maximum weight of cormels per plant (23.86 g) was obtained with the treatment T<sub>2</sub> (GA<sub>3</sub> 200 ppm) which was statistically similar to treatment T<sub>1</sub> GA<sub>3</sub> 100 ppm (20.40 g). As an organic stimulants T<sub>11</sub> (cow dung slurry at 25%) obtained maximum weight of cormels (18.66 g). Whereas, minimum weight of cormels per plant was obtained with treatment T<sub>0</sub> (Control). Gladiolus plants treated with GA<sub>3</sub> have much higher cormel weights per plant. This may be because the GA<sub>3</sub> treated plants stay physiologically active longer to accumulate enough food, which leads to improved plant growth and development and ultimately higher cormel weights per plant. Similar findings are observed by Baskaran *et al.* (2009) [8], Gaur *et al.* (2003) [19] and Padmalatha *et al.* (2012) [32].

#### Conclusion

It can be inferred from the results of the present investigation, that the treatment with GA<sub>3</sub> at 200 ppm proved to be the best in improving vegetative and flowering characteristics, including days taken for sprouting, plant height, number of leaves per plant, number of days taken to spike initiation, spike length, number of florets, vase life, weight of corms and cormels per plant. The maximum number of sprouts per corm, number of spike and number of corms per plant was obtained in BA at 50 ppm. Whereas in case of organic stimulants cow dung slurry at 50% performed best in improving spike length.

In terms of commercial potency, for quality flower production the application of GA<sub>3</sub> at 200 ppm maybe recommended to get good quality spike with longer vase life. On the other hand, from economic point of view, the application of BA at 50 ppm can be considered best for getting more number of spikes and corms.

#### References

1. Arora JS, Kushal S, Grewal NS, Singh K. Effect of GA<sub>3</sub> on corm and growth in gladiolus. *Indian Journal of Plant Physiology*. 1992;35:202-206.
2. Amin NU, Khattak AM, Ahmad I, Ara N, Alam A, Ali A, Ali I. Corm and cormel size of gladiolus greatly influenced growth and development of subsequent corm production. *Pakistan Journal of Botany*. 2013;45(4):1407-1409.
3. Abou-El-Ella EM. Physiological studies on *Acanthus mollis* plants. M.Sc. Thesis, Horticulture Department, Faculty of Agriculture, Benha University, Egypt. 2007;22:571-582.
4. Anju P, Santhosh K, Srivastava R. Effect of floral preservatives on postharvest management in gladiolus spikes. *Journal of Ornamental Horticulture*. 2003;6(4):367-371.
5. Awasthi AK, Karhana PK, Pandey KK. The application of preservatives on quality of gladiolus (*Gladiolus grandiflorus* L.) cut spike. *International Journal of Plant, Animal and Environmental Sciences*. 2012;3(3):1-4.
6. Ashwini A, Munikrishnappa PM, Kulkarni BS, Kumar R, Amreen T, MK S. Effect of plant growth regulators on vegetative and flowering parameters of gladiolus (*Gladiolus hybridus* L.) cv. Adigo yellow. *International Journal of Chemical Studies*. 2019;7(2):1553-1556.
7. Baskaran V, Misra RL. Effect of plant growth regulators on growth and flowering of gladiolus. *Indian Journal of Horticulture*. 2007;64:479-482.
8. Baskaran V, Misra RL, Abirami K. Effect of plant growth regulators on corm production in gladiolus. *Journal of Horticultural Sciences*. 2009;4(1):78-80.
9. Baskaran V, Abirami K, Roy SR. Effect of plant growth regulators on yield and quality in gladiolus under Bay Island conditions. *Journal of Horticultural Sciences*. 2014;9(2):213-216.
10. Bhalla R, Kumar A. Response of plant bio-regulators on dormancy breaking in gladiolus. *Journal of Ornamental Horticulture*. 2008;11(1):18.
11. Bhattacharjee SK. Response of *Lilium tigrinum* Ker-Gawl (tiger lily) to soil drench application of growth regulating chemicals. *Progressive Horticulture*. 1983;15:204-209.
12. Bhattacharjee SK. The effects of growth regulating chemicals on gladiolus. *Gartenbauwissenschaft*. 1984;49:103-106.
13. Chopde N, Gonge VS, Nagre PK. Effect of growth regulators on growth and flowering of gladiolus. *Asian Journal of Horticulture*. 2011;6(2):398-401.
14. Chopde N, Gonge VS, Dalal SR. Growth flowering and corm production of gladiolus as influenced by foliar application of growth regulators. *Plant Archives*. 2012;12(1):41-46.
15. Chopde N, Gonge VS, Warade AD. Influence of growth regulators on gladiolus varieties. *Journal of Agricultural Research and Technology*. 2013;38(3):369-374.
16. Dua IS, Sehgal OP, Chark KS. Gibberellic acid induced earliness and increased production in gladiolus. *Gartenbauwissenschaft*. 1984;49:91-94.
17. Dogra S, Pandey RK, Bhat D. Influence of Gibberellic acid and plant geometry on growth, flowering and corm production in gladiolus under Jammu agroclimate. *International Journal of Pharmacy and Biological Sciences*. 2012;3(4):1083-1090.
18. El-Meligy MM. Effect of gibberellin and radiation on corm formation and anthocyanin content in gladiolus. *Agricultural Research Review*. 1982;60:265-280.
19. Gaur GS, Chaudhary TC, Trivedi JD. Effect of GA<sub>3</sub> and IAA on growth, flowering and corm production in gladiolus cv. Eurovision. *Journal of Farm Sciences*. 2003;12(1):1-3.
20. Khan FN, Rahman MM, Hossain MM. Effect of benzyladenine and gibberellic acid on dormancy breaking, growth and yield of gladiolus corms over different storage periods. *Journal of Ornamental and Horticultural Plants*. 2013;3(1):59-71.
21. Khan FN, Rahman MM, Hossain MM, Hossain T. Effect of benzyladenine and gibberellic acid on dormancy breaking and growth of gladiolus cormels. *Thai Journal of Agricultural Science*. 2011;44:165-174.
22. Kumar R, Dubey RK, Misra RL. Effect of GA<sub>3</sub> on growth, flowering and corm production of gladiolus. In: Misra RL, Misra S, editors. *Floriculture Research Trend*

- in India. New Delhi: Indian Society of Ornamental Horticulture; c2002. p. 110-113.
23. Kumar S, Bhagawat PR, Kumar R, Ronya T. Effect of plant growth regulators on vegetative growth, flowering and corm production of gladiolus in Arunachal Pradesh. *Journal of Ornamental Horticulture*. 2008;11:265-270.
  24. Kumar V, Singh RP. Effect of soaking of mother corms with plant growth regulators on vegetative growth, flowering and corm production in gladiolus. *Journal of Ornamental Horticulture*. 2005;8(4):306-308.
  25. Kumar PN, Reddy YN, Chandrashekhar R. Effect of growth regulators on flowering and corm production in Gladiolus. *Indian Journal of Horticulture*. 2008;65(1):73-78.
  26. Khan A, Bahadur V. Effect of plant growth regulators on growth and spike yield of gladiolus cultivars. *HortFlora Research Spectrum*. 2013;2(4):341-345.
  27. Mahesh KS. Effect of growth regulators on gladiolus var. Snow Princess [M.Sc. Thesis]. New Delhi: Indian Agricultural Research Institute (IARI); c1992.
  28. Maurya RP, Nagda CL. Effect of growth substances on growth and flowering of gladiolus (*Gladiolus grandiflorus* L.) cv. Friendship. *Haryana Journal of Horticultural Sciences*. 2002;31:203-204.
  29. Mukhopadhyay A, Bankar GJ. Pre-planting soaking of corm with gibberellic acid modified growth and flowering of gladiolus cultivar 'Friendship'. *Indian Agriculture*. 1986;30:317-319.
  30. Murti GSR, Upreti KK. Use of growth regulators in ornamental plants. In: Chadda KL, Bhattacharjee SK, editors. *Advances in Horticulture*, Vol. 12: Ornamental Plants. New Delhi: Malhotra Publishing House; c1995. p. 863-883.
  31. Neetu, Singh AK, Sisodiya A, Kumar R. Effect of GA<sub>3</sub> on growth and flowering attributes of gladiolus cultivars. *Annals of Agricultural Research*. 2013;34(4):315-319.
  32. Padmalatha T, Reddy GS, Chandrasekhar R, Shankar AS, Chaturvedi A. Effect of foliar sprays of chemicals and plant growth regulators on corm production and vase life in gladiolus plants raised from cormels. *Plant Archives*. 2012;12(2):685-690.
  33. Roychoudhuri N, Biswas J, Dhua RS, Mitra SK. Effect of chemicals on germination, growth, flowering and corm yield of gladiolus. *Indian Agriculture*. 1985;29(3):215-217.
  34. Raju DVS. Effect of plant growth regulators and disbudding on growth and development of Asiatic hybrid lily cv. Corrida [M.Sc. Thesis]. New Delhi: Indian Agricultural Research Institute (IARI); c2000.
  35. Rajaram R, Mukherjee S, Manuja S. Plant growth regulators affect the development of both corms and cormels in Gladiolus. *HortScience*. 2002;37(2):343.
  36. Rana P, Kumar J, Kumar M. Response of GA<sub>3</sub>, plant spacing and planting depth on growth, flowering and corm production in gladiolus. *Journal of Ornamental Horticulture*. 2005;8(1):41-44.
  37. Reddy GVS, Nageswararao MB, Umajyothi K, Sasikala K. Studies on the effect of plant growth regulators on flowering, corm and cormel production in gladiolus (*Gladiolus grandiflorus* L.) cv. White Prosperity. *Journal of Research ANGRAU*. 2013;41(2):11-15.
  38. Sarkar J, Misra RL, Singh SK, Prasad KV, Arora A. Effect of growth regulators on growth and flowering in tuberose under North India conditions. *Indian Journal of Horticulture*. 2009;66(4):502-507.
  39. Sharma JR, Gupta RB, Panwar RD. Growth, flowering and corm production of gladiolus cv. Friendship as influenced by foliar application of nutrients and growth regulators. *Journal of Ornamental Horticulture*. 2004;7:154-158.
  40. Sudhakar M, Rameshkumar S. Effect of growth regulators on growth, flowering and corm production of gladiolus (*Gladiolus grandiflorus* L.) cv. White Friendship. *Indian Journal of Plant Sciences*. 2012;1(2/3):133-136.
  41. Suresh KK, Chandra SR, Siva SA. Effect of plant growth regulators on dormancy, corm and cormel production in gladiolus (*Gladiolus × grandiflorus* L.). *Journal of Ornamental Horticulture*. 2009;12(3):182-187.
  42. Taiz L, Zeiger E. *Plant Physiology*. 2nd ed. Sunderland (MA): Sinauer Associates, Inc.; c1998. p. 792.
  43. Tawar RV, Sable AS, Giri MD. Effect of growth regulators on growth and flowering of gladiolus cv. Jester. *Annals of Plant Physiology*. 2002;16(2):109-111.
  44. Tawar RV, Sable AS, Kakad GJ, Hage ND, Ingle MB. Effect of growth regulators on corm and cormel production of gladiolus (cv. Jester). *Annals of Plant Physiology*. 2007;21:257-258.
  45. Umrao VK, Singh RP, Singh AR. Effect of gibberellic acid and growing media on vegetative and floral attributes of gladiolus. *Indian Journal of Horticulture*. 2007;64(1):73-76.
  46. Wagh VK, Chawla SL, Gaikwad AR, Parolekar SS. Effect of bulb size and GA<sub>3</sub> on vegetative and floral characters of tuberose (*Polyanthes tuberosa* L.) cvs. Prajwal and Calcutta Single. *Progressive Horticulture*. 2012;44(1):27-31.